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Ameliorating Effects of *Croton Bonplandianus* Leaves against Gentamicin Induced Acute Liver and Kidney Injury: A Biochemical and Histopathological Investigation

Hina Yasin^{1,2*}, Shaukat Mahmud², Raheela Bano³, Hina Abrar⁴, Kaneez Fatima⁵, Rabia Bushra³

¹ Department of Pharmacognosy, Faculty of Pharmacy, Dow University of Health Sciences, Ojha Campus, Gulzar E Hijri, Scheme-33, Karachi, Pakistan

² Department of Pharmacognosy, Baqai Institute of Pharmaceutical Sciences, Baqai Medical University, Super Highway, Gadap Road, Karachi, Pakistan

³ Department of Pharmaceutics, Faculty of Pharmacy, Dow University of Health Sciences, Ojha Campus, Gulzar E Hijri, Scheme-33, Karachi, Pakistan

⁴ Department of Pharmacology, Faculty of Pharmacy, Dow University of Health Sciences, Ojha Campus, Gulzar E Hijri, Scheme-33, Karachi, Pakistan

⁵ Department of Pharmacognosy, Institute of Pharmaceutical Sciences, Jinnah Sindh Medical University, Rafiqui H.J Shaheed Road, Karachi, Pakistan

* Corresponding author: hina.yaseen@duhs.edu.pk

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Abstract: The utilization of natural constituents for the treatment of various medical complaints is well established. The members of the Euphorbiaceae family, especially the *Croton* genus, are reported to be rich in diterpenoids and showing various biological activities. Many activities of *Croton bonplandianus* are still unfolded therefore, current study is designed to evaluate the ameliorative effects of aqueous extract of *C. bonplandianus* against gentamicin-induced liver and kidney injuries in rats. The aqueous extract of leaves (27-81 mg/kg) was administered to albino rats Wistar strain for eight days with concurrent administration of gentamicin (100 mg/kg) daily. The protective effect of the aqueous extracts on the liver was determined by liver function tests (SGPT, ALT, GGT, Bilirubin) and kidney functions were evaluated by assessment of plasma biomarkers including urea, creatinine, uric acid and electrolytes (sodium, potassium, chloride and bicarbonate). Histopathological studies of liver and kidney tissue were carried out by Hematoxylin and Eosin (H& E) staining confirmation of the consequences. Significant alterations in liver and kidney tissues treated groups with aqueous extracts of leaves were obtained in lower doses (27-54 mg/kg). Similarly, serum levels of SGPT, ALT, GGT, bilirubin, creatinine, and urea in the treated group were significantly reduced ($P < 0.01$) at similar doses, whereas no significant changes were founded in levels of ions and uric acids. The histological examination of the liver and kidney confirmed the biochemical changes in treated rats. The underlying results demonstrate the protective effect of aqueous leave extracts *Croton bonplandianus* on an acute liver and kidney injury rat model. The hepato and renal protective effects of *C. bonplandianus* are certainly due to the presence of diterpenoids flavonoids and polyphenols exhibiting free radical or oxygen scavenging activities.

Keywords: *Croton bonplandianus*, gentamicin, histopathological studies, nephroprotective effect.

巴豆邦普兰迪亚努斯叶对庆大霉素诱导的急性肝肾损伤的改善作用：生化和组织病理学研究

摘要：利用天然成分治疗各种医疗问题已经很成熟。据报道，大戟科植物，尤其是巴豆属植物，富含二类化合物，具有多种生物活性。巴豆的许多活性仍未展开，因此，当前的研究旨在评估巴豆水提取物对庆大霉素导的大鼠肝肾损伤的改善作用。将叶子的水提取物(27-81毫克/公斤)给予白化病大鼠威斯达品系8天，同时每天给予庆大霉素(100毫克/公斤)。水提取物通过对肝脏的保护作用通过肝功能测试(血清谷氨酸-丙酮酸转氨、丙氨酸氨基转移酶、 γ -谷氨转移、胆红素)确定,肾功能通过评估血浆生物标志物(包括尿素、肌酸、尿酸和电解质钠、氯、盐和盐素)。对肝脏和肾脏组织进行病理学研究，通过苏木精和伊红染色确认结果。在较低剂量(27-54毫克/公斤)的叶水提取物治疗组中，肝脏和肾脏组织发生了显著变化。同样，治疗组血清谷丙转功能、丙氨酸氨基转移、维生素、维生素、含微毒素素素素素和血清素含量在相似剂量下显著降低($P < 0.01$)，而离子水平没有显著变化和尿酸。肝脏和肾脏的组织学检查证实了治疗大鼠的生化变化。基本结果证明了巴豆叶水提取物对急性肝肾损伤大鼠模型的保护作用。巴豆的肝和肾保护作用肯定是由于二种黄酮和多甾的存在表现出自由基或氧清除活性。

关键词：巴豆，庆大霉素，组织病理学研究，肾保护作用。

Introduction

Pakistan showed rich and diverse plants, from which substances with distinct pharmacological activities have been identified. Moreover, the use of medicinal flora native to Pakistan is the growing area of research for scientists [1]. Wide varieties of medicinal plants have been tested and showed therapeutic potential against drug-induced liver and kidney disorders [2]. Several compounds in the plants such as flavonoids, alkaloids, terpenes, and phenolic compounds possess hepatoprotective and nephroprotective activities [3]. Drug-induced liver and kidney injuries are the main causes of drug discontinuation despite the ultimate need for therapy. *Croton bonplandianus* is a perennial herb indigenous to south Bolivia and founded in Asia. Different species of *Croton* (Euphorbiaceae) showed antiviral, anti-bacterial, anti-fungal, anti-inflammatory, anti-plasmodic, anti-proliferative, and antiseptic activities in the complementary traditional medicine [4]. Several compounds are identified in various croton species such as alkaloids, terpenoids, and saponins that have marked therapeutic activity [5].

Literature revealed that the methanolic and aqueous extracts of *C. bonplandianus* contained rutin, crotosparine and its methylated derivative phorbol, flavonoids, alkaloids and phenolic content, saponin and tannins [6, 7]. Plant material showed analgesic, anti-oxidant, anti-bacterial, anti-fungal, hepatoprotective, anti-coronary, insect repellent, and wound healing effects [8].

Liver and kidney damage are mainly caused by the

used of various chemical agents, when taken in high dose or on prolong used [9]. Kidney damage is usually characterized with increased exposure to medication [10]. Non-selective NSAIDs (non-steroidal anti-inflammatory drugs), anti-cancer drugs, antibiotics, and amino glycosides are well-reported causes of renal damage [11, 12]. Gentamicin is an aminoglycoside antimicrobial used to treat several grams-negative infections but it showed severe adverse drugs such as nephrotoxicity, hepatotoxicity and ototoxicity.

The possible mechanism of hepatic injury of gentamicin is the induction of oxidative stress and inflammation that leads to the mitochondrial dysfunctioning and ultimately leads to elevation in hepatic biomarkers such as ALT, ALP, total bilirubin, and gamma GT [13, 14]. Drug-induced acute liver damage is considered as the membrane deterioration, infiltration of neutrophils, hepatocyte necrosis, release of cytokines, and production of reactive oxygen species (ROS) [9]. Similarly, gentamicin also induces kidney damage by producing reactive oxidation species and apoptosis in the renal proximal tubule [15]. Gentamicin deposits in the renal tubule, where it induces membrane damage, necrosis, inflammation, apoptosis, and release of vasoconstrictor hormone. These events affect the membrane permeability and ultimately lead to the reduction in the glomerular filtration rate (GFR) [3]. These effects make it a suitable agent for the induction of hepatic and nephrotoxicity [16].

