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## Pineapple Rooting in Vitro from Sipahutar, North Sumatra, Indonesia, with Addition of Sucrose and IBA

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**Abstract:** The advantages of Sipahutar Pineapple are that it tastes sweeter, has a low content of water, softer texture, yellowish color, and is suitable with people preference, but up until now, the production is very limited. In vitro rooting was carried out to determine the effect of sucrose addition and Plant Growth Regulator (PGR) Indole Butyric Acid (IBA) on base media MS + 1 ppm BAP of Sipahutar pineapple (*Ananas comosus* L.) plantlet roots with in vitro. This research was conducted using a completely randomized factorial design. Factor 1: Sucrose in a row, 0, 10, 20, 30 gram L<sup>-1</sup>. Factor 2: PGR IBA with the dose is 0, 1.5, and 3 ppm. The interaction of sucrose and IBA on MS + 1 ppm BAP basic medium significantly affected the number of roots, the number of leaves, leaf length, leaf width and had no significant effect on other parameters.

**Keywords:** rooting, indole butyric acid, sucrose, *Ananas comosus* L., Sipahutar pineapple.

## 来自印度尼西亚北苏门答腊西帕胡塔的菠萝在体外生根 · 并添加了蔗糖和国际律师协会

**摘要:** 西帕胡塔菠萝的优点是味道更甜, 水分含量低, 质地更柔软, 颜色偏黄, 适合人们的喜好, 但到目前为止, 产量非常有限。进行体外生根以确定添加蔗糖和植物生长调节剂(遗传资源)吲哚丁酸(国际律师协会)对西帕胡塔菠萝(凤梨升)小植物根的基础培养基多发性硬化症+1百万分之一BAP的影响。这项研究是使用完全随机的因子设计进行的。因子1:连续蔗糖,0、10、20、30克L<sup>-1</sup>。因素2:遗传资源国际律师协会的剂量为0、1.5和3百万分之一。蔗糖和国际律师协会在多发性硬化症+1百万分之一BAP基础培养基上的相互作用显著影响根数、叶数、叶长、叶宽,对其他参数无显著影响。

**关键词:** 生根, 吲哚丁酸, 蔗糖, 凤梨, 西帕胡塔菠萝。

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## 1. Introduction

Sipahutar Pineapple is a horticultural commodity that has long been cultivated. It has prospects and potential to continue to be developed in Sipahutar District. Pineapple has the widest harvest of 1,819 ha, with a production of 32,532.82 tons [1]. Pineapple is a vegetatively propagated plant; now, this commodity is very important in tropical countries [2]. Pineapple contains sugar, acids and is rich in vitamins [3]. Pineapple from Sipahutar is a source of income for the people of North Tapanuli [4].

Generally, farmers plant pineapple from Sipahutar by using seedlings derived from tillers stem and crown [5]. By utilizing seedlings derived from crowns and tillers stem, which are relatively limited in number to plant huge land. The obstacle faced by pineapple farmers in Sipahutar is the difficulty in getting the pineapple seeds [6]. One of the alternatives that can be used to solve the needs of many seedlings is using in vitro culture techniques. In vitro culture has several advantages, including preparing large-scale seedlings in rare, endemic, and endangered plants [7]. The seedlings produced are more tolerant of pests and disease (aseptic propagation) and have a good uniformity of plant material.

The success of in vitro culture is strongly affected by growth regulators (PGR) [8], [9], [10]. MS media is often used in plant tissue culture applications both in embryogenic and organogenesis [11]. According to [12], one of the media components that determines the success of tissue culture is the type and concentration of PGR used. The type and concentration of PGR depend on the purpose and stage of the culture [13]. Auxins are phytohormones that are known to play an important role in plant growth and root development. However, the effect of auxin can vary in various plant species [14]. Indole Butyric Acid (IBA) is a growth regulator which is more often used than other types of auxin because IBA is a synthetic auxin, which is superior and effective in stimulating rooting activities; also, IBA has more stable chemical content and is higher to stimulate root formation [15].

In addition to growth regulators, the carbon source is also an important factor in determining the success of tissue culture. The carbon source serves as a source of energy that is needed by cells to be able to grow. This research will show the further influence of sucrose as an energy source. As a result of sucrose hydrolysis, glucose and fructose can stimulate the growth of some plants [16]. Based on research that has been recorded, the sucrose and IBA test for plant growth on a pineapple is still very limited, while the combination of sucrose + IBA has never been done on Sipahutar pineapple. Therefore, this research is important and has a high degree of novelty for farmers and science in the future.

## 2. Materials and Methods

This research was carried out in the YAHDI tissue culture laboratory, Perum Pelabuhan Jl. Lambung No. 18 Tanah 600 Medan Marelan Indonesia in March – September 2018. The tools used are Laminar Air Flow Cabinet (LAFC), autoclaves, analytical balance, refrigerators, and standard tissue culture tools. The material used was Pineapple Explants from Sipahutar, which was developed at the Yahdi Tissue Culture Laboratory in Medan.

This research used a factorial Completely Randomized Factorial Design experimental method, with 12 treatments. With the basic media Murashige and Skoog (MS) media + Plant Growth Regulator (PGR) of 1 ppm Benzyl Amino Purine (BAP), the factors in this study are: Factor I: Sucrose, consists of 4 levels of treatment:  $S_0 = 0$  ppm (control),  $S_{10} = 10$  gram  $L^{-1}$ ,  $S_{20} = 20$  gram  $L^{-1}$ ,  $S_{30} = 30$  gram  $L^{-1}$ . Factor II: IBA (PGR) consisted of 3 levels of treatment:  $I_0 = 0$  ppm (control),  $I_{1.5} = 1.5$  ppm,  $I_3 = 3$  ppm. Each treatment was repeated three times, resulting in 36 bottles in this research.

All tools used in this study were sterilized in two stages. The first stage was washed thoroughly and followed up by using an autoclave at  $121^\circ C$  with a pressure of 17.5 psi for 1 hour. All ingredients are used according to a predetermined dose. MS + Sucrose media (0; 10; 20; 30) grams  $L^{-1}$  combined with IBA (0; 1.5; 3 ppm) were dissolved in 1 liter of sterile aquades, then measured media pH (5.8) and added to (7 gram  $L^{-1}$ ).

Explant planting used is in vitro Sipahutar Pineapple shoots. Planting is carried out in a LAFC with a sterile planting tool. The in vitro shoots were planted into the rooting medium according to treatment. After all the explants were planted, Sipahutar pineapple explants were transferred to the culture room on the plant rack.

Maintenance was done by placing an explant-filled bottle on a research rack at a temperature range of  $22^\circ C$ . For light treatment, the application was made with a 3000-3200 lux fluorescent lamp in a 16-hour photoperiod.

