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Soaking Time in Lime Solution Increases the Antioxidant Activity, Antidiabetic Activity, and Consumer Acceptance Level of *Sargassum Polycystum* Seaweed Tea

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Abstract: The brown seaweed *Sargassum polycystum* contains bioactive compounds, including antioxidants and antidiabetes, letting it be a functional food ingredient such as seaweed tea. However, seaweed tea is less preferred by consumers because of its fishy smell, which must be reduced using various methods, including soaking in a lime solution. This study aims to determine the effect of the soaking time in lime solution on the antioxidant activity, antidiabetic activity, and consumer acceptance of *S. polycystum* seaweed tea. In this study, the seaweed was soaked in a lime solution at pH 5 at 85°C for 0, 4, 8, 12, and 16 minutes. Analysis was conducted on *S. polycystum* seaweed tea to obtain the moisture content, total phenol, antioxidant activity (inhibition of 1,1-diphenyl-2-picrylhydrazyl (DPPH) and ferric reduction antioxidant power (FRAP)), the inhibitory activity of α -glucosidase, and consumer preference test using hedonic. The results show that the soaking time in the lime solution at pH 5 had no effect on the water content but had a significant effect ($p < 0.05$) on the total phenol, antioxidant and antidiabetic activities and consumer preference of *S. polycystum* seaweed tea. The best treatment was obtained at 16 minutes of soaking time with a water content of $4.68 \pm 0.12\%$, a total phenol content of 74.53 ± 0.59 mg GAE/g, a DPPH inhibitory activity of $52.70 \pm 0.86\%$, a FRAP value of 123.94 ± 1.39 μ M/g, an α -glucosidase inhibitory activity $51.56 \pm 0.70\%$, and consumer preference levels for appearance, color, flavor, taste and overall of 4.63 ± 0.74 , 4.49 ± 0.75 , 4.39 ± 1.07 , 4.43 ± 1.03 , and 4.49 ± 0.11 , respectively.

Keywords: lime, *Sargassum polycystum*, seaweed tea, soaking.

在石灰溶液中浸泡时间可提高马尾藻多囊海藻茶的抗氧化活性、抗糖尿病活性和消费者接受度

摘要: 棕色海藻马尾藻多囊含有生物活性化合物, 包括抗氧化剂和抗糖尿病剂, 使其成为海藻茶等功能性食品成分。然而, 消费者不太喜欢海藻茶, 因为它有鱼腥味, 必须使用各种方法来减少这种气味, 包括浸泡在石灰溶液中。本研究旨在确定在石灰溶液中的浸泡时间对多囊海藻茶的抗氧化活性、抗糖尿病活性和消费者接受度的影响。在这项研究中, 海藻在 85°C、酸碱度值为 5 的石灰溶液中浸泡 0、4、8、12 和 16 分钟。对多囊海藻茶进行水分含量、总酚、抗氧化活性 (1,1-二苯基-2-苦基肼的抑制和铁还原抗氧化能力的抑制)、 α -葡萄糖苷酶, 以及使用享乐的消费者偏好测试。结果表明, 在 pH 的潜力值为 5 的石灰溶液中浸泡时间对含水量没有影响, 但对多囊海藻茶的总酚、抗氧化和抗糖尿病活性以及消费者偏好有显著影响 ($p < 0.05$)。最佳处理为浸泡时间 16 分钟, 含水量为 $4.68 \pm 0.12\%$, 总酚含量为 74.53 ± 0.59 毫克每克没食子酸当量, 二苯基-2-苦基肼抑制活性为 $52.70 \pm 0.86\%$, 铁还原抗氧化能力值为 123.94 ± 1.39 微米/克, α -葡萄糖苷酶抑制活性 $51.56 \pm 0.70\%$, 消费者对外观、颜色、风味、味道和总体的偏好水平为 4.63 ± 0.74 、 4.49 ± 0.75 、 4.39 ± 1.07 、 4.43 、 ± 4.03 分别为 0.11。

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关键词：生石灰、海藻多囊、紫菜茶，浸泡。

1. Introduction

Seaweed is a very abundant biological resource, including brown seaweed. One of the abundant brown seaweed species in the world is *the Sargassum polycystum*. Many studies show that *S. polycystum* has bioactivity, including antioxidant [1], antibacterial [2], anti-inflammatory [3], antistress [4], anticoagulant [5], antidiabetic [6], and anticancer activities [7]. Thus, *S. polycystum* has a high potential to be used as a food and industrial ingredient.

The use of seaweed in the industrial sector includes the food, beverage, medicine, cosmetics, paper, detergent, paint, and textile industries [8]. One of the uses of seaweed in the beverage industry is the manufacture of seaweed tea. Seaweed tea is a health drink that contains nutrients needed by the body, including antioxidants [9]. However, seaweed tea has limitations: the flavor and taste of seaweed tea are generally not preferred by consumers because of the fishy smell. One alternative to reduce the fishy smell is soaking in boiling water [10]. However, consumer preference remained low, so it was necessary to add other natural ingredients, such as lime solution. Soaking in lime solution at pH 5 for 6 hours could increase *Sargassum filipendula* seaweed tea [9].

Additionally, a soaking time that is too long can reduce the antioxidant activity of a food product [11]. Until now, research on the manufacture of seaweed tea from *S. polycystum* is still very rare. Therefore, this study aims to determine the effect of the soaking time in lime solution on the antioxidant activities, antidiabetic activities, and consumer acceptance of *S. polycystum* seaweed tea.

2. Material and Methods

2.1. Materials

The materials in this research are brown seaweed *S. polycystum* obtained from Sepanjang Beach, Gunungkidul Yogyakarta. The commercial seaweed tea for comparison was obtained from SMEs in Bali. Lime (*Citrus aurantifolia*) was obtained from the Kranggan Market Yogyakarta. Other materials were Folin Ciocalteu reagent (produced by Merck, USA), Na₂CO₃ (produced by Merck, USA), 1,1-diphenyl-2-picrylhydrazyl (DPPH) (produced by Merck, USA), ethanol (produced by Merck, USA), gallic acid (produced by Merck, USA), K₂SO₄ (produced by Merck, USA), 2,4,6-tri (2-pyridyl)-s triazine (TPTZ) (produced by Merck, USA), FeCl₃ (produced by Merck, USA), FeSO₄ (produced by Merck, USA), vitamins C (produced by Merck, USA), acarbose (produced by Sigma-Aldrich, Germany), α -glucosidase

from *Saccharomyces cerevisiae* (produced by Sigma-Aldrich, Germany), and p-nitrophenyl- α -D-glucopyranoside (PNPG) (produced by Sigma Aldrich).

2.2. Preparation of Seaweed *Sargassum Polycystum*

Seaweed samples of *S. polycystum* were taken from Sepanjang Beach, Gunungkidul Yogyakarta, in August 2020. Seaweed was collected by cutting the lower thallus near holdfast using scissors. Then, the samples were washed and separated for morphological identification. The other samples were stored in an icebox to be brought to the laboratory and stored in the refrigerator.

2.3. Manufacture of *Sargassum Polycystum* Seaweed Tea

The manufacture of seaweed tea refers to the research of Sinurat & Suryaningrum [10] with several modifications. 200 g of seaweed *S. polycystum* was washed using clean water and subsequently soaked in 2000 mL of pH 5 lime solution. The pH 5 lime solution was prepared by mixing 2000 mL of water at 85°C and approximately 6.4 mL of lime water. The initial temperature of the water was 85°C, and the temperature decreased during the soaking process. The variations in soaking time were 0 (without soaking), 4, 8, 12, and 16 minutes. After soaking, the seaweed was drained and dried for 24 hours in a baking sheet placed on a terrace and not exposed to direct sunlight. Then, the seaweed was roasted in a frying pan for 15-20 minutes and cut with a knife into pieces 0.5 ± 0.1 cm in size. Before use, seaweed tea was placed in a sealed standing pouch and stored at 4°C.