On the basis of former research, the aqueous extracts of leaves of the *Croton bonplandianus* were selected for hepatoprotective and nephroprotective

effects against gentamicin-induced hepatic and renal injuries.

The aqueous fraction is considered safe while administered to animals for liver protective and renal protective effects comparative to methanolic extract. The detailed microscopic examination of liver and kidney tissues was conducted to ascertain the preventative effect of plant extracts on tissues of vital organs. Beside the histopathological studies biomarker related to hepatic and renal injuries were also determined to confirm the findings of the study.

1. Materials and Method

Aqueous extracts of leaves of *Croton bonplandianus* (CB) (27-81 mg/kg), paracalcitrol (PC) (100 mg/kg), gentamicin (GM) (100 mg/kg), distilled water, and feeding tubes.

1.1. Solvents

Hexane (Sigma-Aldrich), chloroform (Sigma-Aldrich), methanol (Sigma-Aldrich), ethyl acetate (Sigma-Aldrich), and distilled water.

1.2. Collection and Identification of Plant Material

Leaves of the *Croton bonplandianus* (Euphorbiaceae) were collected from the botanical garden of the University of Karachi. Leaves were authenticated by the taxonomist from the Department of Botany, University of Karachi. A voucher specimen (no. 03) was preserved and deposited in the Department of Pharmacognosy, Baqai Institute of Pharmaceutical Sciences, Baqai Medical University, Karachi.

1.3. Extraction and Fractionation of Leaves

Dried leaves (1 kg) were extracted in Methanol (7L) for 15 days at room temperature followed by filtration with Whatman filter paper (45 micron). This procedure was repeated with residual material twice and combined all filtrates. The methanolic extract was concentrated using a rotary evaporator under reduced pressure and controlled temperature (40°C). Then, the extract was suspended in H₂O sequentially partitioned with n-hexane, chloroform and EtOAc. All the fractions were lyophilized to obtain dried extracts and were stored at 4°C up to be used for pharmacological work [17, 18].

1.4. Animal Model

Adult male albino rats of wistar strain weighing 200 ± 10 g were selected for nephroprotective and hepatoprotective activity provided by Baqai Institute of Pharmaceutical Sciences, Baqai Medical University Karachi followed using Winter method [19].

Animals were kept as per the standard rule given by animal houses with maintained temperature 25 ± 2 °C

for 12-hour light/dark cycle. Rats fed with a laboratory standard balance diet and water *ad libitum* [20].

1.4.1. Ethical Approval

The study was approved by the Ethical Committee of Baqai Medical University, Karachi, Pakistan Ref. # BUU-EC/2018/01.

1.5. Experimental Design

Animals were weighed and randomly divided in six groups, each of six animals and treated in the following manner:

The control group daily received only 10 ml/kg distilled water for 8 days orally.

GM group: treated with 100 mg/Kg of gentamicin intra-peritoneally daily.

GM-PC group: treated with 100 mg/Kg of gentamicin and 100 mg/Kg paracalcitriole daily.

GM-CB group rats were given three different oral doses (81, 54 and 27 mg/Kg) of aqueous extract of CB concurrent with GM at 100mg/kg daily for 8 days [4, 21].

1.6. Blood Collection and Tissue Preparation

After 24 hours after the last dose, all the animals were anesthetized. The thoracic cage was exposed and approximately 5ml of blood was collected from each rat by the cardiac puncture technique. Blood samples were then transferred to the gel tube for biochemical estimation [22]. The blood was centrifuged for 8-10 minutes at 3500 RPM. The serum was stored after separation into eppendorf tubes at -20 °C for estimation of renal and hepatic biomarkers. After that, the rats were sacrificed by cervical dislocation and their livers and both kidneys were removed immediately and kept in 10% formaldehyde for histopathological processes [23-25].

1.7. Measurement of Serum Biochemical Parameters

The hepatic biomarkers including AST, ALT, GGT and serum total, direct and indirect bilirubin were assayed using commercial reagent kits from Merck (Germany). Renal biochemical markers (creatinine, urea and uric acid) were estimated using commercial reagent kits (Sigma-Aldrich) as per the manufacturer protocol. The serum electrolyte levels were also determined by adopting standard methods.

1.8. Histopathological Examination

The isolated kidneys and liver from all groups were preserved in formalin 10% and embedded in paraffin wax as a standard protocol. Afterward 5-micron thick tissue sections were sliced, mounted on a poly-L-lysine coated glass slide, and stained with haematoxylin and eosin (H & E) for histopathological examination. Stepwise methodology are represented in fig. 1 [26].

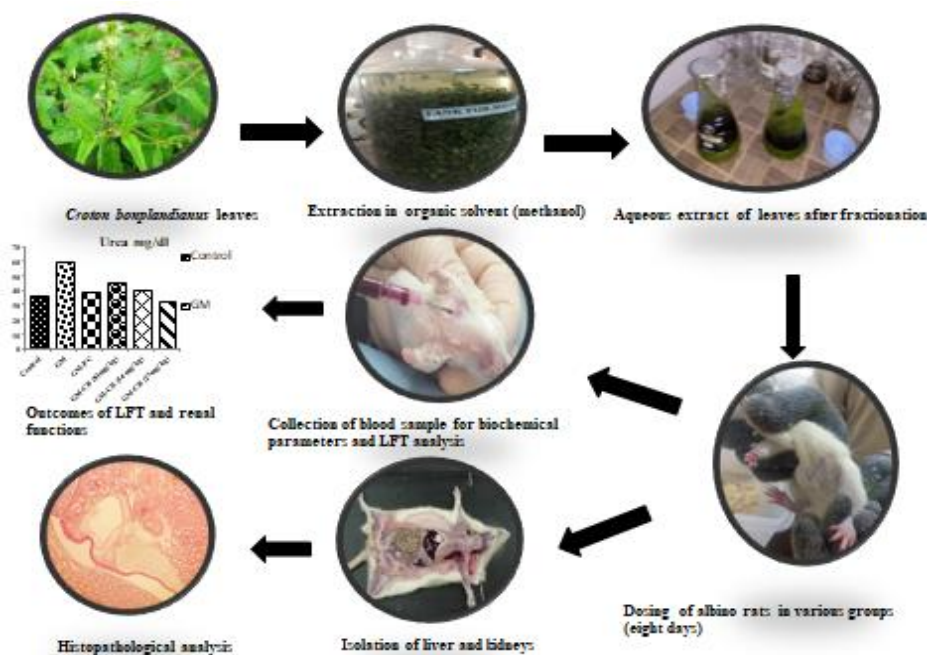


Fig. 1 Leading steps of the research methodology

2. Result

2.1. Biochemical Analysis

2.1.1. Renal Functional Biomarkers

Gentamicin (GM) induced nephrotoxic mode was used to evaluate the nephroprotective effects of plant extracts. The serum electrolyte levels, including Na^{++} , K^{+} , Cl^{-} and HCO_3^{-} were not changed significantly. The lower dose of CB showed a greater reduction in serum

uric acid levels compared to high dose. The GM-treated group showed elevated levels of urea and creatinine, while these parameters were markedly reduced in GM-PC group (standard). Concurrent administration of plant extracts in different doses with gentamicin displayed a protective role and prevented drug-induced nephropathies. The lower dose of CB showed a group maximum protective effect compared to the higher dose indicated by the marked reduction in creatinine level (Table 1, Fig. 2 and 3).

