

Lagerstroemia, *Euphorbia Hirta*, and *Kleinhovia Hospita* as Inhibitors of Heptad Repeat (HR) SARS-CoV-2 Spike Protein Based on an *In Silico* Study

Jantje Wiliem Souhaly, Sapti Puspitarini, M. Hermawan Widyananda, Nashi Widodo

Department of Biology, Faculty of Mathematics and Natural Science, Brawijaya University, Malang, Indonesia

Abstract: Severe acute respiratory syndrome-coronavirus 2 (SARS-CoV-2), the causative agent of coronavirus disease-19 (COVID-19), has become a global health issue. Spike proteins from the virus have a vital role in infection. Herbal medicines such as *Lagerstroemia*, *Euphorbia hirta*, and *Kleinhovia hospita* have several pharmacological functions such as anticancer, antiviral, and antioxidant because of their bioactive compound content. Based on an in silico study, this research was conducted on the possibility of phytochemicals from herbal *Lagerstroemia*, *E. hirta*, and *K. hospita* to inhibit spike protein SARS-CoV-2. A three-dimensional (3D) compound structure of each herbal medicine was docked with HR protein using AutoDock Vina software. The docking result, which has the best binding energy value, is continued with the analysis of molecular dynamics simulation. Lagerine, rutin, and nicotiflorin compounds might bind to proteins with lower binding energy. Protein was unstable when complexed with compounds compared with control, as seen from the root-mean-square deviation (RMSD) value. Therefore, this research is pre-experimental to inhibit SARS-CoV-2 spike proteins by herbal medicines.

Keywords: COVID-19, *Euphorbia hirta*, *Kleinhovia hospita*, *Lagerstroemia*, severe acute respiratory syndrome-coronavirus 2.

基于计算机研究的紫薇、大戟属赫塔和克莱因霍维亚医院作为七肽重复(人力资源)非典-冠状病毒-2 刺突蛋白的抑制剂

摘要: 严重急性呼吸综合征冠状病毒 2(非典-冠状病毒-2)是冠状病毒病 19(新冠肺炎)的病原体,已成为全球健康问题。来自病毒的刺突蛋白在感染中起着至关重要的作用。紫薇、大戟和小花等中草药因其生物活性化合物含量而具有抗癌、抗病毒和抗氧化等多种药理功能。基于一项计算机研究,这项研究是针对草药紫薇、E.希尔塔和 K.医院中的植物化学物质抑制刺突蛋白非典-冠状病毒-2 的可能性进行的。使用自动停靠维娜 软件将每种草药的三维(3D)复合结构与人力资源蛋白对接。结合能值最佳的对接结果继续进行分子动力学模拟分析。紫薇、芦丁和烟花苷化合物可能与结合能较低的蛋白质结合。从均方根偏差(RMSD)值可以看出,与对照相比,当与化合物复合时,蛋白质是不稳定的。因此,这项研究是通过草药抑制非典-冠状病毒-2 刺突蛋白的预实验。

关键词: 新冠肺炎, 大戟, 克莱因霍维亚医院, 紫薇, 严重急性呼吸综合征冠状病毒 2。

1. Introduction

At the end of December 2019, in Wuhan, Hubei Province, China, a novel coronavirus disease (COVID-19) caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) was discovered. It is

rapidly spreading from its origin to the rest of the world, causing a pandemic [1, 2]. As of November 2021, more than 260 million people have become infected, and over 5 million have died because of this pandemic [28]. Although several vaccines are already

found, the war against COVID-19 continues, and complementary medicine and diet are needed to prevent the severity of the disease [2, 3].

Prevent replication and attachment respiratory of syndrome-coronavirus (SARS-CoV-2) become the main target in combating COVID-19 [4, 5]. Previous studies have shown that several proteins from SARS-CoV-2 have been modeled and used as a target to decrease the number of positive cases [6–8]. The primary key of SARS-CoV-2 to enter the host cells is the spike protein, consisting of parts, such as heptad repeat 1 (HR1) and HR2 inside the receptor-binding domain (RBD) to form a fusion of membrane after attachment [7, 9, 10]. Based on this critical role, HR1 and HR2 can be the main targets to evade viral entry and infection [7].