2.4. Serving of *Sargassum Polycystum* Seaweed Tea

As much as 1 g of seaweed tea was put in a teabag. Brewing was done by placing 1 tea bag in 100 mL of boiling water for 6 minutes. During the brewing process, the teabags were raised and lowered into boiling water 5 times and stirred 2-3 times; then, the teabags were removed from the solution [12]. The seaweed tea solution was used for the total phenol analysis, antioxidant activity analysis, α -glucosidase inhibition test, and consumer preference tests.

2.5. Analysis of the Water Content

The water content was analyzed using a moisture analyzer Ohaus MB120. A sample of 0.5 g was placed on the plate in the moisture content analyzer; then, the appliance was closed. Next, the cells were incubated for 2-20 minutes until the drying process was complete. The displayed results on the moisture content analyzer screen were recorded.

2.6. Total Phenol Analysis

The total phenol analysis refers to the research of Sinurat & Suryaningrum [10]. In total, 1 mL of tea solution was put into a Falcon tube, and 1 mL of 96% ethanol, 5 mL of distilled water, and 0.5 mL of 50% Folin Ciocalteu reagent were added. The mixture was allowed to stand for 5 minutes, and 1 mL of 5% Na₂CO₃ was subsequently added. The mixture was homogenized with vortexing and incubated in the dark for 1 hour. Standard solutions were prepared by making gallic acid solutions with 0, 20, 40, 60, 80, and 100 ppm concentrations. Then, 1 mL of gallic acid solution was taken for each concentration and put into a Falcon tube, 1 mL of 96% ethanol, 5 mL of distilled water, and 0.5 mL of 50% Folin Ciocalteu reagent were added. The mixture was allowed to stand for 5 minutes, and 1 mL of 5% Na₂CO₃ was subsequently added. The mixture was homogenized with vortexing and incubated in the dark for 1 hour. The sample solution's absorbance and standard solution were measured using a UV-Vis spectrophotometer (Lambda 25, PerkinElmer) at a wavelength of 725 nm. The formula to calculate the total phenol content is as follows:

$$\text{Total phenol (mg GAE/g)} = x \cdot \frac{v}{m}$$

where:

x = concentration of the test solution (mg/mL)

v = volume of test solution (mL)

m = mass of the test solution (g)

2.7. DPPH Free Radical Scavenging Assay

The scavenging ability against DPPH free radicals was estimated to evaluate the antioxidant activity of seaweed tea using the modified method of Muthia et al. [13]. The DPPH solution was prepared by dissolving 0.39 mg DPPH powder into 100 mL ethanol. Then, the solution mixture was incubated for 30 minutes at 4°C. Tea samples were prepared by dissolving 1 g of seaweed tea for each treatment and commercial seaweed tea in 100 mL of water. Then, 0.1 mL of the tea sample was placed in a Falcon tube wrapped in aluminum foil, and 1 mL of ethanol and 0.7 mL of DPPH solution were added. A vitamin C sample was prepared by dissolving 1 g of vitamin C in 100 mL of water. Then, 0.1 mL of vitamin C solution was taken and put into a Falcon tube wrapped in aluminum foil, and 1 mL of ethanol and 0.7 mL of DPPH solution were added. The lime solution was prepared by making a solution of lime pH 5 at 85°C. A volume of 0.1 mL of the lime solution at pH 5 was placed in a Falcon tube wrapped in aluminum foil, and 1 mL of ethanol and 0.7 mL of DPPH solution were added. The blanks were 0.7 mL of DPPH and 1 mL of ethanol. The solution mixture (tea sample, vitamin C, lime solution, and blank) was homogenized with vortexing and incubated for 15 minutes at room temperature. The absorbance was measured at a wavelength of 515 nm using a UV-Vis

spectrophotometer. The percent inhibition was calculated using the following formula:

$$\text{Inhibitory activity (\%)} = \frac{(C - D) - (A - B)}{(C - D)} \times 100$$

where A = sample (160 µL of sample + 40 µL of 0.76 mM DPPH); B = sample control (160 µL of sample + 40 µL of distilled water); C = negative control (160 µL of distilled water + 40 µL of 0.76 mM DPPH); and D = blank (200 µL of aqua dest).

2.8. Ferric Reduction Antioxidant Power (FRAP) Assay

This assessment required the method described by Suhaila et al. [14], where Fe³⁺ was reduced to Fe²⁺. A spectrophotometer was used to measure the iron(III) chloride modified into Fe²⁺ complexes at 595 nm wavelength. These changes were observed with a solution color transformation to blue. In addition, an acetate buffer solution with pH 3.6 was formulated by adding 0.775 g of sodium acetate trihydrate (CH₃COON.3H₂O) to 4 mL of concentrated acetic acid, followed by dissolution with distilled water to obtain an exact volume of 250 mL. The yield was stored at 4°C as a stock solution. Subsequently, 10 mM/mL 2,4,6-tripyridil-s-triazine (TPTZ) solution was formulated by dissolving 0.15 g of TPTZ in 40 mM/L HCl to achieve an exact volume of 50 mL. In contrast, the 40 mM/L HCl solution was prepared by dissolving 0.828 mL concentrated HCl in 250 mL distilled water. Therefore, the generated TPTZ solution was reserved at 4°C for use within 24 hours. Additionally, 0.54 g FeCl₃.6H₂O was dissolved in distilled water and made approximately 100 mL to produce a 20-mM/L FeCl₃.6H₂O solution, which was stored for up to 24 hours at 4°C. The FRAP reagent was formulated by combining 25 mL acetate buffer, 2.5 mL TPTZ solution, and 2.5 mL FeCl₃.6H₂O solution (10:1:1) before making approximately 100 mL with distilled water. Furthermore, standard FeSO₄.7H₂O (10,000 µM/L) solutions were produced by dissolving 2.78 g FeSO₄.7H₂O in 1000 mL distilled water before serial dilution to optimal concentrations of 50, 100, 150, 200, 250, and 300 ppm. The *S. polycystum* seaweed tea solution, commercial seaweed tea, and vitamin C were prepared by dissolving 1 g of each ingredient in 100 mL of distilled water. The pH 5 lime solution was made by adding lime juice to distilled water at 85°C to the desired pH. In total, 900 µL of FRAP reagent was mixed with 120 µL of each sample solution. Then, the solution mixture was vortexed and incubated for 15 minutes. The absorbance of the solution was measured at a wavelength of 595 nm. Then, the absorbance data were processed using Microsoft Excel. The standard solution of FeSO₄.7H₂O was used as a standard curve by making a line equation for the absorbance value of the FeSO₄.7H₂O solution. The absorbance data of the sample were input into the equation of the line to obtain the FRAP value in µM/g.

2.9. Inhibition of the α -Glucosidase Activity

An inhibition test of α -glucosidase was performed using the method described by Azizi et al. [15]. Acarbose, an antidiabetic drug, and lime solution were used as the standards. The test consists of 50 mL of 0.1 M phosphate buffer (KH_2PO_4) pH 7, 25 mL of 0.5 mM p-nitrophenyl- α -D-glucopyranoside (PNP-G, as the substrate), 10 mL of the sample extract (1 g/100 mL) or standard (1 g/100 mL), and 0.2 U/mL α -glucosidase of 25 mL. The sample was also mixed and incubated at 37°C for approximately 30 min, and the reaction was stopped using 100 mL of 0.2 M Na_2CO_3 . The p-nitrophenol that was formed using a microplate reader was used to determine the enzyme activity, which was inhibited at a wavelength of 405 nm. Furthermore, the absorbance values were used to analyze the percentage inhibition of the enzyme.

$$\text{Percentage inhibition} = [(K - (S1 - S0)) / K] \times 100\%$$

where K, S1, and S0 are the absorbance of the control blank, a sample with enzyme and a sample without enzyme.