Table 1 Effect of aqueous extract of leaves and bark of *Croton bonplandianus* on different kidney functions

Samples	Uric acid mg/dl	Sodium mEq/l	Potassium mEq/l	Chloride mEq/l	Bicarbonate mEq/l
Control	1.6 ± 0.02	143 ± 1.27	4.2 ± 0.15	97 ± 0.24	28 ± 0.18
GM	1 ± 0.0	141 ± 0.16	4.6 ± 0.32	104 ± 0.45	20 ± 0.03
GM-PC	1 ± 0.0	143 ± 0.42	3.9 ± 0.47	103 ± 0.17	22 ± 0.4
GM (81 mg/kg)	1.41 ± 0.36	143 ± 0.77	4.85 ± 0.01	101 ± 0.24	27 ± 0.15
GM (54mg/kg)	1.84 ± 0.27	141 ± 1.58	6 ± 0.23	101.5 ± 0.98	23.5 ± 1.26
GM (27 mg/kg)	1.12 ± 0.06	142 ± 0.10	4.4 ± 0.17	104.5 ± 0.35	25 ± 0.77

Notes: Control - control group treated with distilled water (10 ml/Kg), GM - group treated with gentamicin, GM-PC - group treated with paracalcitriol (100 mg/kg) with gentamicin, GM-CB - groups treated with Aqueous extract of leaves of *C. bonplandianus* (81, 54 and 27 mg/kg). Not significant result obtained compared with control $P > 0.05$. Data expressed as mean ± SEM (n = 6) (ANOVA). P values < 0.05 were considered significant.

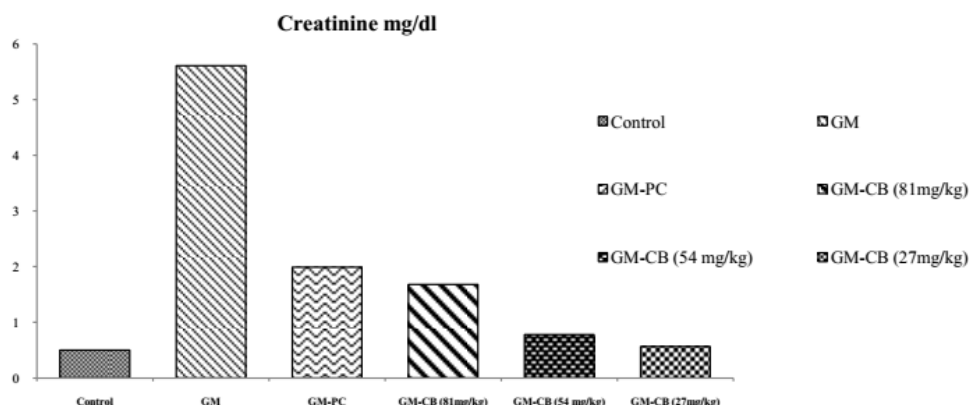


Fig. 2 Effects of different groups on creatinine levels for kidney function

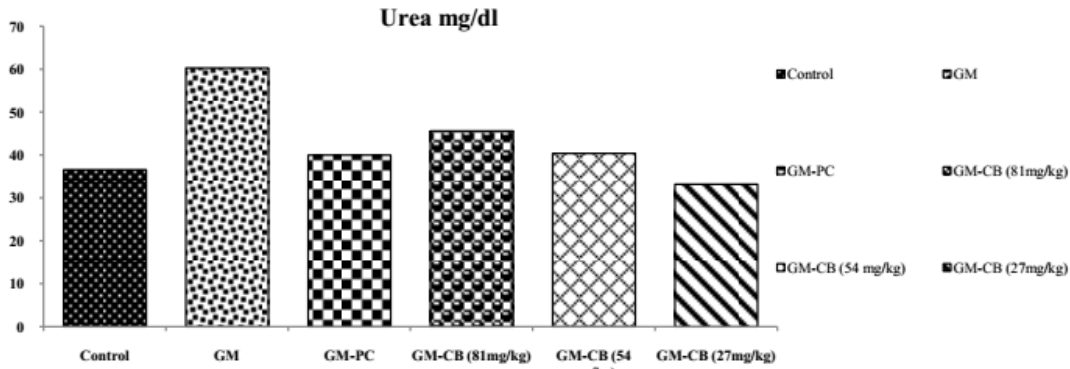


Fig. 3 Effects of different groups on urea levels for kidney function

2.1.2. Hepatic Functional Biomarkers

In the present studies, gentamicin induced liver damage indicated by elevated levels of transaminases (ALT, AST) in serum and minute increase in gamma GT (Fig. 4). The group treated with GM also showed an increased in plasma bilirubin concentration, which is characteristic marker that indicates liver injury and inhibition of detoxifying functions of the liver (fig. 5). Concomitant administration of CB extracts in different Doses along with GM showed a protective effect on the

liver as all hepatic biomarkers were significantly reduced. All the doses of CB produce almost similar protective effects on GM-induced hepatotoxicity. These hepatic damage markers were also recently reported to be decreased by another species of croton called *Croton hypoleucus* [27]. In another investigation, the presence of α -amyrin in the leaf extract of *C. bonplandianus* has been documented against hepatocellular damage [28].

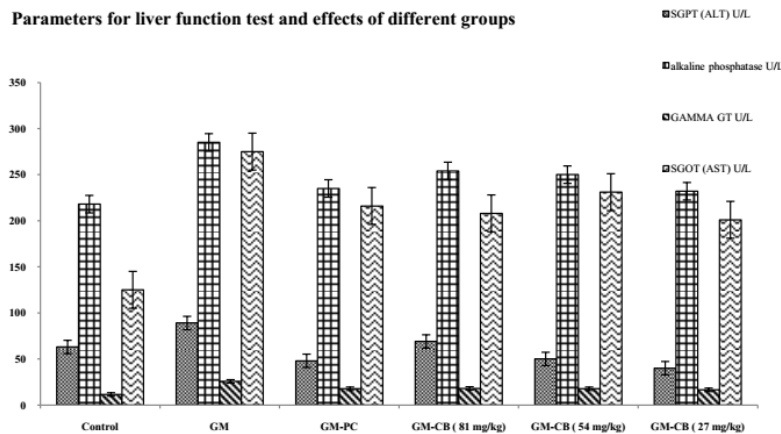


Fig. 4 Effects of different groups on liver function evaluation

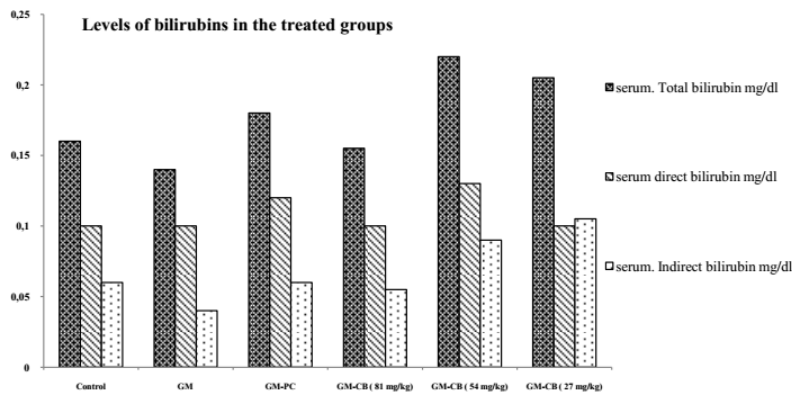


Fig. 5 Effects of different groups on different bilirubin on liver function evaluation

2.2. Histopathological Studies

2.2.1. Kidney

The kidney sections of the control group rats only exhibited a normal histological structure of renal tubules and glomeruli. Prominent macula densa and

distal convoluted tubules were also noticed (Fig. 6a). The kidney section of the GM-treated group rats expressed variable and severe histological alteration. Lymphocyte infiltration, degeneration in the epithelial cell wall, dilated and edematous interstitium, and tubular epithelial necrosis were clearly observed.