After planting, the next step was observing plant growth, starting from one week old until 12 Weeks After Planting (WAP). The parameters observed in this research were:

- (1) Time of root emergence is characterized by swelling of the bulb buds. White bumps appear, followed by the release of fine roots. Observations are carried out every week to see when the roots emerged;
- (2) The number of roots. Counting of the number of roots starts from week 1 to 12 weeks of observation, by counting the number of roots each week;
- (3) Time of emergence of leaves. The time of the emergence of the leaves is calculated since the new leaves are formed. Observations are carried out every week to see when exactly the leaves appear;

(4) The number of leaves. Total leaf counted every week, from the first week to 12 weeks after treatment;

(5) The number of tillers is calculated every week by counting the total tillers every week, from the first week to 12 weeks after treatment;

(6) Time of emergence of tillers. Observations were every week to see how many tillers appear in a specific week;

Other observation parameters:

(7) Root length. This parameter was measured from the root base to the tip of the root;

(8) Leaf width. This parameter was measured by measuring the width of the leaf in the center of the leaf;

(9) Leaf length. This parameter was measured from the leaf base to the tip of the leaf;

(10) Height of tillers. This parameter was observed by measuring the height of new shoots formed;

(11) Plant height. This parameter was measured from the base of the stem to the top (leaf tip);

(12) Overall plant height. This parameter measured from leaf tips to root tips.

Parameters 7–12 were observed on 12 Weeks after Planting (12 WAP). After all the data was obtained, the data is processed by analysis techniques using a factorial completely randomized design model as follows:

$$Y_{ijk} = \mu + \alpha_i + \beta_j + (\beta_j)_{ij} + \epsilon_{ijk}$$

where:  $Y_{ijk}$  - the results of observations on the replication of the k-test which received the treatment of myo-inositol on the i-level and IBA treatment on the j-level;  $\mu$  - middle value;  $\alpha_i$  - concentration effect of myo-inositol on the i-level;  $\beta_j$  - concentration effect of IBA on the j level;  $(\beta_j)_{ij}$  - interaction effect of myo-inositol on the i-level with IBA on the j-level;  $\epsilon_{ijk}$  - error effect of myo-inositol on the i-level with IBA on the j-level at the k-test.

### 3. Results

#### 3.1. Number of Roots

It appears that as a single factor, sucrose or PGR IBA do not affect increasing the number of roots (Table 1), but the interaction of both has a significant and very significant effect on increasing the number of roots, especially starting from Week 3 until the end of observation (Week 12). The highest number of roots ( $2 \pm 0.49$ ) strands resulted from the treatment of medium base MS + 1 ppm BAP with the addition of 20 grams of sucrose  $L^{-1}$  and without the addition of IBA ( $S_{20} I_0$ ), followed by 20 grams of sucrose  $L^{-1}$  treatment and addition of 3 ppm IBA ( $S_{20} I_3$ ) produced  $1.96 \pm 0.67$  roots. The lowest root number was  $0.71 \pm 0.00$ , resulting from the treatment without sucrose and 1.5 ppm IBA ( $S_0 I_{1.5}$ ).

#### 3.2. Number of Leaves

Sucrose, PGR IBA, and their interactions had not affected the initial stage of the plant. In subsequent observations, sucrose has a significant and very significant effect, especially starting at Week 5 until the end of the observation (Table 1). PGR IBA from the beginning to the end of the observation did not have a significant effect. The interaction between the two (sucrose and PGR IBA) formed a significant effect from week 8 to the 12<sup>th</sup> week of observation. The highest number of leaves was produced from MS + 1 ppm BAP base medium with the addition of 20 grams of sucrose  $L^{-1}$  and 3 ppm IBA ( $S_{20} I_3$ ) with an average number of leaves  $31.33 \pm 15.89$ . Without the addition of sucrose but given 3 ppm IBA ( $S_0 I_3$ ) produced the lowest number of leaves ( $2.67 \pm 0.58$ ) (Table 1).

#### 3.3. Number of Tillers Buds

Sucrose generally has a significant and very significant effect, especially starting in Week 5 until the end of the observation. In contrast, PGR IBA and the interaction of both have no significant effect on the number of tiller buds (Table 1). During Weeks 3-12, the highest number of shoots resulted from 20 grams  $L^{-1}$  sucrose treatment and 3 ppm PGR IBA ( $S_{20} I_3$ ), with an average number of shoots of 2.67. Treatment of 0 gram  $L^{-1}$  sucrose and 3 ppm IBA ( $S_0 I_3$ ), 10-gram  $L^{-1}$  sucrose and 3 ppm IBA ( $S_{10} I_3$ ), and 30-gram  $L^{-1}$  sucrose and 1.5 ppm IBA ( $S_{30} I_{1.5}$ ) did not produce shoots shown in Fig. 1.

#### 3.4. Leaf Length

Table 2 shows that the rooting induction treatment with MS + 1 ppm BAP base medium with 12 combinations of treatments above gives different responses for the observed parameters, namely number of roots, number of leaves, leaf length, and leaf width. The highest leaf length was produced from the treatment without sucrose and 3 ppm IBA ( $S_0 I_3$ ) with an average leaf length of  $2.35 \pm 0.04$ , then decreased with a leaf length of  $2.28 \pm 0.10$ , resulting from 20 grams  $L^{-1}$  sucrose treatment of and PGR 3 ppm IBA ( $S_{20} I_3$ ). The shortest leaves averaged  $1.15 \pm 0.13$ , resulting from the treatment of 20 grams sucrose  $L^{-1}$  and 1.5 ppm IBA ( $S_{20} I_{1.5}$ ). Table 2 shows that sucrose has no effect, PGR IBA has a significant effect, and the interaction between sucrose and PGR IBA has a very significant effect on leaf length.

#### 3.5. Leaf Width

In general, leaf width is near evenly distributed. The highest leaf width resulted from 10 grams sucrose  $L^{-1}$  treatment and 1.5 ppm IBA ( $S_{10} I_{1.5}$ ) with leaf width of  $1.12 \pm 0.11$  (Table 2), while the lowest leaf width resulted from administering the same dose of 10 grams sucrose  $L^{-1}$  and 3 ppm IBA ( $S_{10} I_3$ ) with leaf width  $0.97 \pm 0.03$  (Fig. 2). The widest leaf is not followed by the longest leaf length and vice versa, while the narrowest leaf does not have the shortest leaf length. Data from

Fig. 2 shows that sucrose, PGR IBA have no effect, but the interaction between the two has a very significant effect on leaf width.

### 3.6. Root Length

Table 3 shows that sucrose, PGR IBA, and their interactions do not affect the root length, buds height, and plant height. The highest root length was obtained

from the treatment of base media MS +1 ppm BAP with the addition of 20-gram L<sup>-1</sup> sucrose and 0 ppm PGR IBA (S<sub>20</sub>I<sub>0</sub>), with an average root length of 2 cm, with the same amount of sucrose concentration and 3 ppm PGR IBA producing root length an average of 1.96 cm. The lowest root length resulted from treatment of 0-gram sucrose L<sup>-1</sup> and 1.5 ppm IBA (Fig. 3).