2.10. Analysis of the Consumer Acceptance

The level of consumer acceptance was analyzed using the hedonic test based on Suryono et al. [16]. This analysis used 80 untrained panelists. The steps were as follows: brew a sample of *S. polycystum* seaweed tea for each treatment by soaking 1 bag (1 gram) of the sample in 100 mL of boiling water for 6 minutes while raising and lowering 5 times and stirring 2-3 times; then, remove the teabag from the solution. In total, 25 mL of *S. polycystum* seaweed tea solution was put into the cup. Each sample was coded, and the panelists were asked to provide a sample response by directly tasting the tested sample and giving an assessment (score 1-5) of the sample, including the parameters of appearance, color, flavor, and taste [17]. The level of consumer acceptance of *S. polycystum* seaweed tea was obtained from the score given by the panelists with the following values: 5 = very like, 4 = like, 3 = somewhat like, 2 = dislike, 1 = very dislike.

2.11. Statistical Analysis

The analysis phase was conducted using 3 replications ($n = 3$). Data analysis was performed using SPSS (developed by SPSS Inc., USA). Data analysis included a normality test using the one-sample Kolmogorov-Smirnov test followed by a real difference test using multiple comparisons.

3. Results and Discussion

3.1. Water Content of *Sargassum Polycystum* Seaweed Tea

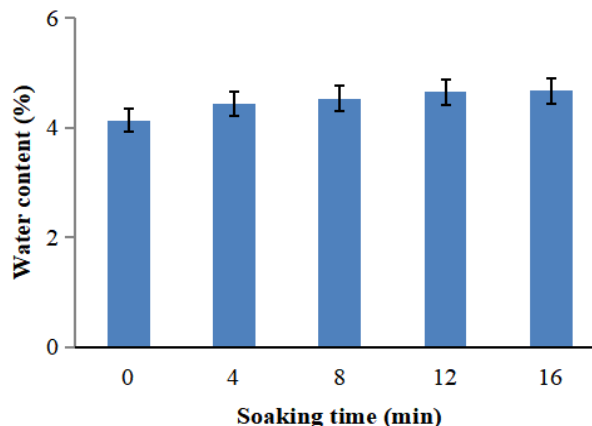


Fig. 1 Effect of the soaking time in lime solution at pH 5 on the water content of *S. polycystum* seaweed tea

The effect of the soaking time in lime solution at pH 5 on the water content of *S. polycystum* seaweed tea is shown in Fig. 1. The water content of *S. polycystum* seaweed as raw material was 92.96%. After the seaweed tea was made by soaking in the pH 5 lime solution for 0 ~ 16 minutes, the water content of *S. polycystum* seaweed tea was $4.14 \pm 0.11 \sim 4.68 \pm 0.12\%$. The analysis indicates that the soaking time in lime solution at pH 5 had no significant effect ($p > 0.05$) on the water content of seaweed tea. The water content of *S. polycystum* seaweed tea was consistent with the Indonesian National Standard (SNI) for dry packaged tea with a maximum water content of 8% [18]. Sinurat & Suryaningrum [10] reported that *Sargassum sp.* soaked in boiling water for 0 ~ 5 minutes were 2 ~ 6%, while Supirman et al. [9] reported that the water content of *S. filipendula* seaweed tea was 9.67%. The water content plays a major role in the deterioration and shelf life of food products. The moisture content of dry ingredients should be below 10% to prevent enzymatic processes and the growth of microbes [10]. The reason is that dry materials are generally stored for a long time; if an enzymatic process occurs, it will change its stored chemical composition, so the effects of the active compounds therein can also change.

3.2. Total Phenol Content

The total phenol content of *S. polycystum* seaweed tea is shown in Fig 2. *S. polycystum* seaweed tea that was soaked for 0, 4, 8, 12, and 16 minutes had total phenol contents of 15.26 ± 0 , 38.2547 ± 0.59 , 35.54 ± 0.58 , 55.75 ± 0.75 , and 74.53 ± 0.59 mg GAE/g, respectively. The soaking time had a significant effect ($p < 0.05$) on the increase in total phenol content of seaweed tea because soaking with an acid solution at 100°C can increase the total phenol content [19]. Soaking using hot water can minimize the damage to polyphenol compounds to increase the total phenol in ingredients [20].

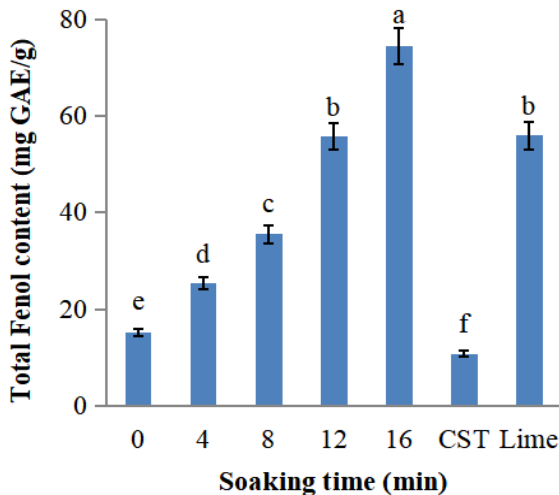


Fig. 2 Effect of the soaking time in lime solution at pH 5 on the total phenol content of *S. polycystum* seaweed tea. Note: CST = commercial seaweed tea

Supirman et al. [9] reported that the total phenol content of *S. filipendula* seaweed tea soaked in lime solution at pH 5 for 6 hours was 6.48 ± 0.03 mg GAE/g. Meanwhile, the total phenol content of seaweed tea from *Sargassum sp.* that was soaked in hot water for 0, 1, 3, and 5 minutes was $1.80 \sim 2.22$ mg GAE/g [10]. The *S. polycystum* seaweed tea had a higher total phenol content than *Sargassum sp.* and *S. filipendula* seaweed tea. This result can occur due to temperature and time when one soaks and roasts seaweed tea. According to Dewata et al. [21], high temperatures will increase the total phenol levels because high temperatures can increase the release of phenol compounds in the cell walls.

Additionally, a brewing time that is too long will decrease the total phenol levels because it can damage phenolic compounds in cell components [22]. Brewing tea for too short a time will make the compounds in the tea sample not properly dissolve, so the total phenol content decreases [23]. During brewing, phenol compounds are damaged at high temperatures, with an optimal temperature range of $0 \sim 90^\circ\text{C}$ [22].

3.3. DPPH Radical Scavenging Activity

The antioxidant activity of *S. polycystum* seaweed tea using the DPPH method is shown in Fig. 3. The DPPH inhibitory activities of *S. polycystum* seaweed tea that was soaked for 0, 4, 8, 12, and 16 minutes were 8.35 ± 0.71 , 18.88 ± 1.15 , 29.75 ± 0.87 , 40.99 ± 1.08 , and $52.70 \pm 0.86\%$, respectively. The data analysis results show that the soaking time significantly affected ($p < 0.05$) the DPPH inhibitory activity. The *S. polycystum* seaweed tea had higher antioxidant activity than *Sargassum sp.* seaweed tea ($4.00 \pm 1.34 \sim 18.00 \pm 2.01\%$) [10] but a lower antioxidant activity than *S. filipendula* seaweed tea ($64.13 \pm 0.40\%$) soaked in solution lime at pH 5 [9]. The reason is that the *S. filipendula* seaweed in the pH 5 lime solution was soaked for a longer time, i.e., for 6 hours.