Overall architecture showed marked inflammation, periarteriolar hemorrhages, and damaged renal glomeruli. Congested and dilated capillaries with degenerated and damaged glomerular were seen (Fig. 6b). The kidney section of the group of rats treated with GM-PC exhibited moderate lymphocyte infiltration in proximal convoluted tubules (PCT), mild dilation, and congestion in capillaries. The epithelial lining is slightly ruptured with inflammation (Fig. 6c).

These pathological alterations were observed in groups that received aqueous extract of leaves of *C. bonplandianus* with concurrent administration of GM in different doses (81, 54 and 27 mg/kg). The alleviated changes were indicated in low doses of aqueous leave extract. Renal glomeruli showed normal architecture with intact bowman capsules and minimal inflammation in GM-CB (27 mg/Kg) group (fig. 7c). The kidney section of rats of group GM-CB (54 mg/Kg) indicated mild damaged in epithelial cells and moderate inflammation, normal renal architecture, and intact bownans capsules (Fig. 7b). The kidney section of rats that were treated with high doses i.e., GM-CB (81 mg/Kg) showed moderate inflammatory cell infiltration, minor hemorrhages, and mild thickness in the epithelial lining (fig. 7a).

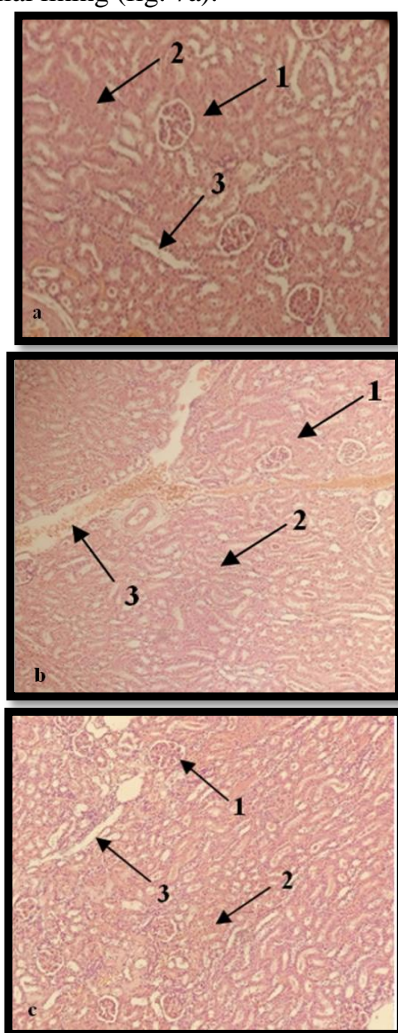


Fig. 6 Photomicrograph of 5-micron thick H & E-stained paraffin section from kidney rats: 1 - Bowman's capsule, 2 - Renal tubules, 3 - Interstitium (a: Control; b: GM; c: PC-GM)

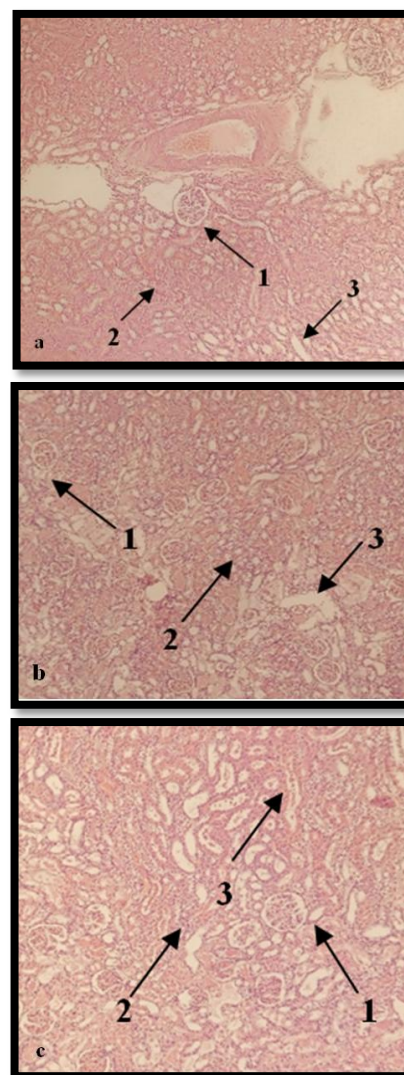


Fig. 7 Photomicrograph of 5 micron thick H & E-stained paraffin section from kidney rats: 1 - Bowman's capsule, 2 - Renal tubules, 3 - Interstitium (a: GM-CBn (81 mg/kg); b: GM-CB (54 mg/kg); c: GM-CB (27 mg/kg))

2.2.2. Liver

Histopathological analysis of the liver of albino rats was carried out to determine the changes in the normal architecture of the liver with a treated sample along the standard drug, to evaluate the degree of damaged cells compared with untreated rat's liver. In the control groups, overall architecture seems intact and normal with normal diameter of the central vein and sinusoids (fig. 8a). In the GM-treated group, dilated, congested and distorted central and portal veins were seen. Moderate inflammatory cell infiltration, sinusoidal dilation and hemorrhages and ruptured hepatic architecture were observed (fig. 8b). 3 portal veins were observed in GM-PC group (fig. 8c).

The series of groups treated with aqueous extracts of *C. bonplandianus* with different doses and concurrent administration of GM showed the following results:

In GM-CB (81 mg/Kg) normal diameter of the central vein with severe congestion has been indicated.

Disturbed portal triad, dilation, and inflammation in the portal vein with moderate hemorrhages were observed. Sinusoidal dilation, distorted, and necrotized hepatocytes were also present (fig. 9a). Group GM-CB (54mg/Kg) showed moderate dilation in the central and portal veins and inflammation in the portal triad (fig. 9b). GM-CB (27mg/Kg) group displayed intact hepatic architecture and moderate sinusoidal dilation with no hemorrhages. A normal portal triad with mild central vein dilation was noted (fig. 9c).

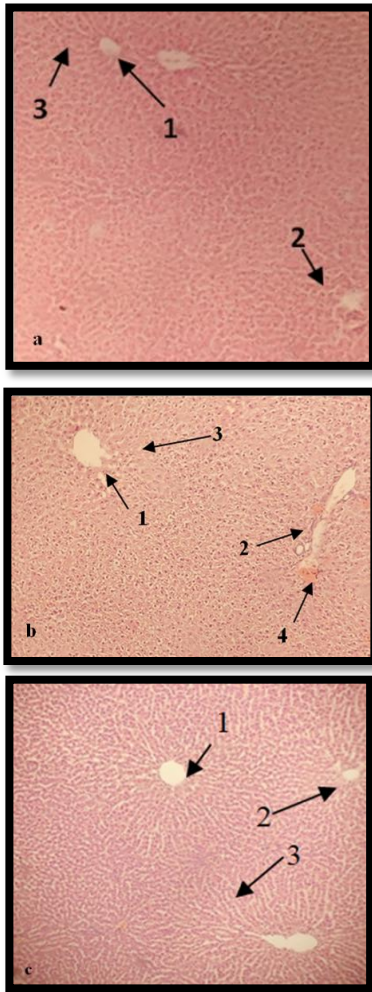


Fig. 8 Photomicrograph of 5-micron thick H & E-stained paraffin section from the liver of rats (a: Control: 1 - Central vein, 2 - Portal vein, 3 - Hepatic architecture; b: GM: 1 - Central vein, 2 - Portal triad, 3 - Necrosis, 4 - Hemorrhages; c: GM-PC: 1 - Central vein, 2 - Portal triad, 3 - Hepatic architecture)