Table 1 Recapitulation of variance of various variables observation of pineapple rooting from Sipahutar

Week	Observation 1 - 12 WAP								
	Number of root			Number of leaves			Number of tillers buds		
	S	I	S&I	S	I	S&I	S	I	S&I
1	nd	nd	nd	0.21 <sup>ns</sup>	2.91 <sup>ns</sup>	0.60 <sup>ns</sup>	1 <sup>ns</sup>	1 <sup>ns</sup>	1 <sup>ns</sup>
2	1.11 <sup>ns</sup>	1.54 <sup>ns</sup>	2.46 <sup>ns</sup>	1.84 <sup>ns</sup>	2.77 <sup>ns</sup>	0.35 <sup>ns</sup>	2.2 <sup>ns</sup>	0.55 <sup>ns</sup>	0.55 <sup>ns</sup>
3	1.52 <sup>ns</sup>	0.19 <sup>ns</sup>	2.6 <sup>*</sup>	2.04 <sup>ns</sup>	0.08 <sup>ns</sup>	2.78 <sup>*</sup>	3.94 <sup>*</sup>	2.05 <sup>ns</sup>	1.21 <sup>ns</sup>
4	1.88 <sup>ns</sup>	0.34 <sup>ns</sup>	5.14 <sup>**</sup>	2.75 <sup>ns</sup>	0.13 <sup>ns</sup>	0.99 <sup>ns</sup>	4.92 <sup>**</sup>	0.46 <sup>ns</sup>	0.91 <sup>ns</sup>
5	1.5 <sup>ns</sup>	0.35 <sup>ns</sup>	5.59 <sup>**</sup>	4.33 <sup>*</sup>	0.08 <sup>ns</sup>	1.84 <sup>ns</sup>	6.1 <sup>**</sup>	0.76 <sup>ns</sup>	1.7 <sup>ns</sup>
6	0.6 <sup>ns</sup>	1.08 <sup>ns</sup>	4.9 <sup>**</sup>	4.53 <sup>*</sup>	0.19 <sup>ns</sup>	2.16 <sup>ns</sup>	6.58 <sup>**</sup>	0.31 <sup>ns</sup>	1.33 <sup>ns</sup>
7	0.07 <sup>ns</sup>	1.85 <sup>ns</sup>	3.64 <sup>*</sup>	4.08 <sup>*</sup>	0.10 <sup>ns</sup>	2.27 <sup>ns</sup>	6 <sup>**</sup>	0.18 <sup>ns</sup>	1.43 <sup>ns</sup>
8	1.6 <sup>ns</sup>	1.21 <sup>ns</sup>	2.13 <sup>ns</sup>	5.62 <sup>**</sup>	0.36 <sup>ns</sup>	3.15 <sup>*</sup>	5.45 <sup>**</sup>	0.26 <sup>ns</sup>	1.58 <sup>ns</sup>
9	1.96 <sup>ns</sup>	0.89 <sup>ns</sup>	3.22 <sup>*</sup>	4.94 <sup>**</sup>	0.27 <sup>ns</sup>	3.37 <sup>*</sup>	5.47 <sup>**</sup>	0.46 <sup>ns</sup>	1.75 <sup>ns</sup>
10	2.24 <sup>ns</sup>	1.31 <sup>ns</sup>	2.91 <sup>*</sup>	5.70 <sup>**</sup>	0.79 <sup>ns</sup>	5.14 <sup>*</sup>	4.9 <sup>**</sup>	0.27 <sup>ns</sup>	1.46 <sup>ns</sup>
11	2.24 <sup>ns</sup>	1.31 <sup>ns</sup>	2.91 <sup>*</sup>	5.91 <sup>**</sup>	0.37 <sup>ns</sup>	3.62 <sup>*</sup>	5.47 <sup>**</sup>	0.36 <sup>ns</sup>	1.59 <sup>ns</sup>
12	2.24 <sup>ns</sup>	1.31 <sup>ns</sup>	2.91 <sup>*</sup>	6.13 <sup>**</sup>	0.29 <sup>ns</sup>	3.11 <sup>*</sup>	5.58 <sup>**</sup>	0.4 <sup>ns</sup>	1.55 <sup>ns</sup>

Notes: \*\* - highly significant; \* - significant; nd - no data; ns - not significant; S - sucrose; I - IBA; S&I - sucrose and IBA interaction

Table 2 Average number of roots, number of leaves, leaf length, and leaf width on the observation of root formation of Sipahutar pineapple in 12 WAP

Treatment	Number of root	Number of leave	Leaf length	Leaf width
S <sub>0</sub> I <sub>0</sub>	1.34 ± 0.21 <sup>abc</sup>	9.00 ± 6.24 <sup>ab</sup>	1.80 ± 0.25 <sup>bcde</sup>	0.98 ± 0.08 <sup>ab</sup>
S <sub>0</sub> I <sub>1,5</sub>	0.71 ± 0.00 <sup>a</sup>	10.00 ± 6.08 <sup>ab</sup>	1.45 ± 0.26 <sup>ab</sup>	1.00 ± 0.00 <sup>abcd</sup>
S <sub>0</sub> I <sub>3</sub>	1.76 ± 0.31 <sup>bc</sup>	2.67 ± 0.58 <sup>a</sup>	2.35 ± 0.04 <sup>e</sup>	1.11 ± 0.05 <sup>cd</sup>
S <sub>10</sub> I <sub>0</sub>	1.65 ± 0.38 <sup>bc</sup>	12.33 ± 3.06 <sup>a</sup>	2.10 ± 0.32 <sup>cde</sup>	1.08 ± 0.09 <sup>bcd</sup>
S <sub>10</sub> I <sub>1,5</sub>	1.81 ± 0.56 <sup>bc</sup>	13.33 ± 6.43 <sup>ab</sup>	1.97 ± 0.33 <sup>bcde</sup>	1.12 ± 0.11 <sup>d</sup>
S <sub>10</sub> I <sub>3</sub>	1.46 ± 0.21 <sup>bc</sup>	3.00 ± 1.73 <sup>a</sup>	1.51 ± 0.44 <sup>ab</sup>	0.97 ± 0.03 <sup>a</sup>
S <sub>20</sub> I <sub>0</sub>	2.00 ± 0.49 <sup>c</sup>	12.00 ± 12.12 <sup>a</sup>	1.78 ± 0.49 <sup>bcde</sup>	0.98 ± 0.08 <sup>abc</sup>
S <sub>20</sub> I <sub>1,5</sub>	1.25 ± 0.94 <sup>ab</sup>	16.00 ± 12.12 <sup>b</sup>	1.15 ± 0.13 <sup>a</sup>	0.91 ± 0.08 <sup>a</sup>
S <sub>20</sub> I <sub>3</sub>	1.96 ± 0.67 <sup>bc</sup>	31.33 ± 15.89 <sup>c</sup>	2.28 ± 0.10 <sup>de</sup>	1.09 ± 0.12 <sup>bcd</sup>
S <sub>30</sub> I <sub>0</sub>	1.56 ± 0.33 <sup>bc</sup>	6.33 ± 5.13 <sup>ab</sup>	1.80 ± 0.58 <sup>bcde</sup>	1.00 ± 0.08 <sup>abcd</sup>
S <sub>30</sub> I <sub>1,5</sub>	1.83 ± 0.44 <sup>bc</sup>	6.33 ± 1.15 <sup>ab</sup>	1.71 ± 0.39 <sup>bcd</sup>	1.05 ± 0.08 <sup>bcd</sup>
S <sub>30</sub> I <sub>3</sub>	1.34 ± 0.21 <sup>abc</sup>	11.33 ± 6.03 <sup>ab</sup>	1.64 ± 0.40 <sup>abc</sup>	1.06 ± 0.10 <sup>bcd</sup>

Note: Numbers followed by the same lowercase letter in the same column or row show no significant difference at the Duncan Multiple Range Test (DMRT) 5%

Table 3 Recapitulation of variance of root length, leaf length, leaf width, buds' height, overall plant height, plant height