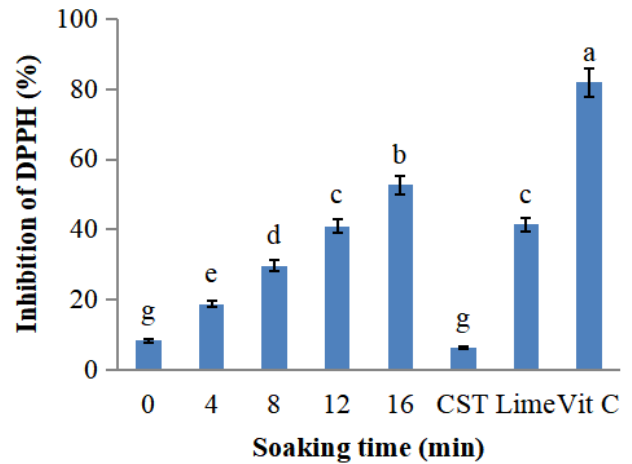


Fig. 3 Effect of the soaking time in lime solution at pH 5 on the DPPH inhibitory activity of *S. polycystum* seaweed tea. Note: CST = commercial seaweed tea

Fig. 3 shows that a longer soaking time in the pH 5 lime solution corresponds to a higher antioxidant activity of *S. polycystum* seaweed tea. In general, the heating treatment of ingredients can reduce the antioxidant activity, but heating by soaking can increase the antioxidant activity [24]. According to Suri et al. [11], a longer soaking time can reduce antioxidant activity. Sinurat & Suryaningrum [10] reported that the heating process with the right time by soaking could increase the antioxidant activity. Boonkorn [19] explained that soaking in hot water could release antioxidant components from within the cells to increase their antioxidant activity.

3.4. Ferric Reduction Antioxidant Power (FRAP)

The effect of the soaking time in lime solution at pH 5 on the FRAP value of *S. polycystum* seaweed tea is shown in Fig. 4. The FRAP values of *S. polycystum* seaweed tea that was soaked for 0, 4, 8, 12, and 16 minutes were 170.91 ± 0.91 , 159.70 ± 1.89 , 149.09 ± 0.91 , 138.79 ± 1.89 , and 123.94 ± 1.39 $\mu\text{M/g}$, respectively. The highest antioxidant activity was found in *S. polycystum* seaweed tea that was soaked for 16 minutes (123.94 ± 1.39 $\mu\text{M/g}$), while the lowest antioxidant activity was found in unsoaked seaweed tea (170.91 ± 0.91 $\mu\text{M/g}$). The antioxidant activity is higher if the FRAP value is lower because a lower sample concentration is required to achieve the absorbance produced by the FeSO_4 standard solution to convert Fe^{3+} to Fe^{2+} [25]. Based on FRAP value, *S. polycystum* seaweed tea's antioxidant activity was higher than *S. muticum* fucoidan [26].

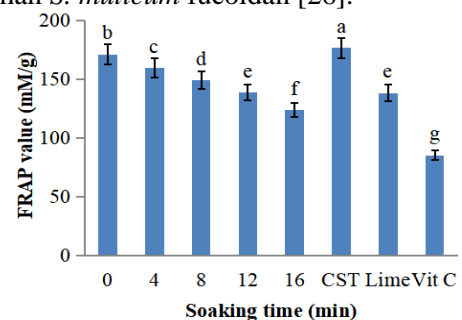


Fig. 4 Effect of the soaking time in lime solution at pH 5 on the FRAP value of *S. polycystum* seaweed tea. Note: CST = commercial seaweed tea

The antioxidant activity of *S. polycystum* seaweed tea increased with increasing soaking time. Thus, soaking in lime solution at pH 5 and 85°C can increase the antioxidant activity of *S. polycystum* seaweed tea. The increase in antioxidant activity during soaking was thought to occur because the compounds in seaweed, such as flavonoids, become more active [19]. In addition, the soaking process under acidic conditions can result in flavonoid compounds in the form of glycosides, which will be degraded into aglycones and sugars, which increase the antioxidant activity [27]. More OH groups donate H⁺ atoms with a higher polyphenol content of the flavonoid group, more OH groups donate H⁺ atoms, so the antioxidant activity increases [28].

3.5. Inhibitory Activity of α -Glucosidase

Data on the effect of the soaking time in the pH 5 lime solution on the α -glucosidase inhibitory activity of *S. polycystum* seaweed tea are shown in Fig. 5. *S. polycystum* seaweed tea-soaked for 0, 4, 8, 12, and 16 minutes had inhibitory activities of 11.47 ± 0.82 , 20.71 ± 1.02 , 32.34 ± 0.68 , 41.59 ± 0.60 , and $51.56 \pm 0.70\%$, respectively. The data show that a longer soaking time resulted in a higher α -glucosidase inhibitory activity. The highest α -glucosidase inhibitory activity was produced in the 16-minute soaking treatment ($51.56 \pm 0.70\%$), while the lowest was produced without soaking ($11.47 \pm 0.82\%$). This result can occur because one of the main ingredients of lime is flavonoids [29]. Flavonoids inhibit α -glucosidase in the breakdown of carbohydrates before being absorbed as monosaccharides [30].

Based on Fig. 5, the highest α -glucosidase inhibitory activity was obtained after soaking in the pH 5 lime solution for 16 minutes ($51.56 \pm 0.70\%$), while the lowest was obtained without soaking ($11.47 \pm 0.82\%$). The inhibitory activity of α -glucosidase by lime solution at pH 5 was quite high ($42.82 \pm 0.31\%$); thus, soaking seaweed in lime solution at pH 5 affects the α -glucosidase inhibitory activity of *S. polycystum* seaweed tea. Lime contains phenolic or polyphenol compounds such as flavonoids, glycosides, tannins, and saponins, which affect the inhibitory activity of α -glucosidase [31].

The *S. polycystum* seaweed tea had higher α -glucosidase inhibitory activity than commercial seaweed tea but lower than acarbose. Acarbose is often used for α -glucosidase inhibitors. Acarbose works competitively to inhibit the action of α -glucosidase within cells from reducing glucose absorption and postprandial hyperglycemia. In diabetic patients, acarbose can reduce postprandial hyperglycemia 30-50% and HbA1c 0.5-1% [32]. Acarbose can also competitively inhibit the hydrolysis of carbohydrates

with α -glucosidase to inhibit the hydrolysis process and prevents blood sugar levels from increasing. However, this drug was reported to cause gastrointestinal side effects such as diarrhea and flatulence [33].