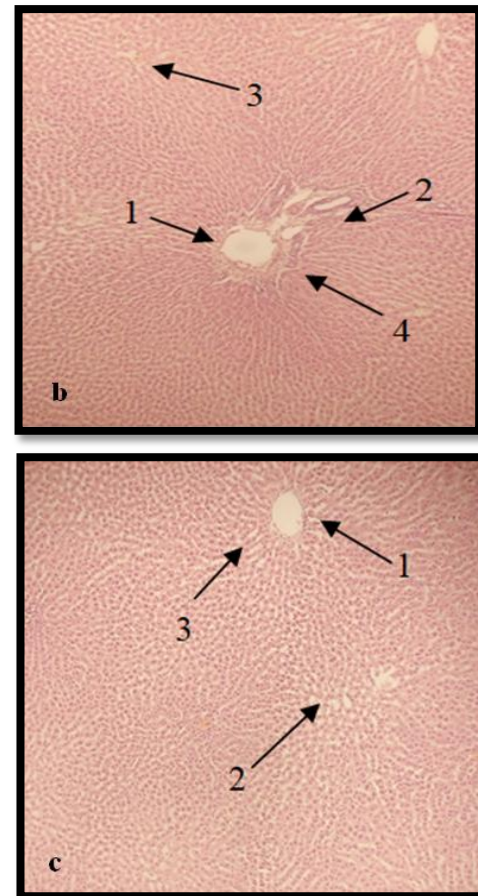
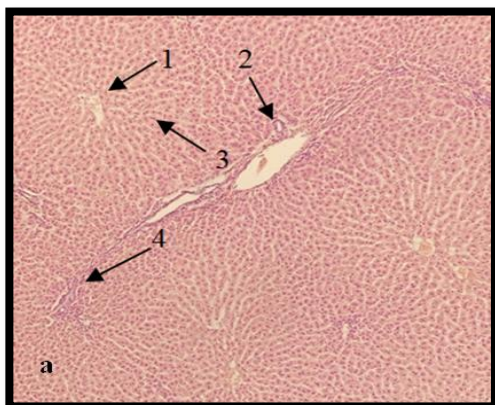


Fig. 9 Photomicrograph of 5-micron thick H & E-stained paraffin section from the liver of rats. a: GM-CB (81 mg/kg): 1 - Central vein, 2 - Portal triad, 3 - Sinusoid, 4 - Necrosis; b: GM-CB (54 mg/kg): 1 - Central vein, 2 - Portal triad, 3 - Sinusoid, 4 - Inflammation; c: GM-CB (27 mg/kg): 1 - Central vein, 2 - Portal triad, 3 - Hepatic architecture

3. Discussion

Gentamicin is the most widely used antibiotic as an effective aminoglycoside for various pathogens. Recent research shown that it may cause hepatorenal injuries [29]. In this study gentamicin-induces liver and kidney damage was selected for estimating the hepatoprotective and nephroprotective effects of aqueous extract of leaves of *C. bonplandianus*. Paracalciteriol was used as a reference or standard drug [23, 29]. Routine analysis of the above-mentioned parameters consolidates and confirms the kidney function [30, 31]. No significant changes were observed in all groups that indicated that administration of gentamicin alone and in combination may not alter levels of sodium, potassium, chloride, and uric acid (Table 1). However, significant reductions in creatinine and urea were exhibited in the treated groups compared with the group that received gentamicin alone, which showed that improvement in kidney function in the treated groups (Fig. 2, 3, 6 and 7). Similarly, most of the liver diseases are associated with the used of antibiotics such as gentamicin, carbon tetrachloride, or intake of alcohol etc. [32]. In the present study, the hepatoprotective and nephroprotective effects of leaves extracts are may be due to the abundance of flavonoids

and polyphenolic compounds in the aerial parts of plants [33, 34]. These compounds possess antioxidant and free radical scavenging properties.

4. Conclusion

The hepatoprotective and nephroprotective effects based on histopathological screening were observed in the underlying study. Limited literature is available on the renal and hepatic protection of leaves extract of *Croton bonplandianus*. The aforementioned findings led to undertake that the promising hepatoprotective and nephroprotective therapeutic potential to reduce GM-induced renal and hepatic functional and structural abnormalities. Hepatotoxicity and nephrotoxicity induced by GM are reduced by antioxidant action. The production of free radicles due to metabolic reactions lead toward the oxidative stresses. The leaf extract holds the free radical scavenging activities consequently, exhibiting anti-oxidant role. The study provides a scientific justification for the usage of the leaves extracts of *C. bonplandianus*. However, in the future, detailed molecular studies with isolation of active compounds will be deemed to prove the mentioned activities. Furthermore, extensive preclinical and clinical investigation in different ways must be complemented toward the diversity of a new drug molecule that may be used as an active moiety in the pharmaceutical industries.

References

[1] AKHTAR M. F., SALEEM A., and SALEEM M. A comprehensive review on ethnomedicinal, pharmacological and phytochemical basis of anticancer medicinal plants in Pakistan. *Current Cancer Drug Targets*, 2019, 19(2): 120-151. <https://doi.org/10.2174/1568009618666180706164536>

[2] ADEWUSI E., & AFOLAYAN A. J. A review of natural products with hepatoprotective activity. *Journal of Medicinal Plants Research*, 2010, 4(13): 1318-1334. <https://doi.org/10.3748/wjg.v20.i40.14787>

[3] BABAEENEZHAD E., NOURYAZDAN N., NASRI M., AHMADVAND H., and SARABI M. M. Cinnamic acid ameliorate gentamicin-induced liver dysfunctions and nephrotoxicity in rats through induction of antioxidant activities. *Heliyon*, 2021, 7(7): e07465. <https://doi.org/10.1016/j.heliyon.2021.e07465>

[4] YASIN H., MAHMUD S., ABRAR H., ARSALAN A., JAHAN N., FATIMA K., BANO R., HUSSAIN R. A., and ISLAM M. Anti-inflammatory and histopathological studies of leaves extracts of *Croton bonplandianus* in Albino rats. *Pakistan Journal of Pharmaceutical Sciences*, 2021, 34(2): 747-753. <https://pubmed.ncbi.nlm.nih.gov/34275810/>

[5] JADHAV V., GHAWATE V., and SINGH N. Chemical composition and antibacterial activity of *Croton bonplandianum* Baill leaves. *International Journal of Pharmacy & Life Sciences*, 2020, 11(7).

[6] DIVYA S., NAVEEN KRISHNA K., RAMACHANDRAN S., and DHANARAJU M. Wound healing and in vitro antioxidant activities of *Croton bonplandianum* leaf extract in rats. *Global Journal of Pharmacology*, 2011, 5(3): 159-163.