Week	Observation 12 WAP								
	Root Length			Leaf Length			Leaf Width		
	S	I	S&I	S	I	S&I	S	I	S&I
12	1.30 <sup>ns</sup>	0.73 <sup>ns</sup>	1.09 <sup>ns</sup>	0.61 <sup>ns</sup>	4.95 <sup>*</sup>	4.95 <sup>**</sup>	1.32 <sup>ns</sup>	1.74 <sup>ns</sup>	3.98 <sup>**</sup>
12	Buds height			Plant height					
	2.79 <sup>ns</sup>	0.13 <sup>ns</sup>	2.24 <sup>ns</sup>	2.11 <sup>ns</sup>	0.81 <sup>ns</sup>	1.66 <sup>ns</sup>			

Notes: \*\* - highly significant; \* - significant; nd - no data; ns - not significant; S - sucrose; I - IBA; S&I - sucrose and IBA interaction

### 3.7. Tiller Buds Height

Data from observations of the average height of shoots of pineapple shoots from Sipahutar showed the results of 20 grams sucrose L<sup>-1</sup> treatment and 3 ppm PGR IBA (S<sub>20</sub>I<sub>3</sub>) higher than all treatments with an average of 1.72 ± 0.56 and the lowest in the treatment of 30 grams sucrose L<sup>-1</sup> and 0 ppm PGR IBA (S<sub>30</sub>I<sub>0</sub>) with an average of 0.81 ± 0.17, while in the 0 grams sucrose L<sup>-1</sup> treatment and 3 ppm IBA (S<sub>0</sub>I<sub>3</sub>), 10 grams sucrose L<sup>-1</sup> and 3 ppm IBA (S<sub>10</sub>I<sub>3</sub>), and 30-gram L<sup>-1</sup>

sucrose and 1.5 ppm IBA (S<sub>30</sub>I<sub>1,5</sub>) did not produce new shoots (Fig. 4).

### 3.8. Plant Height

Data from observations of the highest average plant height is from 10 grams L<sup>-1</sup> sucrose and 0 ppm IBA treatments, with an average plant height of 5.67 cm, not significantly different from 10 grams sucrose L<sup>-1</sup> treatment with 1.5 ppm IBA or 0-gram sucrose L<sup>-1</sup> and 3 ppm IBA and 20-gram L<sup>-1</sup> sucrose and 3 ppm IBA (Fig. 5).

### 3.9. Time of Root Emergence

The emergence of roots produced from a base medium of MS +1 ppm BAP with the addition of 20-gram sucrose L<sup>-1</sup> with 3 ppm IBA (S<sub>20</sub> I<sub>3</sub>) began in the second week. The rest of the roots generally begin at week 4 (Fig. 6). In this study, media without the addition of sucrose combined with 1.5 ppm IBA (S<sub>0</sub>I<sub>1.5</sub>) did not show the appearance of new roots (Table 4).

### 3.10. Time of Leaves' Increase

The number of leaves has begun to increase since the first and second week after planting. Exception for

media without the addition of sucrose and PGR IBA (S<sub>0</sub>I<sub>0</sub>), and media with the addition of 10 grams sucrose L<sup>-1</sup> and 3 ppm PGR IBA (S<sub>10</sub>I<sub>3</sub>), new leaves begin to appear in the third week.

### 3.11. Time of Tillers' Bud Emergence

The emergence time of tillers bud varies greatly from the first to the eighth. With moderate treatment, the addition of 20 grams sucrose L<sup>-1</sup> and 1.5 ppm PGR IBA (S<sub>20</sub> I<sub>1.5</sub>) produced the fastest tillers bud that is one week after planting (Table 4). Certain treatments did not produce new tillers bud (Fig. 7).

Table 4 Time of root emergence, time of number of leaves increases, time of tillers bud emergence

Treatment	Time of root emergence	Time of number of leaves increases	Time of tillers bud emergence
S <sub>0</sub> I <sub>0</sub>	Week 5	Week 3	Week 6
S <sub>0</sub> I <sub>1.5</sub>	No root appeared	Week 1	Week 5
S <sub>0</sub> I <sub>3</sub>	Week 4	Week 2	No tillers appeared
S <sub>10</sub> I <sub>0</sub>	Week 5	Week 1	Week 8
S <sub>10</sub> I <sub>1.5</sub>	Week 4	Week 1	Week 4
S <sub>10</sub> I <sub>3</sub>	Week 4	Week 3	No tillers appeared
S <sub>20</sub> I <sub>0</sub>	Week 4	Week 1	Week 4
S <sub>20</sub> I <sub>1.5</sub>	Week 8	Week 1	Week 1
S <sub>20</sub> I <sub>3</sub>	Week 2	Week 1	Week 2
S <sub>30</sub> I <sub>0</sub>	Week 4	Week 1	Week 7
S <sub>30</sub> I <sub>1.5</sub>	Week 4	Week 1	No tillers appeared
S <sub>30</sub> I <sub>3</sub>	Week 3	Week 2	Week 3
<b>Abbreviations</b>			
S <sub>0</sub> I <sub>0</sub>	= Sucrose 0gram L <sup>-1</sup> + IBA 0 gram/l	S <sub>20</sub> I <sub>0</sub>	= Sucrose 20 gram L <sup>-1</sup> + IBA0 gram L <sup>-1</sup>
S <sub>0</sub> I <sub>1.5</sub>	= Sucrose 0gram L <sup>-1</sup> + IBA1.5 gram/l	S <sub>20</sub> I <sub>1.5</sub>	= Sucrose 20 gram L <sup>-1</sup> + IBA1.5 gram L <sup>-1</sup>
S <sub>0</sub> I <sub>3</sub>	= Sucrose 0gram L <sup>-1</sup> + IBA3 gram/l	S <sub>20</sub> I <sub>3</sub>	= Sucrose 20 gram L <sup>-1</sup> + IBA3 gram L <sup>-1</sup>
S <sub>10</sub> I <sub>0</sub>	= Sucrose 10 gram L <sup>-1</sup> + IBA0 gram L <sup>-1</sup>	S <sub>30</sub> I <sub>0</sub>	= Sucrose 30 gram L <sup>-1</sup> + IBA0 gram L <sup>-1</sup>
S <sub>10</sub> I <sub>1.5</sub>	= Sucrose 10 gram L <sup>-1</sup> + IBA1.5 gram L <sup>-1</sup>	S <sub>30</sub> I <sub>1.5</sub>	= Sucrose 30 gram L <sup>-1</sup> + IBA1.5 gram L <sup>-1</sup>
S <sub>10</sub> I <sub>3</sub>	= Sucrose 10 gram L <sup>-1</sup> + IBA3 gram L <sup>-1</sup>	S <sub>30</sub> I <sub>3</sub>	= Sucrose 30 gram L <sup>-1</sup> + IBA3 gram L <sup>-1</sup>

## 4. Discussion

This study showed information that sucrose was needed for root formation. This can be clarified by the research results showing that the highest number of roots is produced from sucrose-treated treatments, without ignoring the role of PGR IBA. It was seen in all treatments with PGR IBA. Also, the acceleration of root emergence is faster than without PGR IBA. The data above showed that media without the addition of sucrose combined with 1.5 ppm IBA did not produce roots, meaning that although IBA is present but without an energy source, namely sucrose, the roots will not appear. Research from [17] found that sugar has provided the tissue culture plant with carbon in an

organic form that is not required for those grown from seeds. This process indicates that the PGR IBA serves to accelerate the emergence of roots.