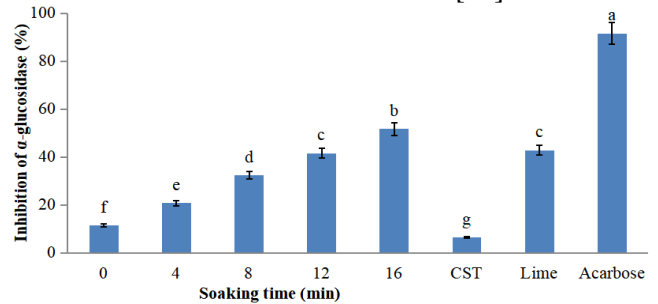


Fig. 5 Effect of immersion time in lime solution at pH 5 on the α -glucosidase inhibitory activity of *S. polycystum* seaweed tea. Note: CST = commercial seaweed tea

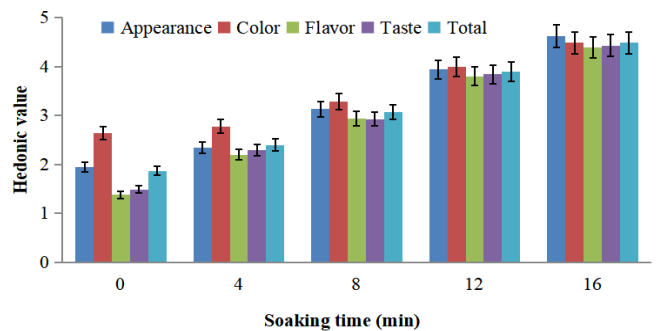


Fig. 6 Effect of the soaking time in lime solution at pH 5 on the hedonic value of *S. polycystum* seaweed tea

3.6. Level of Consumer Acceptance

Hedonic testing was conducted to determine the level of consumer acceptance of *S. polycystum* seaweed tea in terms of appearance, color, flavor, and taste. Appearance greatly affects the quality of a product [34]. Products with an attractive appearance can increase consumer attractiveness. The effect of soaking in lime solution at pH 5 on the color preference of *S. polycystum* seaweed tea is shown in Figure 6. The consumer preference value for the appearance of seaweed tea was 1.95-4.63. The sample without soaking had the lowest preference value (1.95), i.e., between very dislike and dislike, while the 16-minute soaking treatment had the most preferred appearance (4.63), i.e., between like and very like. Sinurat & Suryaningrum [10] reported that the level of preference for the appearance was 4.7 for *Sargassum sp.* that was soaked in boiling water for 1 minute, i.e., "like it until you truly like it". A good appearance will increase the value of other parameters such as color, flavor, and taste [35].

Color visually determines the level of acceptance of the product by consumers. The level of consumer acceptance of food products is influenced by color attributes [36]. The consumer acceptance of *S. polycystum* seaweed tea color preferences is presented in Fig. 6. The level of consumer acceptance of tea colors was 2.64-4.49. The sample immersed in the pH 5 lime solution for 16 minutes had the highest color value (4.49), i.e., between like and very like. The one

without soaking had the lowest color preference value (2.64), i.e., between dislike and somewhat like. The hedonic value of color in this study was higher than that of *S. filipendula* seaweed tea that was soaked in a pH 5 lime solution for 6 hours (4.10) [9] but still lower than that of *Sargassum sp.* seaweed tea that was immersed in boiling water for 1 minute (4.65) [10]. A possible reason is the different treatments and samples.

The consumer acceptance of the flavor value of *S. polycystum* seaweed tea is presented in Fig. 6. The level of consumer acceptance of flavor tea was 1.38-4.39. The sample soaked in the pH 5 lime solution for 16 minutes had the highest value of flavor preference (4.39), i.e., between like and very like. The results of this study were higher than the level of preference for the tea flavor of *S. filipendula* (2.50) soaked in pH 5 lime solution for 6 hours [9] and *Sargassum sp.* tea (3.71) soaked in boiling water for 1 minute [10].

The level of taste preference determines whether consumers can accept a product. Products that have many functions for health but unacceptable taste by consumers do not sell [17]. The effect of immersion on the taste preference level of *S. polycystum* seaweed tea is presented in Figure 6. The level of consumer preference for the taste of the tea was 1.49-4.43. The soaking time had a significant effect ($p < 0.05$) on the taste preference level of *S. polycystum* seaweed tea. A longer soaking time corresponds to a stronger lime taste in *S. polycystum* seaweed tea. In addition, according to Sinurat & Suryaningrum [10], the soaking process with hot water made the most dominant polymer compounds in seaweed change to oligomeric compounds, which the panelists preferred. The tannins caused the taste of seaweed tea in the brown seaweed *Sargassum sp.* Soaking in a hot lime solution can reduce the tartness of *S. polycystum* seaweed tea. The highest level of taste preference (4.43) was produced by soaking in a pH 5 lime solution for 16 minutes, i.e., between like and very like. The results of this study were higher than the taste preference level of *S. filipendula* seaweed tea (3.30) soaked in pH 5 lime solution for 6 hours [9]. Still, it was lower than the taste preferences of *Sargassum sp.* seaweed tea (4.65) that was immersed in boiling water for 1 minute [10].

Fig. 6 also shows the overall consumer preference of *S. polycystum* seaweed tea. The level of consumer acceptance of tea was 1.87-4.49. Based on statistical tests, the most preferred *S. polycystum* seaweed tea was seaweed tea soaked in the pH 5 lime solution for 16 minutes, and the least preferred tea by consumers was the one without soaking. The soaking time in lime solution at pH 5 affects the level of consumer acceptance. With a longer soaking time, the *S. polycystum* seaweed tea products were more preferred. *S. polycystum* seaweed tea's overall consumer preference was higher compared to *Sargassum sp.* seaweed tea [10].

S. polycystum seaweed can be made into functional beverage products in the form of tea through the following steps: 1) Seaweed *S. polycystum* was washed using clean water and then soaked pH 5 lime solution for 16 minutes. 2) After soaking, the seaweed was drained and dried for 24 hours in a baking sheet, which was placed on a terrace and not exposed to direct sunlight. 3) The seaweed was roasted in a frying pan for 15-20 minutes and cut with a knife into pieces 0.5 ± 0.1 cm in size. 4) Seaweed tea was placed in a sealed standing pouch and ready to use.

4. Conclusion

In conclusion, the soaking time in lime solution at pH 5 affects the antioxidant and antidiabetic activities and the consumer acceptance of *S. polycystum* seaweed tea. Soaking seaweed in lime solution pH 5 for 16 minutes was the best treatment for *S. polycystum* seaweed tea with the following characteristics: water content $4.68 \pm 0.12\%$; total phenol 74.53 ± 0.59 mg GAE/g; inhibition of DPPH $52.70 \pm 0.86\%$; FRAP value 123.94 ± 1.39 μ M/g; α -glucosidase inhibitory activity $51.56 \pm 0.70\%$; hedonic values for appearance, color, taste, flavor and overall: 4.63 ± 0.74 , 4.49 ± 0.75 , 4.39 ± 1.07 , 4.43 ± 1.03 , and 4.49 ± 0.11 , respectively. In producing *S. polycystum* seaweed tea, it is strongly influenced by the manufacturing process, one of which is the length of immersion in lime. Therefore, it is necessary to optimize the optimum time to get the best quality and the most preferred by consumers in the future.

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References

- [1] CAHYANINGRUM K., HUSNI A., and BUDHIYANTI S. A. Antioxidant activity of brown seaweed extract (*Sargassum polycystum*). *Agritech*, 2016, 36(2): 137-144. <https://doi.org/10.22146/agritech.12857>
- [2] BUDI P. H., THAIB E. A., and JULITA M. Use of *Sargassum polycystum* ethanol extract as antibacterial for increasing shelf life tilapia fillet (*Oreochromis niloticus*) stored in chilling temperature. *Institute of Physics Conference Series: Earth and Environmental*, 2019, 278: 012012. <https://doi.org/10.1088/1755-1315/278/1/012012>
- [3] BUWONO N. R., RISJANI Y., and ARSAD, S. Anti-inflammatory and analgesic activity from brown algae *Sargassum polycystum*. *Jurnal of Pharmaceutical Sciences*