[https://www.idosi.org/gjp/5\(3\)11/8.pdf](https://www.idosi.org/gjp/5(3)11/8.pdf)

[7] SURESH M., ALFONISAN M., ALTURAIKI W., AL ABOODY M. S., ALFAIZ F. A., PREMANATHAN M., VIJAYAKUMAR R., UMAMAGHESWARI K., AL GHAMDI S., and ABDALLAH ALSAGABY S. Investigations of bioactivity of *Acalypha indica* (L.), *Centella asiatica* (L.) and *croton bonplandianus* (Baill) against multidrug resistant bacteria and cancer cells. *Journal of Herbal Medicine*, 2020, 28(2): 100359. <http://dx.doi.org/10.1016/j.hermed.2020.100359>

[8] DE OLIVEIRA A. M., DE FREITAS A. F. S., COSTA W. K., MACHADO J. C. B., BEZERRA I. C. F., FERREIRA M. R. A., GUEDES PAIVA P. M., NAPOLEÃO T. H., and LIRA SOARES L. A. Flavonoid-rich fraction of *Croton blanchetianus* Baill. (Euphorbiaceae) leaves: Chemical profile, acute and subacute toxicities, genotoxicity and antioxidant potential. *South African Journal of Botany*, 2022, 144(5): 238-249. <http://dx.doi.org/10.1016/j.sajb.2021.08.040>

[9] SOZEN H., CELIK O. I., CETIN E. S., YILMAZ N., AKSOZEK A., TOPAL Y., CIGERCI I. H., and BEYDILLI H. Evaluation of the protective effect of silibinin in rats with liver damage caused by itraconazole. *Cell Biochemistry and Biophysics*, 2015, 71(2): 1215-1223. <https://doi.org/10.1007/s12013-014-0331-8>

[10] CHOUDHURY D., & AHMED Z. Drug-induced nephrotoxicity. *The Medical clinics of North America*, 1997, 81(3): 705-717. [https://doi.org/10.1016/s0025-7125\(05\)70541-1](https://doi.org/10.1016/s0025-7125(05)70541-1)

[11] COLOMBO A., MERONI C. A., CIPOLLA C. M., and CARDINALE D. Managing cardiotoxicity of chemotherapy. *Current Treatment Options in Cardiovascular Medicine*, 2013, 15(4): 410-424. <https://doi.org/10.1007/s11936-013-0248-3>

[12] RAD A. K., MOHEBBATI R., and HOSSEINIAN S. Drug-induced nephrotoxicity and medicinal plants. *Iranian Journal of Kidney Diseases*, 2017, 11(3): 169. <https://pubmed.ncbi.nlm.nih.gov/28575877/>

[13] BULBOACĂ A. E., PORFIRE A. S., RUS V., NICULA C. A., BULBOACĂ C. A., and BOLBOACĂ S. D. Protective effect of liposomal epigallocatechin-gallate in experimental gentamicin-induced hepatotoxicity. *Antioxidants*, 2022, 11(2): 412. <https://doi.org/10.3390/antiox11020412>

[14] JAFARIPOUR L., NASERZADEH R., AHMADVAND H., HADIPOUR MORADI F., GHOBADI K., ALIZAMANI E., and NOURYAZDAN N. Effects of L-glutamine on oxidative stress in gentamicin induced hepatotoxicity rats. *Journal of Kerman University of Medical Sciences*, 2019, 26(1): 36-42. https://www.researchgate.net/publication/332950615_Effects_of_L-glutamine_on_oxidative_stress_in_gentamicin_induced_hepatotoxicity_rats

[15] AL-KURAIISHY H. M., AL-GAREEB A., and RASHEED H. A. The antioxidant and anti-inflammatory effects of curcumin contribute into attenuation of acute gentamicin-induced nephrotoxicity in rats. *Asian Journal of Pharmaceutical and Clinical Research*, 2019, 12(3): 466-468. <http://dx.doi.org/10.22159/ajpcr.2019.v12i3.30875>

[16] KHAKSARI M., ESMALI S., ABEDLOO R., and KHASTAR H. Palmatine ameliorates nephrotoxicity and hepatotoxicity induced by gentamicin in rats. *Archives of Physiology and Biochemistry*, 2021, 127(3): 273-278.

- <https://doi.org/10.1080/13813455.2019.1633354>
- [17] AMALIA R., AULIFA D. L., ZAIN D. N., PEBIANSYAH A., and LEVITA J. The cytotoxicity and nephroprotective activity of the ethanol extracts of angelica keiskei Koidzumi stems and leaves against the NAPQI-induced human embryonic kidney (HEK293) cell line. *Evidence-Based Complementary and Alternative Medicine*, 2021, 2021: 6458265. <https://doi.org/10.1155/2021/6458265>
- [18] SADASHIVA C., HUSSAIN H. F., and NANJUNDAIAH S. Evaluation of hepatoprotective, antioxidant and cytotoxic properties of aqueous extract of turmeric rhizome (Turmesac®). *Journal of Medicinal Plants Research*, 2019, 13(17): 423-430. <https://doi.org/10.5897/JMPR2019.6824>
- [19] WINTER C. A., RISLEY E. A., and NUSS G. W. Carrageenin-induced edema in hind paw of the rat as an assay for antiinflammatory drugs. *Proceedings of the Society for Experimental Biology and Medicine*, 1962, 111(3): 544-547. <https://doi.org/10.3181/00379727-111-27849>
- [20] DHRUVE P., NAUMAN M., KALE R. K., and SINGH R. P. A novel hepatoprotective activity of Alangium salviifolium in mouse model. *Drug and Chemical Toxicology*, 2022, 45(2): 576-588. <https://doi.org/10.1080/01480545.2020.1733593>
- [21] MOHAMED A. S., GAMAL M. A., EL-ZAYAT E., and SULIMAN H. Sea cucumbers-saponin ameliorates hepatorenal toxicity induced by Gentamicin in rats. *GSC Biological and Pharmaceutical Sciences*, 2021, 14(3): 129-136. <http://dx.doi.org/10.30574/gscbps.2021.14.3.0069>
- [22] ABRAR H., NAQVI S., AHMED M. R., ALI A. B., and YASIN H. Ameliorating Effect of a Beta-Blocker, Propranolol on Carbamazepine-Induced Hepatotoxicity in Rabbits. *Pakistan Journal of Zoology*, 2019, 51(1): 341-346. <http://dx.doi.org/10.17582/journal.pjz/2019.51.1.341.346>
- [23] OKOKON J. E., NWAFOR P. A., and NOAH K. Nephroprotective effect of Croton zambesicus root extract against gentimicin-induced kidney injury. *Asian Pacific Journal of Tropical Medicine*, 2011, 4(12): 969-972. [https://doi.org/10.1016/s1995-7645\(11\)60228-9](https://doi.org/10.1016/s1995-7645(11)60228-9)
- [24] AYZA M. A., RAJKAPOOR B., WONDAFRASH D. Z., and BERHE A. H. Protective Effect of Croton macrostachyus (Euphorbiaceae) Stem Bark on Cyclophosphamide-Induced Nephrotoxicity in Rats. *Journal of Experimental Pharmacology*, 2020, 12: 275. <https://doi.org/10.2147/jep.s260731>
- [25] IQBAL S. M., HUSSAIN L., HUSSAIN M., AKRAM H., ASIF M., JAMSHED A., SALEEM A., and SIDDIQUE R. Nephroprotective potential of a standardized extract of bambusa arundinacea: in vitro and in vivo studies. *ACS Omega*, 2022, 7(21): 18159-18167. <https://doi.org/10.1021/acsomega.2c02047>
- [26] THANG HOANG D., HIEN TRUONG T. T., VIET DUC N., ANH HOANG L. T., DO T. T., VINH L. B., YANG S. Y., GAO D., and LE T. Hepatoprotective Effects of Extract of Helicteres hirsuta Lour. on Liver Fibrosis Induced by Carbon Tetrachloride in Rats. *Applied Sciences*, 2021, 11(18): 8758. <http://dx.doi.org/10.3390/app11188758>
- [27] URRUTIA-HERNÁNDEZ T. A., SANTOS-LÓPEZ J. A., BENEDÍ J., SÁNCHEZ-MUNIZ F. J., VELÁZQUEZ-GONZÁLEZ C., DE LA O-ARCINIEGA M., JARAMILLO-MORALES O. A., and BAUTISTA M. Antioxidant and hepatoprotective effects of croton hypoleucus extract in an induced-necrosis model in rats. *Molecules*, 2019, 24(14): 2533. <https://doi.org/10.3390/molecules24142533>
- [28] DUTTA S., CHAKRABORTY A. K., DEY P., KAR P., GUHA P., SEN S., KUMAR A., SEN A., and CHAUDHURI T. K. Amelioration of CCl4 induced liver injury in Swiss albino mice by antioxidant rich leaf extract of Croton bonplandianus Baill. *PLoS One*, 2018, 13(4): e0196411. <https://doi.org/10.1371/journal.pone.0196411>
- [29] ALI F. E., HASSANEIN E. H., BAKR A. G., EL-SHOUBA E. A., EL-GAMAL D. A., MAHMOUD A. R., and ABD-ELHAMID T. H. Ursodeoxycholic acid abrogates gentamicin-induced hepatotoxicity in rats: Role of NF- κ B-p65/TNF- α , Bax/Bcl-xl/Caspase-3, and eNOS/iNOS pathways. *Life Sciences*, 2020, 254: 117760. <https://doi.org/10.1016/j.lfs.2020.117760>
- [30] BAGSHAW S. M., and GIBNEY R. T. N. Conventional markers of kidney function. *Critical Care Medicine*, 2008, 36(4): S152-S158. <https://doi.org/10.1097/ccm.0b013e318168c613>
- [31] DEN BAKKER E., GEMKE R. J., and BÖKENKAMP A. Endogenous markers for kidney function in children: a review. *Critical Reviews in Clinical Laboratory Sciences*, 2018, 55(3): 163-183. <https://doi.org/10.1080/10408363.2018.1427041>
- [32] HSU L.-S., HO H.-H., LIN M.-C., CHYAU C.-C., PENG J.-S., and WANG C.-J. Mulberry water extracts (MWEs) ameliorated carbon tetrachloride-induced liver damage in rats. *Food and Chemical Toxicology*, 2012, 50(9): 3086-3093. <https://doi.org/10.1016/j.fct.2012.05.055>
- [33] YASIN H., MAHMUD S., RIZWANI G. H., PERVEEN R., ABRAR H., and FATIMA K. Effects of aqueous leaves extract of Holoptelea integrifolia (Roxb) Planch on liver and kidney histopathology of albino rats. *Pakistan Journal of Pharmaceutical Sciences*, 2019, 32(2): 569-573. <https://pubmed.ncbi.nlm.nih.gov/31081768/>
- [34] OKOKON J. E., & NWAFOR P. A. Antiplasmodial activity of root extract and fractions of Croton zambesicus. *Journal of Ethnopharmacology*, 2009, 121(1): 74-78. <https://doi.org/10.1016/j.jep.2008.09.034>