Early emergence of roots is a success factor in pineapple rooting from Sipahutar with in vitro technique. The interaction between sucrose and IBA gives a better effect than a single factor. Data from Table 4 indicate that the addition of IBA to the base media of MS without adding the sucrose is not enough to induce the emergence of roots. It is shown from the treatment of sucrose 0 gram L<sup>-1</sup> with 1.5 ppm IBA (S<sub>0</sub>I<sub>1.5</sub>). Hence, to induce the emergence of new roots, it takes a source of energy to increase metabolic processes, in this study is sucrose.

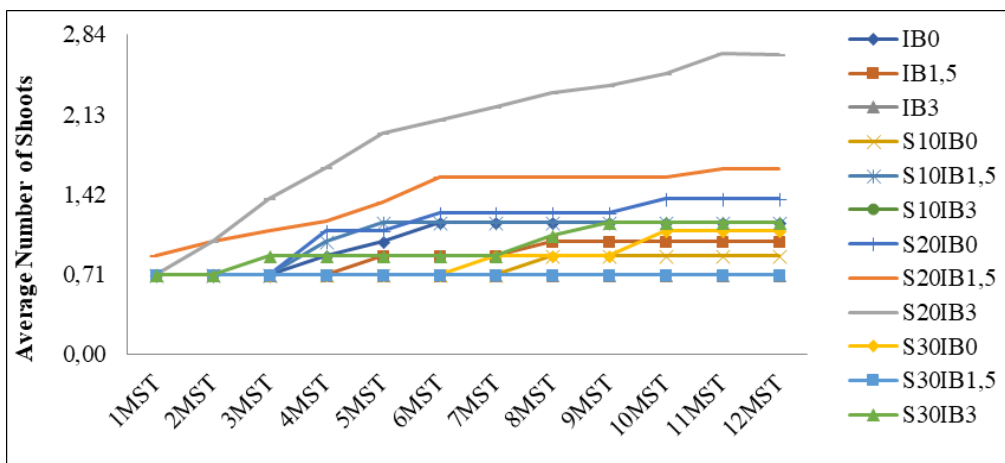


Fig. 1 Number of shoots from Sipahutar pineapple for 12 weeks

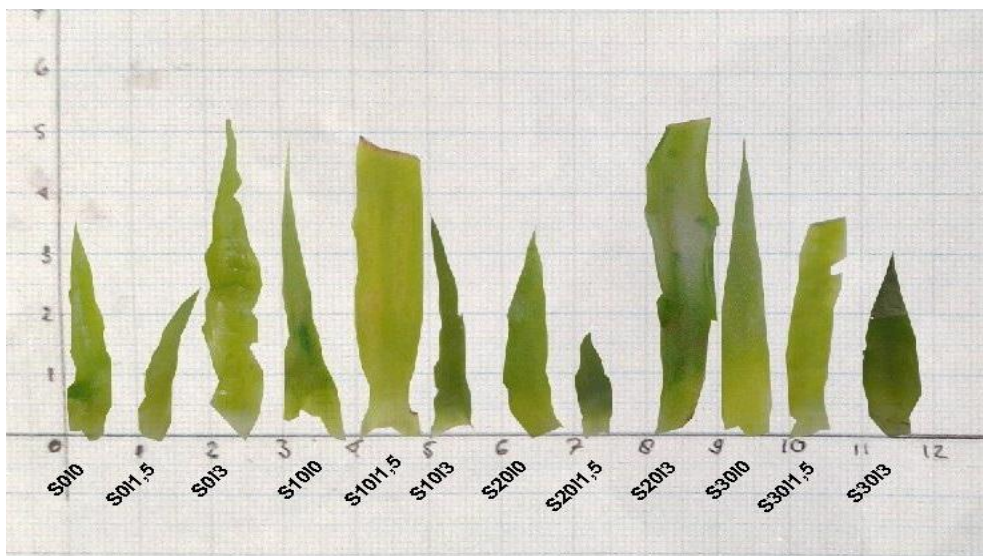


Fig. 2 Documentation of leaf length and leaf width on various concentrations of sucrose and IBA 12 WAP, with treatment S<sub>0</sub>I<sub>0</sub>, S<sub>0</sub>I<sub>1.5</sub>, S<sub>0</sub>I<sub>3</sub>, S<sub>10</sub>I<sub>0</sub>, S<sub>10</sub>I<sub>1.5</sub>, S<sub>10</sub>I<sub>3</sub>, S<sub>20</sub>I<sub>0</sub>, S<sub>20</sub>I<sub>1.5</sub>, S<sub>20</sub>I<sub>3</sub>, S<sub>30</sub>I<sub>0</sub>, S<sub>30</sub>I<sub>1.5</sub>, S<sub>30</sub>I<sub>3</sub>