Several herbal medicines with active compounds used as antioxidants, anticancer, antibacterial, and antiviral drugs were reported [11]. Antioxidants are used to prevent damage to cells caused by free radicals. In China, herbal medicines are used as conventional therapies for patients infected with SARS-CoV-2 [12, 13]. In Indonesia, herbal medicines are expected to provide treatment for SARS-CoV-2 patients. The current study focused on three herbal medicines, *Lagerstroemia*, *E. hirta*, and *K. hospita*, as potential treatments for COVID-19. In addition, several compounds from herbal medicines, such as rutin and nicotiflorin, were reported to have antioxidant and antiviral effects [14–16].

2. Materials and Method

2.1. Ligand and Protein Selection

The compounds of herbal medicines were collected from the KNApSAcK database [30]. The structures of three-dimensional (3D) compounds were assessed through the PubChem database. The 3D structures of protein were obtained from the Protein Data Bank (PDB) [29]. The HR structures were retrieved from the PDB (PDB ID: 6LXT).

2.2. Molecular Docking

Protein-ligand docking was analyzed using AutoDock Vina integrated into PyRx 0.8 [17, 18]. Docking was performed on the active side of HR. Herbal compounds were screened using molecular docking. Protein-protein docking was performed using HEX 8.0, was run using default settings, and was operated under the Shape + DARS correlation type [19]. The results of docking and protein-ligand interactions were analyzed using BIOVIA Discovery Studio.

2.3. Molecular Dynamics Simulation

The best binding affinity from the docking results was continued to molecular dynamics simulation using the software Yet Another Scientific Artificial Reality

Application (YASARA) [20]. The parameters used were in accordance with the cell physiological conditions (37°C, 1 atm, pH 7.4, and salt content of 0.9%) for 20 ns with autosave every 25 ps. The simulation was run using the md_runfast macro program, and the root-mean-square deviation (RMSD) was displayed using the macro md_analyze program.

2.4. Potential Compounds

Potential compounds from *Lagerstroemia*, *E. hirta*, and *K. hospita* were analyzed using Way2Drug Pass Server prediction to determine antiviral activity based on Pa value (Probability to Be Active).

3. Results and Discussion

Through the KNApSAcK retrieval, 22 compounds from *Lagerstroemia*, *E. hirta*, and *K. hospita* were collected. The results of the docking HR based on binding energy showed that a compound from each herbal medicine, such as lagerine, rutin, and nicotiflorin, has a lower binding energy value of -7.5 , -6.9 , and -7 kcal/mol for HR1 (Table 1) and -1175.3 , -986.9 , and -1027.8 kcal/mol for HR1–HR2 (complex) (Table 2). The binding energy of the HR complex after binding to this compound could decrease compared to that of the HR complex without the ligand (Table 2). This compound could also change the HR1–HR2 binding motif (Fig. 1a). Among the three compounds that have the best affinity to HR1, rutin was able to modify the HR1–HR2 interaction. This was described by the binding energy value of the HR complex with rutin, which had the highest value among the complexes, compared to the HR complex alone. Further analysis used molecular dynamics simulation to evaluate the interaction of compounds when interacting with HR1–HR2.