参考文献:

- [1] AKHTAR M. F., SALEEM A., 和 SALEEM M. 巴基斯坦抗癌药用植物的民族药理学、药理学和植物化学基础综合综述。当前癌症药物靶点, 2019年, 19(2): 第 120-151 页. <https://doi.org/10.2174/1568009618666180706164536>
- [2] ADEWUSI E., & AFOLAYAN A. J. 具有保肝活性的天然产物的综述。药用植物研究杂志, 2010, 4(13): 第 1318-1334页. <https://doi.org/10.3748/wjg.v20.i40.14787>
- [3] BABAEENEZHAD E., NOURYAZDAN N., NASRI M., AHMADVAND H., 和 SARABI M. M. 肉桂酸通过诱导抗氧化活性改善庆大霉素诱导的大鼠肝功能障碍和肾毒性。赫利永, 2021年, 7(7): 文章电子0 7465. <https://doi.org/10.1016/j.heliyon.2021.e07465>
- [4] YASIN H., MAHMUD S., ABRAR H., ARSALAN A., JAHAN N., FATIMA K., BANO R., HUSSAIN R. A., 和 ISLAM M. 巴豆叶提取物对白化病大鼠的抗炎和组织病理学研究。巴基斯坦药理学杂志, 2021, 34(2): 第 747-753 页. <https://pubmed.ncbi.nlm.nih.gov/34275810/>
- [5] JADHAV V., GHAWATE V., 和 SINGH N. 巴豆

叶的化学成分和抗菌活性。国际药学与生命科学杂志, 2020, 11(7).

[6] DIVYA S., NAVEEN KRISHNA K., RAMACHANDRAN S., 和 DHANARAJU M. 巴豆邦普兰叶提取物在大鼠中的伤口愈合和体外抗氧化活性。全球药理学杂志, 2011, 5(3): 第159-163页。

[https://www.idosi.org/gjp/5\(3\)11/8.pdf](https://www.idosi.org/gjp/5(3)11/8.pdf)

[7] SURESH M., ALFONISAN M., ALTURAIKI W., AL ABOODY M. S., ALFAIZ F. A., PREMANATHAN M., VIJAYAKUMAR R., UMAMAGHESWARI K., AL GHAMDI S., 和 ABDALLAH ALSAGABY S. 研究刺桐、积雪草和巴豆邦普兰迪亚努斯(保尔)对多重耐药细菌和癌细胞的生物活性。草药杂志, 2020年, 28(2): 文章100359。

<http://dx.doi.org/10.1016/j.hermed.2020.100359>

[8] DE OLIVEIRA A. M., DE FREITAS A. F. S., COSTA W. K., MACHADO J. C. B., BEZERRA I. C. F., FERREIRA M. R. A., GUEDES PAIVA P. M., NAPOLEÃO T. H., 和 LIRA SOARES L. A. 巴豆富含类黄酮的部分。(大戟科)叶子: 化学特征、急性和亚急性毒性、遗传毒性和抗氧化潜力。南非植物学杂志, 2022, 144(5): 第238-249页。

<http://dx.doi.org/10.1016/j.sajb.2021.08.040>

[9] SOZEN H., CELIK O. I., CETIN E. S., YILMAZ N., AKSOZEK A., TOPAL Y., CIGERCI I. H., 和 BEYDILLI H.

水飞蓟宾对伊曲康唑致肝损伤大鼠的保护作用评价。细胞生物化学和生物物理学, 2015, 71(2): 第1215-1223页。<https://doi.org/10.1007/s12013-014-0331-8>

[10] CHOUDHURY D., & AHMED Z. 药物引起的肾毒性。北美的医疗诊所, 1997, 81(3): 第705-717页。[https://doi.org/10.1016/s0025-7125\(05\)70541-1](https://doi.org/10.1016/s0025-7125(05)70541-1)

[11] COLOMBO A., MERONI C. A., CIPOLLA C. M., 和 CARDINALE D. 管理化疗的心脏毒性。目前心血管医学的治疗选择, 2013, 15(4): 第410-424页。<https://doi.org/10.1007/s11936-013-0248-3>

[12] RAD A. K., MOHEBBATI R., 和 HOSSEINIAN S. 药物引起的肾毒性和药用植物。伊朗肾脏病杂志, 2017, 11(3): 文章169。<https://pubmed.ncbi.nlm.nih.gov/28575877/>

[13] BULBOACĂ A. E., PORFIRE A. S., RUS V., NICULA C. A., BULBOACĂ C. A., 和 BOLBOACĂ S. D. 脂质体表没食子儿茶素没食子酸酯对实验性庆大霉素诱导的肝毒性的保护作用。抗氧化剂, 2022, 11(2): 文章412。<https://doi.org/10.3390/antiox11020412>