- and Research, 2018, 10(8): 2092-2096. <https://www.jpsr.pharmainfo.in/Documents/Volumes/vol10Issue08/jpsr10081850.pdf>
- [4] LAILATSSIFA R., HUSNI A., and NUGROHO A. E. Anti-stress activity of *Sargassum polycystum* extracts using a cold restraint stress model. *Food Science and Biotechnology*, 2016, 25(2): 589-594. <https://doi.org/10.1007/s10068-016-0082-y>
- [5] PURUKAN J. A., KUSMARDI K., LAKSMITAWATI D. R., ABDILLAH S., and PRIOSOERYANTO B. P. Comparison of lipid profile in white rats were given crude fucoidan from brown seaweed (*Sargassum polycystum*) that induced high-fat diet. *Jurnal Ilmu Kefarmasian Indonesia*, 2019, 17(1): 46-55. <http://jifi.farmasi.univpancasila.ac.id/index.php/jifi/article/view/648>
- [6] MUBASHEERA M. G., KONERI R., and JHA D. K. A study on the type II antidiabetic activity of methanolic extract of marine algae, *Gracilaria edulis* and *Sargassum polycystum*. *International Journal Pharmacy Science*, 2017, 47(1): 154-159. <https://globalresearchonline.net/journalcontents/v47-1/28.pdf>
- [7] NINGRUM R. R., HARDOKO D. S., and SASMITO B. B. Effect of crude extract of brown algae fucoidan *Sargassum polycystum* as anticancer on Hela cell viability. *Jurnal Mahasiswa Teknologi Hasil Perikanan*, 2013, 1(1): 83-92. <http://thpi.studentjournal.ub.ac.id/index.php/thpi/article/view/8>
- [8] HOLINESTI R., & NURHAYANI. Effect of substitution of brown seaweed extract on quality of chicken sausage afkir. *Jurnal Pendidikan Tata Boga dan Teknologi*, 2020, 1(2): 54-59. <http://boga.ppj.unp.ac.id/index.php/jptb/article/view/31>
- [9] SUPIRMAN, HARTATI K., and KARTINI Z. The effect of differences in pH soaking lime (*Citrus auratifolia*) with sun drying on the chemical quality of brown algae tea (*Sargassum fillipendula*). *Jurnal Mahasiswa Teknologi Hasil Perikanan*, 2012, 1(1): 46-52. <http://thpi.studentjournal.ub.ac.id/index.php/thpi/article/view/6>
- [10] SINURAT E., & SURYANINGRUM T. D. The effect of blanching time on antioxidant activity and sensory characteristic of brown seaweed *Sargassum sp.* tea. *Jurnal Pengolahan Hasil Perikanan Indonesia*, 2019, 22(3): 581-588. <https://doi.org/10.17844/jphpi.v22i3.29228>
- [11] SURI S., KUMAR V., TANWAR B., GOYAL B., & GAT Y. Impact of soaking and germination time on nutritional composition and antioxidant activity of *Nigella sativa*. *Current Research in Nutrition and Food Science*, 2019, 07(1): 142-149. <http://dx.doi.org/10.12944/CRNFSJ.7.1.14>
- [12] SENG J. L., WAN A. W. M., and MOHAMAD Y. M. Seaweed tea: fucoidan-rich functional food product development from Malaysian brown seaweed, *Sargassum binderi*. *Sains Malaysiana*, 2017, 46(9): 1573-1579. <http://dx.doi.org/10.17576/jsm-2017-4609-28>
- [13] MUTHIA R., SAPUTRI R., and VERAWATI S. A. Antioxidant activity test of ethanol extract of mundar fruit peel (*Garcinia forbesii* King.) using the DPPH method (2,2-diphenyl-1-picrylhydrazil). *Jurnal Pharmascience*, 2019, 6(1): 78-82. <https://ppjp.ulm.ac.id/journal/index.php/pharmascience/article/view/6079>
- [14] SUHAILA K., HUSNI A., and SINURAT E. Characteristics and antioxidant activity of fucoidan from the brown seaweed *Sargassum hystrix*. *Aquaculture, Aquarium, Conservation & Legislation Bioflux*, 2019, 12(6): 2319-2329. <http://www.bioflux.com.ro/docs/2019.2319-2329.pdf>
- [15] AZIZI W. A., EKANTARI N., and HUSNI A. Inhibitor activity of *Sargassum hystrix* extract and its methanolic fractions on inhibiting α -glucosidase activity. *Indonesian Journal of Pharmacy*, 2019, 30(1): 35-42. <https://doi.org/10.14499/indonesianjpharm30iss1pp35>
- [16] SURYONO C., NINGRUM L., and DEWI T. Descriptive and organoleptic test of 5 packages of Thousand Islands products. *Jurnal Pariwisata*, 2018, 5(2): 96-107. <https://ejournal.bsi.ac.id/ejournal/index.php/jp/article/view/3526>
- [17] PUTRI R. M. S., & MARDESCI H. Hedonic test of scallop shell biscuits (*Placuna placenta*) from Indragiri Hilir waters. *Jurnal Teknologi Pertanian*, 2018, 7(2): 19-29. <https://ejournal.unisi.ac.id/%20index.php/jtp/article/view/279>
- [18] NATIONAL STANDARDIZATION AGENCY. *SNI 3836: 2013. Packaged dried tea*. National Standardization Agency of Indonesia, Jakarta, 2013. <https://docplayer.info/32051689-Teh-kering-dalam-kemasan.html>
- [19] BOONKORN P. Impact of hot water soaking on antioxidant enzyme activities and some qualities of storage tomato fruits. *International Food Research Journal*, 2016, 23(3): 934-938. [http://www.ifrj.upm.edu.my/23\(03\)2016/\(4\).pdf](http://www.ifrj.upm.edu.my/23(03)2016/(4).pdf)
- [20] NURHAYATI N., MARSENO D. W., SETYABUDI F. M. C. S., and SUPRIYANTO S. Effect of steam blanching on polyphenol oxidase activities, total polyphenols, and antioxidant activities of cocoa beans. *Jurnal Aplikasi Teknologi Pangan*, 2018, 7(3): 95-103. <https://doi.org/10.17728/jatp.2314>
- [21] DEWATA I. P., WIPRADNYADEWI P. A. S., and WIDARTA I. W. R. Effect of temperature and brewing time on antioxidant activity and avocado leaf herbal tea (*Persea americana* Mill). *Jurnal Ilmu dan Teknologi Pangan*, 2017, 6(2): 30-39. <https://ojs.unud.ac.id/index.php/itepa/article/view/36700>
- [22] PEREZ-BURILLO S., GIMÉNEZ R., RUFÍAN-HENARES J. A., and PASTORIZA S. Effect of brewing time and temperature on antioxidant capacity and phenols of white tea: Relationship with sensory properties. *Food Chemistry*, 2018, 248: 111-118. <https://doi.org/10.1016/j.foodchem.2017.12.056>
- [23] TAMBUN R., LIMBONG H. P., PINEM C., and MANURUNG E. Effect of particle size, time and temperature on phenol extraction from red galangal. *Jurnal Teknik Kimia*, 2016, 5(4): 53-56. <https://doi.org/10.32734/jtk.v5i4.1555>
- [24] GARRETSON L., TYL C., and MARTI A. Effect of processing on antioxidant activity, total phenols, and total flavonoids of pigmented heirloom beans. *Journal of Food Quality*, 2018, 2018: 7836745. <https://doi.org/10.1155/2018/7836745>
- [25] AB-RAHIM N., ZAKARIA N., DZULKARNAIN S. M. H., AZAHAR N. M. Z. M., and ABDULLA M. A. Antioxidant activity of *Alstonia angustifolia* ethanolic leaf extract. *American Institute of Physics Conference Proceedings*, 2017, 1891: 020012. <https://doi.org/10.1063/1.5005345>