Table 1 Binding energy of *Lagerstroemia*, *Euphorbia hirta*, and *Kleinhovia hospita*

Herbal	Compound	CID	Binding Energy
			HR1
<i>Lagerstroemia</i>	Cryogenine	61002	-5.9
	Petunidin 3-araboside	6325802	-6.6
	Petunidin 3-O-beta-D-glucopyranoside	443651	-6.3
	Malvidin 3-araboside	12137511	-6.5
	Oenin	443652	-6.2
	Lagerstronolide	101445553	-6.1
	(+/-)-Epiepoxydon	331736	-5
	Methyl 3,4,5-trihydroxybenzoate	7428	-4.7
	Isoprene	6557	-4
	5-Epi-dihydrolyfoline	42640298	-7.1
	Dihydrolyfoline	42640297	-6.5
	Lagerine	101473393	-7.5
	Decamine	2993	-6.1
<i>Euphorbia hirta</i>	Lagerstremine	78385190	-6
	alpha-Amyrin	73170	-6.6
	beta-Amyrin	73145	-6.6
	Taraxerol	92097	-6.9
	Rutin	5280805	-6.9
	Ellagic acid	5281855	-6.2
<i>Kleinhovia hospita</i>	Euphorbianin	44259305	-6.4
	Nicotiflorin	5318767	-7
	Rutin	5280805	-6.9

Note: Compounds with high binding energy are marked in bold

Table 2 Binding energy of HR complex + ligand

Herbal	Compound	CID	Binding Energy
HR Complex + ligand			
<i>Lagerstroemia</i>	Lagerine	101473393	-1175.3
<i>Euphorbia hirta</i>	Rutin	5280805	-986.9
<i>Kleinhovia hospita</i>	Nicotiflorin	5318767	-1027.8
HR1 & HR2 (Control)		12137511	-3136.8

Molecular dynamics simulations were carried out to analyze the structural stability and conformational fluctuations of HR protein interaction with compounds. The important information regarding the stability and flexibility of protein-ligand complexes during the simulation is shown in RMSD. High deviation and variability indicate low stability [21]. Proteins that were stable during the simulation had RMSD values less than 3Å. An RMSD value greater than 3Å indicates that the protein has undergone some structural changes [22]. The molecular dynamics simulation results show that the protein has structural changes (Fig. 1b), and the HR complex with compound inside showed that the RMSD value was more than 3Å compared with the HR complex alone (Fig. 1c). The crucial step for SARS-CoV-2 fusion to membranes is the interaction of HR1–HR2 to form a helix bundle [23, 24]. Modification of the helix bundle formation has been studied to prevent viral entry [7]. A compound such as Lagerine, nicotiflorin, and rutin has an RMSD value of more than 3Å, which may have an excellent potency in inhibiting viral infection. The fusion process is essential for virus entry into the cell host. HR1 and HR2 are located in the S2 subunit regions, which, when exposed, form the fusion core. The formation of the fusion core induces virus membrane fusion with the host cell membrane. The herbal compound can bind to HR1 more strongly and prevent HR2 from binding to HR1 to form the fusion core (Fig. 2).

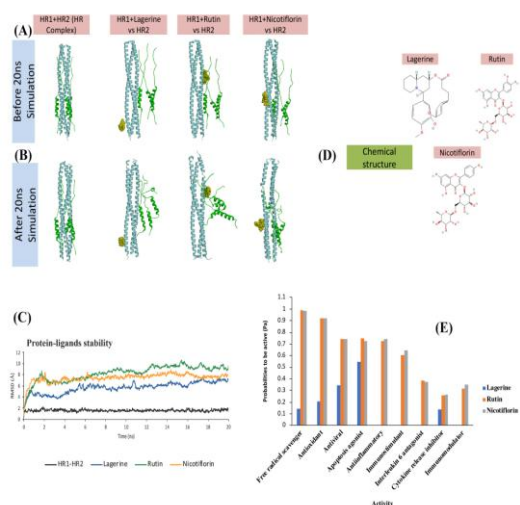


Fig. 1 Structural orientation and binding energy of the heptad repeat (HR) complex after being bound with phytochemical ligands (A) and structural alignment of HR before simulation (B) and after being bound with phytochemical ligands after simulation. The cyan ribbons in Fig. A represent the HR protein complex, and the yellow spheres describe the phytochemical ligand. (C) The stability of protein-ligand complex interactions. (D) Chemical structure of the ligand. (E) Probability to be active and activity of the ligands