[14] JAFARIPOUR L., NASERZADEH R., AHMADVAND H., HADIPOUR MORADI F., GHOBADI K., ALIZAMANI E., 和 NOURYAZDAN N. L-谷氨酰胺对庆大霉素肝毒性大鼠氧化应激的影响。克尔曼医科大学学报, 2019, 26(1): 第36-42页。

https://www.researchgate.net/publication/332950615_Effects_of_L-glutamine_on_oxidative_stress_in_gentamicin_induced_hepatotoxicity_rats

[15] AL-KURAIISHY H. M., AL-GAREEB A., 和 RASHEED H. A. 姜黄素的抗氧化和抗炎作用有助于减轻庆大霉素诱导的大鼠急性肾毒性。亚洲制药与临床研究杂志, 2019, 12(3):

第466-468页。

<http://dx.doi.org/10.22159/ajpcr.2019.v12i3.30875>

[16] KHAKSARI M., ESMAILI S., ABEDLOO R., 和 KHASTAR H. 巴马汀可改善庆大霉素在大鼠中引起的肾毒性和肝毒性。生理生化档案, 2021, 127(3): 第273-278页。<https://doi.org/10.1080/13813455.2019.1633354>

[17] AMALIA R., AULIFA D. L., ZAIN D. N., PEBIANSYAH A., 和 LEVITA J. 当归 庆经已经恋爱了茎和叶的乙醇提取物对否-乙酰基-对苯醌亚胺诱导的人胚肾(HEK293)细胞系的细胞毒性和肾保护活性。循证补充和替代医学, 2021, 2021: 文章6458265。<https://doi.org/10.1155/2021/6458265>

[18] SADASHIVA C., HUSSAIN H. F., 和 NANJUNDAIAH S. 姜黄根茎(图尔梅萨克®)水提取物的保肝、抗氧化和细胞毒性特性的评估。药用植物研究杂志, 2019, 13(17): 第423-430页。<https://doi.org/10.5897/JMPR2019.6824>

[19] WINTER C. A., RISLEY E. A., 和 NUSS G. W. 角叉菜胶诱导的大鼠后爪水肿作为抗炎药物的测定。实验生物学和医学学会会刊, 1962, 111(3): 第544-547页。<https://doi.org/10.3181/00379727-111-27849>

[20] DHRUVE P., NAUMAN M., KALE R. K., 和 SINGH R. P. 八角枫在鼠模型中的新型保肝活性。药物和化学毒理学, 2022, 45(2): 第576-588页。<https://doi.org/10.1080/01480545.2020.1733593>

[21] MOHAMED A. S., GAMAL M. A., EL-ZAYAT E., 和 SULIMAN H. 海参皂苷改善庆大霉素诱导的大鼠肝肾毒性。全球学术交流生物和制药科学, 2021, 14(3): 第129-136页。<http://dx.doi.org/10.30574/gscbps.2021.14.3.0069>

[22] ABRAR H., NAQVI S., AHMED M. R., ALI A. B., 和 YASIN H. β -受体阻滞剂普萘洛尔对卡马西平诱导的家兔肝毒性的改善作用。巴基斯坦动物学杂志, 2019, 51(1): 第341-346页。<http://dx.doi.org/10.17582/journal.pjz/2019.51.1.341.346>

[23] OKOKON J. E., NWAFOR P. A., 和 NOAH K. 巴豆根提取物对庆大霉素引起的肾损伤的肾保护作用。亚太热带医学杂志, 2011, 4(12): 第969-972页。[https://doi.org/10.1016/s1995-7645\(11\)60228-9](https://doi.org/10.1016/s1995-7645(11)60228-9)

[24] AYZA M. A., RAJKAPOOR B., WONDAFRASH D. Z., 和 BERHE A. H. 巴豆(大戟科)茎皮对环磷酰胺诱导的大鼠肾毒性的保护作用。实验药理学杂志, 2020, 12: 文章275。<https://doi.org/10.2147/jep.s260731>

[25] IQBAL S. M., HUSSAIN L., HUSSAIN M., AKRAM H., ASIF M., JAMSHED A., SALEEM A., 和 SIDDIQUE R.

野竹标准化提取物的肾保护潜力: 体外和体内研究。美国化学学会欧米茄, 2022, 7(21): 第18159-18167页。<https://doi.org/10.1021/acsomega.2c02047>

[26] THANG HOANG D., HIEN TRUONG T. T., VIET DUC N., ANH HOANG L. T., DO T. T., VINH L. B., YANG S. Y., GAO D., 和 LE T. 长毛毛虫提取物的保肝作用。对大鼠四氯化碳诱导的肝纤维化的影响。应用科学, 2021, 11(18): 文章8758。

<http://dx.doi.org/10.3390/app11188758>

[27] URRUTIA-HERNÁNDEZ T. A., SANTOS-LÓPEZ J. A., BENEDÍ J., SÁNCHEZ-MUNIZ F. J., VELÁZQUEZ-GONZÁLEZ C., DE LA O-ARCINIEGA M., JARAMILLO-MORALES O. A., 和 BAUTISTA M. 巴豆小白鱼提取物在大鼠诱导性坏死模型中的抗氧化和保肝作用。分子, 2019, 24(14): 文章2533.

<https://doi.org/10.3390/molecules24142533>

[28] DUTTA S., CHAKRABORTY A. K., DEY P., KAR P., GUHA P., SEN S., KUMAR A., SEN A., 和 CHAUDHURI T. K. 巴豆

富含抗氧化剂的叶提取物改善四氯化碳诱导的瑞士白化小鼠肝损伤。公共科学图书馆一号, 2018, 13(4): 文章e0196411.

<https://doi.org/10.1371/journal.pone.0196411>

[29] ALI F. E., HASSANEIN E. H., BAKR A. G., EL-SHOURA E. A., EL-GAMAL D. A., MAHMOUD A. R., 和 ABD-ELHAMID T. H.

巴豆熊去氧胆酸消除庆大霉素诱导的大鼠肝毒性: NF-

κB-p65/肿瘤坏死因子-α、乙细胞淋巴瘤2相关X蛋白/

B细胞淋巴瘤-特大/蛋白酶-

3和内皮/诱导型一氧化氮合酶通路的作用。生命科学, 2020, 254: 文章117760.

<https://doi.org/10.1016/j.lfs.2020.117760>

[30] BAGSHAW S. M., 和 GIBNEY R. T. N. 肾功能的常规标志物。重症监护医学, 2008, 36(4): 第S152-S158页.

<https://doi.org/10.1097/ccm.0b013e318168c613>

[31] DEN BAKKER E., GEMKE R. J., 和 BÖKENKAMP A.

儿童肾功能的内源性标志物: 综述。临床实验室科学的批判性评论, 2018, 55(3): 第163-183页.

<https://doi.org/10.1080/10408363.2018.1427041>

[32] HSU L.-S., HO H.-H., LIN M.-C., CHYAU C.-C., PENG J.-S., 和 WANG C.-J. 桑葚水提取物可改善四氯化碳引起的大鼠肝损伤。食品和化学毒理学, 2012, 50(9): 第3086-3093页.

<https://doi.org/10.1016/j.fct.2012.05.055>

[33] YASIN H., MAHMUD S., RIZWANI G. H., PERVEEN R., ABRAR H., 和 FATIMA K. 全叶全叶木(罗素)平面叶水提取物对白化大鼠肝肾组织病理学的影响。巴基斯坦药学杂志, 2019, 32(2): 第569-573页. <https://pubmed.ncbi.nlm.nih.gov/31081768/>

[34] OKOKON J. E., & NWAFOR P. A. 巴豆根提取物和部分的抗疟原虫活性。民族药理学杂志, 2009, 121(1): 第74-78页.

<https://doi.org/10.1016/j.jep.2008.09.034>