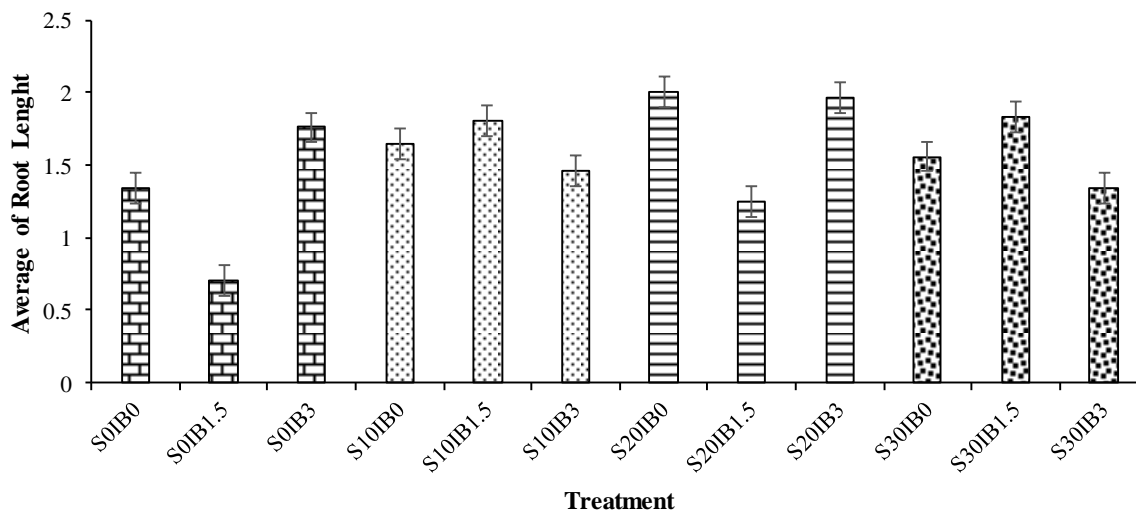


Fig. 3 Average length of pineapple roots from Sipahutar age 12 WAP

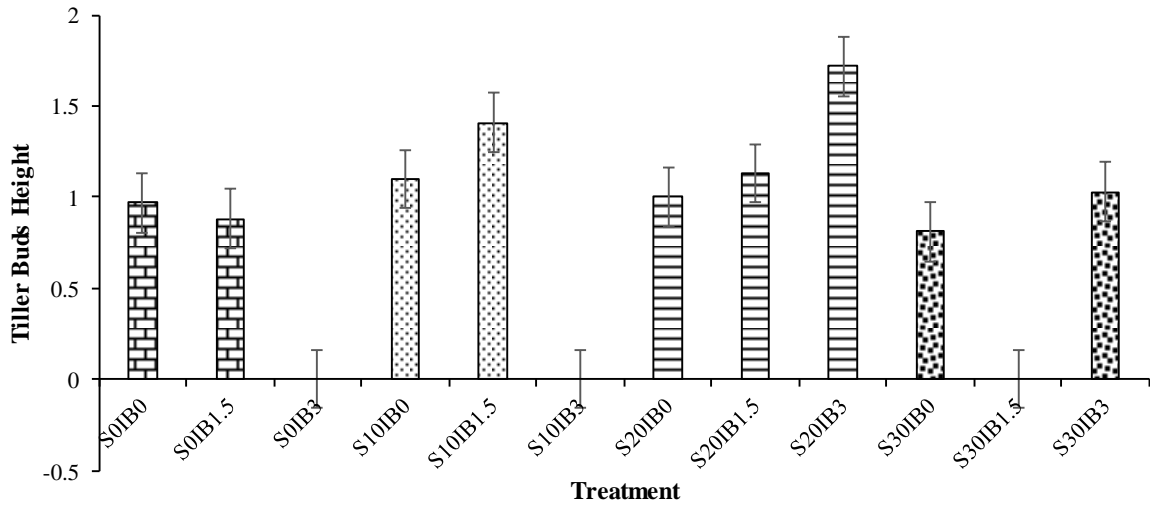


Fig. 4 The average tiller buds height of Sipahutar pineapple

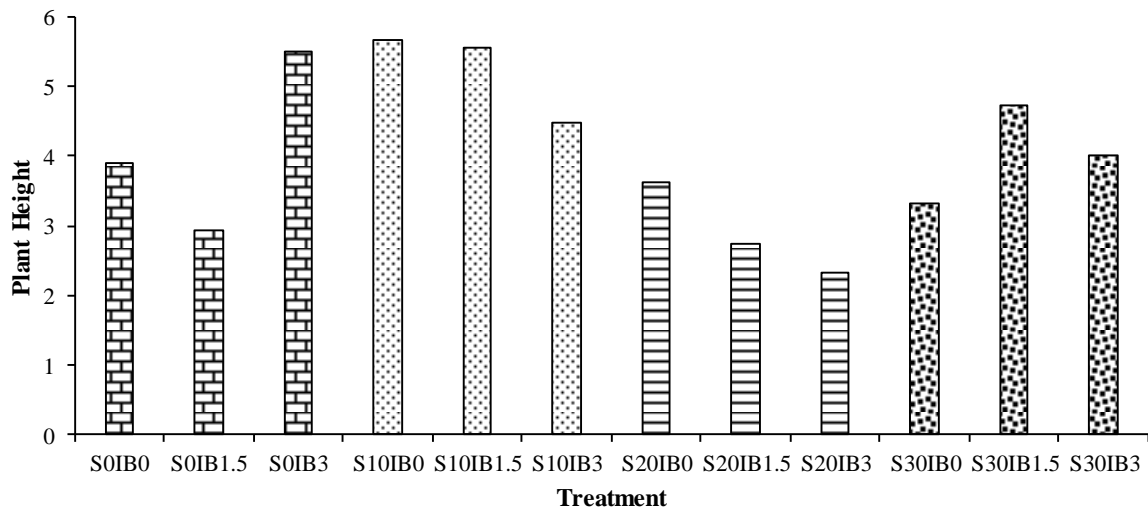


Fig. 5 Plant height of pineapple from Sipahutar age 12 WAP

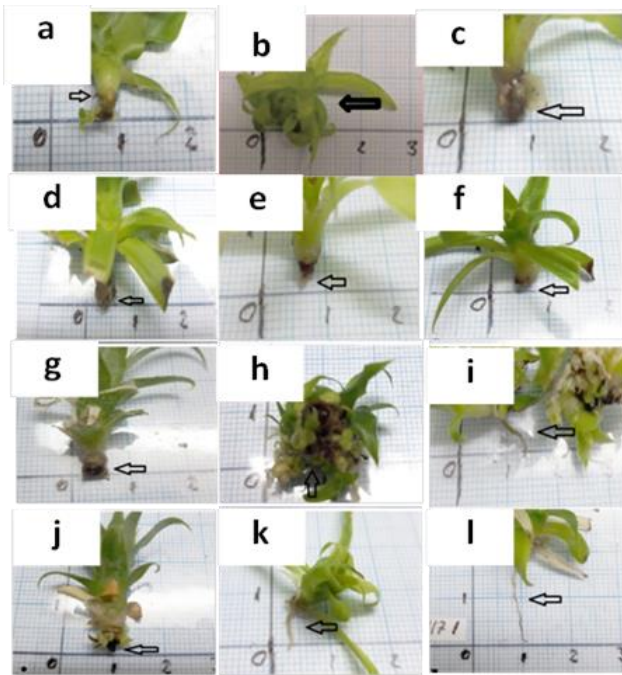


Fig. 6 Documentation of root lengths of various concentrations of sucrose and IBA 12 WAP, on the treatment

- a. A S<sub>0</sub>I<sub>0</sub> = Sucrose 0 gram L<sup>-1</sup> + IBA 0 gram L<sup>-1</sup>
- b. B S<sub>0</sub>I<sub>1.5</sub> = Sucrose 0 gram L<sup>-1</sup> + IBA 1.5 gram L<sup>-1</sup>
- c. C S<sub>0</sub>I<sub>3</sub> = Sucrose 0 gram L<sup>-1</sup> + IBA 3 gram L<sup>-1</sup>

- d. D S<sub>10</sub>I<sub>0</sub> = Sucrose 10 gram L<sup>-1</sup> + IBA 0 gram L<sup>-1</sup>
- e. E S<sub>10</sub>I<sub>1.5</sub> = Sucrose 10 gram L<sup>-1</sup> + IBA 1.5 gram L<sup>-1</sup>
- f. F S<sub>10</sub>I<sub>3</sub> = Sucrose 10 gram L<sup>-1</sup> + IBA 3 gram L<sup>-1</sup>
- g. G S<sub>20</sub>I<sub>0</sub> = Sucrose 20 gram L<sup>-1</sup> + IBA 0 gram L<sup>-1</sup>
- h. H S<sub>20</sub>I<sub>1.5</sub> = Sucrose 20 gram L<sup>-1</sup> + IBA 1.5 gram L<sup>-1</sup>
- i. I S<sub>20</sub>I<sub>3</sub> = Sucrose 20 gram L<sup>-1</sup> + IBA 3 gram L<sup>-1</sup>
- j. J S<sub>30</sub>I<sub>0</sub> = Sucrose 30 gram L<sup>-1</sup> + IBA 0 gram L<sup>-1</sup>
- k. K S<sub>30</sub>I<sub>1.5</sub> = Sucrose 30 gram L<sup>-1</sup> + IBA 1.5 gram L<sup>-1</sup>
- l. L S<sub>30</sub>I<sub>3</sub> = Sucrose 30 gram L<sup>-1</sup> + IBA 3 gram L<sup>-1</sup>

According to [18], IBA is a type of auxin that is most often used in inducing roots compared to other types of auxin because of its high ability to initiate roots. Likewise, according to [19], the concentration of 20 grams of sucrose L<sup>-1</sup> is the best in influencing pineapple plant growth. The addition of 20 grams of sucrose L<sup>-1</sup> and 1.5 ppm IBA (S<sub>20</sub> I<sub>1.5</sub>) resulted in the acceleration of the emergence of new tiller buds.