- [26] KURNIALAHI I. D., HUSNI A., SINURAT E., and ISNANSETYO A. Antioxidant activity of tropical seaweed *Sargassum muticum* fucoidan. *Aquaculture, Aquarium, Conservation & Legislation Bioflux*, 2020, 13(1): 230-240. <http://bioflux.com.ro/docs/2020.230-240.pdf>
- [27] PERDANA A. G., PRATIWI E., and KUNARTO B. Effect of blanching time on antioxidant activity and phenolic compound levels of red ginger extract (*Zingiber officinale* var. *rebrum*). *Jurnal Mahasiswa, Food Technology and Agricultural Products*, 2018. <https://repository.usm.ac.id/detail-jurnalmahasiswa-131.html>
- [28] CHEN J., YANG J., MA L., LI J., SHAHZAD N., and KIM C. K. Structure-antioxidant activity relationship of methoxy, phenolic hydroxyl, and carboxylic acid groups of phenolic acids. *Scientific Reports*, 2020, 10: 2611. <https://doi.org/10.1038/s41598-020-59451-z>
- [29] PRASTIWI S. S., & FERDIANSYAH F. Content and pharmacological activity of lime (*Citrus aurantifolia* S.). *Farmaka*, 2017, 15(2): 1-8. <https://doi.org/10.24198/jf.v15i2.12964>
- [30] PERMATASARI R., ANDRIANE Y., GARNA H., HARIBUDIMAN O., and EKOWATI R. A. R. The effect of water fraction of lemon (*Citrus limon*) on blood glucose levels of old mice that are given a high-fat diet. *Jurnal Integrasi Kesehatan & Sains*, 2019, 1(1): 54-58. <https://ejournal.unisba.ac.id/index.php/jiks/article/view/4322>
- [31] SERANG Y., & BANI F. Test of anti-hyperglycemic activity, and oxidative stress inhibition of lime (*Citrus aurantifolia*) leaf ethanol extract in alloxan-induced diabetic rats. *Biomedika*, 2017, 10(1): 85-92. <http://dx.doi.org/10.31001/biomedika.v10i1.232>
- [32] DINICOLANTONIO J. J., BHUTANI J., and O'KEEFE J. H. Acarbose: safe and effective for lowering postprandial hyperglycaemia and improving cardiovascular outcomes. *Open Heart*, 2015, 2: e000327. <http://dx.doi.org/10.1136/openhrt-2015-000327>
- [33] SUN W., ZENG C., LIAO L., CHEN J., and WANG Y. Comparison of acarbose and metformin therapy in newly diagnosed type 2 diabetic patients with overweight and/or obesity. *Current Medical Research and Opinion*, 2016, 32(8): 1389-1396. <https://doi.org/10.1080/03007995.2016.1176013>
- [34] SCHNURR B., BRUNNER-SPERDIN A., and STOKBURGER-SAUER N. E. The effect of context attractiveness on product attractiveness and product quality: the moderating role of product familiarity. *Marketing Letters*, 2017, 28: 241-253. <https://doi.org/10.1007/s11002-016-9404-3>
- [35] LUTHFIYANA N., NURJANAH, NURILMALA M., ANWAR E., and HIDAYAT T. Porridge ratio of *Eucheuma cottonii* and *Sargassum* sp. as a sunscreen cream formula. *Jurnal Pengolahan Hasil Perikanan Indonesia*, 2016, 19(3): 183-195. <https://doi.org/10.17844/jphpi.2016.19.3.183>
- [36] APANDI I., RESTUHADI F., and YUSMARINI. Analysis of consumer's preference mapping on sensory attributes of soygurt products among students of the Faculty of Agriculture, University of Riau. *Jurnal Online Mahasiswa*, 2016, 3(1): 1-16. <https://jom.unri.ac.id/index.php/JOMFAPERTA/article/view/9599/9263>
- [1] CAHYANINGRUM K., HUSNI A., and BUDHIYANTI S. A. A. 棕色海藻提取物 (马尾藻) 的抗氧化活性. *农业科技*, 2016, 36(2): 137-144. <https://doi.org/10.22146/agritech.12857>
- [2] BUDI P. H., THAIB E. A., and JULITA M. 使用马尾藻多囊乙醇提取物作为抗菌剂来延长冷藏温度下储存的罗非鱼片 (尼罗罗非鱼) 的保质期。物理研究所会议系列: 地球与环境, 2019, 278: 012012. <https://doi.org/10.1088/1755-1315/278/1/012012>
- [3] BUWONO N. R., RISJANI Y., and ARSAD, S. 褐藻马尾藻多囊的抗炎和镇痛活性. *药学研究杂志*, 2018, 10(8): 2092-2096. <https://www.jpsr.pharmainfo.in/Documents/Volumes/vol10Issue08/jpsr10081850.pdf>
- [4] LAILATSSIFA R., HUSNI A., and NUGROHO A. E. 使用冷束缚应激模型的马尾藻多囊提取物的抗应激活性。食品科学与生物技术, 2016, 25(2): 589-594. <https://doi.org/10.1007/s10068-016-0082-y>
- [5] PURUKAN J. A., KUSMARDI K., LAKSMITAWATI D. R., ABDILLAH S., and PRIOSOERYANTO B. P. 白鼠的脂质分布比较从诱导高脂肪饮食的棕色海藻 (多囊马尾藻) 中得到粗褐藻糖胶. *印度尼西亚药学杂志*, 2019, 17(1): 46-55. <http://jifi.farmasi.univpancasila.ac.id/index.php/jifi/article/view/648>
- [6] MUBASHEERA M. G., KONERI R., and JHA D. K. 海藻、江蓠和马尾藻多囊藻甲醇提取物的II型抗糖尿病活性研究. *国际期刊药科学*, 2017, 47(1): 154-159. <https://globalresearchonline.net/journalcontents/v47-1/28.pdf>
- [7] NINGRUM R. R., HARDOKO D. S., and SASMITO B. B. 褐藻褐藻多囊粗提物抗癌作用对海拉细胞活力的影响. *水产品技术学生杂志*, 2013, 1(1): 83-92. <http://thpi.studentjournal.ub.ac.id/index.php/thpi/article/view/8>
- [8] HOLINESTI R., and NURHAYANI. 替代褐海藻提取物对阿夫基尔鸡肉香肠品质的影响. *餐饮科技教育杂志*, 2020, 1(2): 54-59. <http://boga.ppi.unp.ac.id/index.php/jptb/article/view/31>
- [9] SUPIRMAN, HARTATI K., and KARTINI Z. 不同pH浸泡石灰 (柑橘) 与晒干对褐藻茶 (马尾藻) 化学品质的影响. *水产品技术学生杂志*, 2012, 1(1): 46-52. <http://thpi.studentjournal.ub.ac.id/index.php/thpi/article/view/6>
- [10] SINURAT E., and SURYANINGRUM T. D. 热烫时间对褐海藻马尾藻属抗氧化活性和感官特性的影响. *茶*. *印度尼西亚水产品加工杂志*, 2019, 22(3): 581-588. <https://doi.org/10.17844/jphpi.v22i3.29228>
- [11] SURI S., KUMAR V., TANWAR B., GOYAL B., and GAT Y. 浸泡和发芽时间对黑种草营养成分和抗氧化活性的影响。营养与食品科学的最新研究, 2019, 07(1): 142-149. <http://dx.doi.org/10.12944/CRNFSJ.7.1.14>
- [12] SENG J. L., WAN A. W. M., and MOHAMAD Y. M. 海藻茶: 从马来西亚褐海藻马尾藻中开发出富含褐藻糖胶的功能性食品. *马来西亚圣贤*, 2017, 46(9): 1573-1579. <http://dx.doi.org/10.17576/jsm-2017-4609-28>