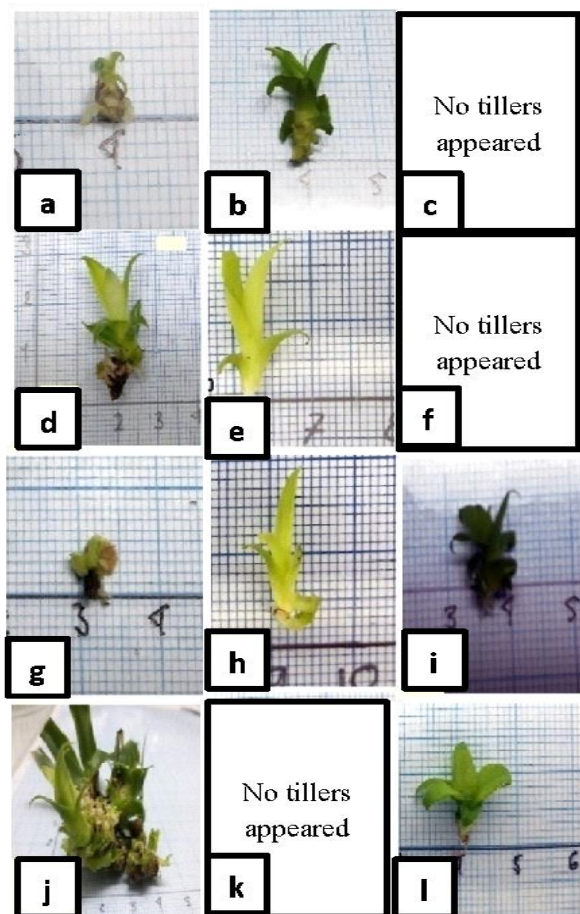


Fig. 7 Documentation of tillers' bud of various concentrations of sucrose and IBA 12 WAP, on the treatment

a.	A	SoI <sub>0</sub>	= Sucrose 0 gram L <sup>-1</sup> + IBA 0 gram L <sup>-1</sup>
b.	B	S <sub>0</sub> I <sub>1.5</sub>	= Sucrose 0 gram L <sup>-1</sup> + IBA 1.5 gram L <sup>-1</sup>
c.	C	SoI <sub>3</sub>	= Sucrose 0 gram L <sup>-1</sup> + IBA 3 gram L <sup>-1</sup>
d.	D	S <sub>10</sub> I <sub>0</sub>	= Sucrose 10 gram L <sup>-1</sup> + IBA 0 gram L <sup>-1</sup>
e.	E	S <sub>10</sub> I <sub>1.5</sub>	= Sucrose 10 gram L <sup>-1</sup> + IBA 1.5 gram L <sup>-1</sup>
f.	F	S <sub>10</sub> I <sub>3</sub>	= Sucrose 10 gram L <sup>-1</sup> + IBA 3 gram L <sup>-1</sup>
g.	G	S <sub>20</sub> I <sub>0</sub>	= Sucrose 20 gram L <sup>-1</sup> + IBA 0 gram L <sup>-1</sup>
h.	H	S <sub>20</sub> I <sub>1.5</sub>	= Sucrose 20 gram L <sup>-1</sup> + IBA 1.5 gram L <sup>-1</sup>
i.	I	S <sub>20</sub> I <sub>3</sub>	= Sucrose 20 gram L <sup>-1</sup> + IBA 3 gram L <sup>-1</sup>
j.	J	S <sub>30</sub> I <sub>0</sub>	= Sucrose 30 gram L <sup>-1</sup> + IBA 0 gram L <sup>-1</sup>
k.	K	S <sub>30</sub> I <sub>1.5</sub>	= Sucrose 30 gram L <sup>-1</sup> + IBA 1.5 gram L <sup>-1</sup>
l.	L	S <sub>30</sub> I <sub>3</sub>	= Sucrose 30 gram L <sup>-1</sup> + IBA 3 gram L <sup>-1</sup>

Sucrose, which at first prioritized to stimulate root growth, is less effective in giving effect to the rooting of pineapple from Sipahutar. It is suspected that the level of treatment factor and concentration are not suitable, making the Sipahutar pineapple plantlet less optimal in rooting. The use of optimal concentration is a factor that can determine the emergence of root explants planted by [18]. Sucrose has a significant effect on the number of leaves and the number of tillers bud.

This study showed that the interaction between sucrose and IBA has a significant effect on the parameters of root number. In S<sub>20</sub>I<sub>3</sub> treatment with an average of 1.96, the concentration of 20 grams of sucrose L<sup>-1</sup> with the addition of 3 ppm IBA significantly affected the number of pineapple roots from Sipahutar. The study of [20] also stated that the interaction of sucrose with IBA in medium

concentrations was more effective in increasing the number of roots. In contrast, high concentrations resulted in a decreasing number of roots. Likewise, according to [21], the interaction of sucrose and IBA can also increase root growth, but a single factor of sucrose is more to stimulate the formation of the buds. According to [22], sucrose will provide energy for the growth of pineapple explants from Sipahutar and, as a building substance, produce larger molecules needed for growth. Therefore, giving IBA as rooting stimulation interacting with sucrose can affect the growth of the number of pineapple explant roots from Sipahutar.

According to [23], sucrose, as a carbohydrate synthesized and transported naturally in plants, helps the number of roots grow. According to [24], IBA has better properties and effects of accelerating root growth, increasing the number of roots, and improving root quality. The effect of auxin on plant roots depends on concentration, method of application, time of addition, part of the plant, and age of plant tissue.

Sucrose is also needed to increase the number of leaves; this can be seen from the treatment without sucrose, which is given with a high dose of 3 ppm PGR IBA, which will produce a low number of leaves. This pictures sucrose contributes greatly to the supply of energy for the process of metabolism in tissue culture. [25] stated that carbohydrate is supplemented as a carbon source to maintain the carbon supply and osmotic potential of cells. Sucrose is widely used in plant tissue culture due to its most favorable effect on growth. This is a basic description that a very basic physiological process happened, that the treatment which produced the fewest number of leaves will have the longest leaves (Table 2).

The delayed emergence of new leaves causes of there is no addition of sucrose and IBA. However, high concentrations (3 ppm) of IBA also cause new leaves to be delayed. It can be obtained that to maximize growth (producing new leaves), it is needed an additional source of energy and growth regulators in certain doses. In tissue culture, certain optimum concentrations are needed for growth. This study illustrates that there is an optimum dose to increase the acceleration of Sipahutar pineapple rooting. This is indicated by adding 20 grams of sucrose L<sup>-1</sup> and 1.5 ppm IBA (S<sub>20</sub>I<sub>1.5</sub>), resulting in the acceleration of the emergence of new tiller buds.