- [13] MUTHIA R., SAPUTRI R., 和 VERAWATI S. A. 用2,2-二苯基-1-苦基肼法测定芒达果皮(藤黄王)乙醇提取物的抗氧化活性. 药学杂志, 2019, 6(1): 78-82. <https://ppjp.ulm.ac.id/journal/index.php/pharmascience/article/view/6079>
- [14] SUHAILA K., HUSNI A., 和 SINURAT E. 褐海藻马尾藻岩藻依聚糖的特性及抗氧化活性. 水产养殖、水族馆、保护和立法生物通量, 2019, 12(6): 2319-2329. <http://www.bioflux.com.ro/docs/2019.2319-2329.pdf>
- [15] AZIZI W. A., EKANTARI N., 和 HUSNI A. 马尾藻提取物及其甲醇组分对 α -葡萄糖苷酶活性的抑制活性. 印度尼西亚药理学杂志, 2019, 30(1): 35-42. <https://doi.org/10.14499/indonesianjpharm30iss1pp35>
- [16] SURYONO C., NINGRUM L., 和 DEWI T. 5包千岛产品的描述和感官测试. 旅游杂志, 2018, 5(2): 96-107. <https://ejournal.bsi.ac.id/ejournal/index.php/jp/article/view/3526>
- [17] PUTRI R. M. S., 和 MARDESCI H. 来自因陀罗希利尔水域的扇贝壳饼干(普拉库纳胎盘)的快感测试. 农业技术杂志, 2018, 7(2): 19-29. <https://ejournal.unisi.ac.id/%20index.php/jtp/article/view/279>
- [18] 国家标准化机构. 印度尼西亚国家标准3836: 2013. 包装干茶. 印度尼西亚国家标准化机构, 雅加达, 2013. <https://docplayer.info/32051689-Teh-kering-dalam-kemasan.html>
- [19] BOONKORN P. 热水浸泡对抗氧化酶活性和贮藏番茄果实某些品质的影响. 国际食品研究杂志, 2016, 23(3): 934-938. [http://www.ifrj.upm.edu.my/23\(03\)2016/\(4\).pdf](http://www.ifrj.upm.edu.my/23(03)2016/(4).pdf)
- [20] NURHAYATI N., MARSENO D. W., SETYABUDI F. M. C. S., 和 SUPRIYANTO S. 蒸汽热烫对可可豆多酚氧化酶活性、总多酚和抗氧化活性的影响. 工业应用技术杂志, 2018, 7(3): 95-103. <https://doi.org/10.17728/jatp.2314>
- [21] DEWATA I. P., WIPRADNYADEWI P. A. S., 和 WIDARTA I. W. R. 温度和冲泡时间对抗氧化活性和鳄梨叶凉茶的影响(美洲磨坊). 食品科学与技术杂志, 2017, 6(2): 30-39. <https://ojs.unud.ac.id/index.php/itepa/article/view/36700>
- [22] PEREZ-BURILLO S., GIMÉNEZ R., RUFÍAN-HENARES J. A., 和 PASTORIZA S. 冲泡时间和温度对白茶抗氧化能力和酚类物质的影响: 与感官特性的关系. 食品化学, 2018, 248: 111-118. <https://doi.org/10.1016/j.foodchem.2017.12.056>
- [23] TAMBUN R., LIMBONG H. P., PINEM C., 和 MANURUNG E. 粒径、时间和温度对红高良姜酚提取的影响. 化学工程学报, 2016, 5(4): 53-56. <https://doi.org/10.32734/jtk.v5i4.1555>
- [24] GARRETSON L., TYL C., 和 MARTI A. 加工对有色传家宝豆的抗氧化活性、总酚和总黄酮的影响. 食品质量杂志, 2018, 2018: 7836745. <https://doi.org/10.1155/2018/7836745>
- [25] AB-RAHIM N., ZAKARIA N., DZULKARNAIN S. M. H., AZAHAR N. M. Z. M., 和 ABDULLA M. A. 小叶阿尔斯通乙醇叶提取物的抗氧化活性. 美国物理学会会议论文集, 2017, 1891: 020012. <https://doi.org/10.1063/1.5005345>
- [26] KURNIALAHI I. D., HUSNI A., SINURAT E., 和 ISNANSETYO A. 热带海藻马尾藻岩藻多糖的抗氧化活性. 水产养殖、水族馆、保护和立法生物通量, 2020, 13(1): 230-240. <http://bioflux.com.ro/docs/2020.230-240.pdf>
- [27] PERDANA A. G., PRATIWI E., 和 KUNARTO B. 热烫时间对红姜提取物(生姜变种. 雷布鲁姆)的抗氧化活性和酚类化合物水平的影响. 学生日记, 食品技术和农产品, 2018. <https://repository.usm.ac.id/detail-jurnalmahasiswa-131.html>
- [28] CHEN J., YANG J., MA L., LI J., SHAHZAD N., 和 KIM C. K. 酚酸的甲氧基、酚羟基和羧酸基团的结构-抗氧化活性关系. 科学报告, 2020, 10: 2611. <https://doi.org/10.1038/s41598-020-59451-z>
- [29] PRASTIWI S. S., 和 FERDIANSYAH F. 石灰(枳实)的含量和药理活性. 法马卡, 2017, 15(2): 1-8. <https://doi.org/10.24198/jf.v15i2.12964>
- [30] PERMATASARI R., ANDRIANE Y., GARNA H., HARIBUDIMAN O., 和 EKOWATI R. A. R. 柠檬(柑橘柠檬)的水分对高脂饮食老年小鼠血糖水平的影响. 健康与科学整合杂志, 2019, 1(1): 54-58. <https://ejournal.unisba.ac.id/index.php/jiks/article/view/4322>
- [31] SERANG Y., 和 BANI F. 在四氧嘧啶诱导的糖尿病大鼠中测试抗高血糖活性和酸橙(枳实)叶乙醇提取物的氧化应激抑制作用. 生物医学, 2017, 10(1): 85-92. <http://dx.doi.org/10.31001/biomedika.v10i1.232>
- [32] DINICOLANTONIO J. J., BHUTANI J., 和 O'KEEFE J. H. 阿卡波糖: 安全有效降低餐后高血糖和改善心血管结局. 放开心灵, 2015, 2: e000327. <http://dx.doi.org/10.1136/openhrt-2015-000327>
- [33] SUN W., ZENG C., LIAO L., CHEN J., 和 WANG Y. 阿卡波糖和二甲双胍治疗初诊超重和/或肥胖的2型糖尿病患者的比较. 当前的医学研究和观点, 2016, 32(8): 1389-1396. <https://doi.org/10.1080/03007995.2016.1176013>
- [34] SCHNURR B., BRUNNER-SPERDIN A., 和 STOKBURGER-SAUER N. E. 情境吸引力对产品吸引力和产品质量的影响: 产品熟悉度的调节作用. 营销信函, 2017, 28: 241-253. <https://doi.org/10.1007/s11002-016-9404-3>
- [35] LUTHFIYANA N., NURJANAH, NURILMALA M., ANWAR E., 和 HIDAYAT T. 紫檀和马尾藻属的粥比例. 作为防晒霜配方. 印度尼西亚水产品加工杂志, 2016, 19(3): 183-195. <https://doi.org/10.17844/jphpi.2016.19.3.183>
- [36] APANDI I., RESTUHADI F., 和 YUSMARINI. 廖内大学农学院学生消费者对豆浆产品感官属性偏好映射分析. 学生在线杂志, 2016, 3(1): 1-16. <https://jom.unri.ac.id/index.php/JOMFAPERTA/article/view/9599/9263>