The average time to increase the number of leaves in all treatments is 1 WAP. This showed that both single and combined factors could increase the number of leaves quickly due to the balance between the composition of growth regulators in the media. According to [26], the balance of growth regulators in the media with the addition of the right dose of sucrose can provide optimal nutrients in plants to accelerate the growth of the number of pineapple leaves from Sipahutar. Likewise, according to [27], in their role as



determining the direction of tissue development, the addition of auxins and cytokines must consider its concentration and comparison in the media.

The 20-gram sucrose L<sup>-1</sup> treatment gave a good stimulation for the growth of the number of pineapple leaves from Sipahutar. During plant tissue culture, sucrose acts as a fuel source to maintain photomixotrophic metabolism and a carbon precursor or metabolite signal to ensure optimal development [28]. The leaves are one of the important organs in plants, the higher the leaves, the better the growth of a plant. According to [29], leave is one of the most important plant organs, especially for photosynthesis, to produce photosynthate experience optimum growth. According to [30], the number of leaves is one of the indicators of plant growth and can be used as supporting data to explain the growth process; sucrose as a carbohydrate substance for plant growth and development affects the increase of pineapple leaves from Sipahutar.

S<sub>20</sub>I<sub>3</sub> treatment with an average of 31.33 showed that the addition of 20-gram sucrose L<sup>-1</sup> and 3 ppm IBA gave a good stimulation for the growth of the number of pineapple leaves from Sipahutar. Sucrose is the most common source of carbohydrates and carbon used in plant tissue culture media. This is because carbohydrates supply energy to plants, especially when it is not ready to photosynthesize during the early stages of tissue culture [31]. Increased addition of sucrose and auxin is proven can increase the number of leaves. [32] stated in her research that MS media added with sucrose can optimally stimulate leaf length. According to [30], the number of leaves is one of the indicators of plant growth that can be used as supporting data to explain the growth process. The addition of sucrose as a carbohydrate substance for plants for growth and development affects the increase of pineapple leaves from Sipahutar.

Observation showed that the earliest for the emergence of tillers bud was in the sucrose treatment of 20 gram L<sup>-1</sup> by giving 1.5 ppm IBA (S<sub>20</sub>I<sub>1.5</sub>). This shows sucrose as a carbon source, and the energy available in the media can be absorbed by the Sipahutar pineapple plantlet. According to [33], the growth and propagation of shoots in vitro are influenced by many factors, one of which is the type of carbon source added to the media. Carbon sources function as energy and osmotic agents to support plant tissue growth. Likewise, according to [34], the addition of sucrose in the media will be a source of energy and a source of carbon for explant cells to grow. Increasing the concentration of sucrose will cause the explant to obtain more energy and carbon sources to accelerate the growth of explants. The higher energy sources resulted in faster cell division so that the growth of tillers bud will be faster.

Explants treated with sucrose have a large number of tillers which first appear like white lumps, then these

lumps form young tillers with large numbers. According to [44], sucrose with high concentrations and added with high BAP will stimulate cell division multiple times and produce higher tillers. [35] stated that high BAP concentrations would inhibit the work of auxin so that it will accelerate the growth of tillers in explants.

According to [19], the concentration of sucrose 20 grams L<sup>-1</sup> can affect the growth of the number of roots, but in this study, sucrose has a significant effect on the growth of the number of leaves and the number of Sipahutar pineapple tillers. This might be caused by adding 1 ppm BAP to base media whose main function is to promote tillers' growth. According to [36], adding sucrose will produce lots of tillers because sucrose and BAP can stimulate cell division in plants, sucrose as a source of carbohydrates to meet the needs of plants in cell division and plant tillers growth. Formed tillers are indicated as a success of the regeneration of explants that were inoculated in tissue culture media [37]. The faster the tillers appear, the faster the plant propagation material is produced. Exogenous applications of cytokinins and auxins have been known to be important for the induction of buds and elongation in many plant species in vitro [38].

The results of the research analysis showed that IBA has a significant effect on leaf length and has no significant effect on the following parameters: the number of leaves, leaf width, length of tillers bud, number of tillers, root length, and number of roots. IBA has a significant effect on the leaf length of Pineapple from Sipahutar at IBA 3 ppm. The addition of auxin, which is often used for root growth, can also affect the growth of leaf length showed in this study; therefore, the basal medium used in this research is MS medium + 1 ppm BAP. [39] stated that the number and length of roots influence the height and number of leaves in plants which with a positive correlation between the number and lengths of roots with the nutrients absorbed are relatively larger, so plants have the opportunity to form more leaves and vice versa, if a little amount of the roots are produced and short, the nutrients absorbed are relatively small and the leaves are formed smaller. Indol-3-Butyric-Acid (IBA) is better than other auxins for rooting stimulation [40]. The main function of IBA as a root growth stimulation hormone does not provide a significant effect on the growth of pineapple roots from Sipahutar. It is suspected because BAP given to the base media, which was already high enough in concentration, made IBA stop stimulating the growth of pineapple roots from Sipahutar. According to [39], IBA has activity as a rooting hormone, so that IBA activity can affect the number of roots. If the addition of IBA is too high, it can inhibit root growth. It is suspected that the use of a high concentration of IBA following the addition of 1 ppm of BAP in base media is too high to stimulate the growth of pineapple roots. When combined with high concentrations, auxin and

cytokinin can inhibit the growth of pineapple roots from Sipahutar. However, in the study of [41], the interaction of IBA and BAP can stimulate long and dense root growth compared to the single factor.

Interaction of sucrose with IBA significantly affected the length of pineapple leaves from Sipahutar, with an average of 2.35 in the S<sub>20</sub>I<sub>3</sub> treatment. The addition of 20 grams of sucrose L<sup>-1</sup> with a concentration of 3 ppm IBA and 1 ppm BAP on MS base media can affect the length of pineapple plantlets from Sipahutar. The study of [40] stated that MS media with BAP could affect the length of the leaves in the tillers bud that emerges. According to [42], the addition of sucrose and BAP to MS media can initiate the growth of plant parts because sucrose, which provides carbohydrates for plants, acts optimally in the growth of plant parts.

The interaction between sucrose and IBA significantly affected leaf width. Observation results showed that the highest average value in leaf width was in the S<sub>10</sub>I<sub>1.5</sub> treatment with an average of 1.12, interactions of 10 grams sucrose L<sup>-1</sup> with 1.5 ppm IBA giving a significant effect on the width of pineapple leaves from Sipahutar. According to [14], adding auxin and cytokinins in accordance with plants can affect the development of plant parts; adding organic substances such as sucrose with the appropriate concentration can provide energy substances for plants. Sucrose is easily available to cells and directly participates in the glycolytic and pentose phosphate pathways for cell growth and is also found to act as osmotic [43].

## 5. Conclusion

The addition of sucrose significantly affected the number of leaves and the number of tillers. While IBA (Indole Bricity Acid) as PGR significantly affects the length of the pineapple leaves from Sipahutar. This showed that IBA stimulates root growth and can stimulate leaf length with the appropriate concentration. The interaction between sucrose and IBA significantly affects the number of roots, leaf length, and leaf width of pineapple plants from Sipahutar. IBA, which interacted with sucrose, can stimulate the number of roots and growth of pineapple from Sipahutar. The limitation of this study is that the test was only conducted in vitro and not yet in vivo. In addition, the best results from each treatment need to be further tested in vivo.

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