Furthermore, the herbal compound was analyzed with Way2Drug Pass Server. Probability to be active (P_a) > 0.7 indicated that the compound is predicted to have a high potential as an immunomodulator. P_a > 0.3 and less than 0.7 indicates that the compound has the lowest potential as an immunomodulator. The results of the Way2Drug analysis have shown that compounds such as rutin (P_a 0.743) and nicotiflorin (P_a 0.742) have potential as antivirals (Fig. 2d), and lagerine (P_a 0.549) has potential as an apoptosis agonist. The highest activity is with free radical scavengers with a P_a value of 0.988 for rutin and 0.984 for nicotiflorin and antioxidants (0.923, 0.924). Stress conditions caused by lipid oxidation will be responded to in the body with an inflammatory mechanism. If stress occurs continuously, it would be chronic inflammation. The role of herbal medicines as anti-oxidative stress is supported by P_a values as those of antioxidants and free radical scavengers. Cytokine storms are inflammation that occurs through the deregulation of the immune response, leading to the disturbance of tissue homeostasis and severe organ damage [25, 26]. Herbal medicines such as *E. hirta* and *K. hospita* are potential anti-inflammatory drugs with P_a values of 0.728 and 0.743. From another perspective, immunostimulants enhance the immune system to suppress inflammation [27]. Rutin and nicotiflorin have immunostimulant activity (with P_a values of 0.607 and 0.642).

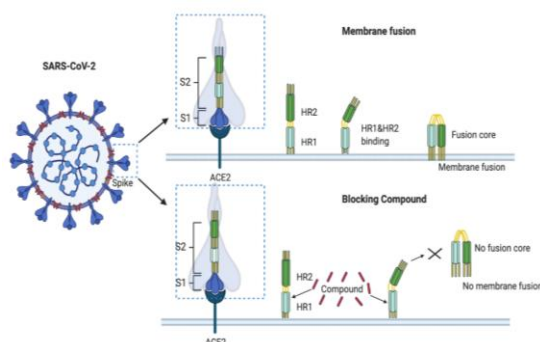


Fig. 2 Fusion core formation mechanism and blocking effect of an antiviral. The S1 subunit contains one receptor-binding domain (RBD). The entry of the virus is mediated by the S2 subunit for the virus/cell membrane fusion. The RBD of the S1 subunit binds to the host angiotensin-converting enzyme 2 receptor when the SARS-CoV-2 is in contact with the cell membrane. The fusion of S2 is exposed and inserted into the target cell membrane. HR1s and HR2s combine to form the fusion core, pulling the viral membrane to fuse with the host cell membrane. The designed and computationally optimized antiviral can bind to HR1 more tightly, preventing the HR1s and HR2s from forming the fusion core

4. Conclusion

Three compounds from *Lagerstroemia*, *E. hirta*, and *K. hospita* have good treatment potential based on binding energy with spike protein HR. Lagerine, rutin, and nicotiflorin compounds showed potential to prevent viral–host fusion by modifying the HR complex structure. In addition, Rutin and nicotiflorin are good immunomodulators. The results of this study

can be used as pre-experimental evidence to support the use of herbal medicines against COVID-19.

Acknowledgment

The authors would like to thank the Rector of Brawijaya University for providing a grant to conduct this research under scheme HGB.

References

- [1] NUGRAHA R. V., RIDWANSYAH H., GHOZALI M., KHAIRANI A. F., and ATIK N. Traditional Herbal Medicine Candidates as Complementary Treatments for COVID-19: A Review of Their Mechanisms, Pros and Cons. *Evidence-Based Complementary and Alternative Medicine*, 2020, 2020: e2560645. <https://doi.org/10.1155/2020/2560645>
- [2] PANYOD S., HO C.T., and SHEEN L.Y. Dietary Therapy and Herbal Medicine for COVID-19 Prevention: A Review and Perspective. *Journal of Traditional and Complementary Medicine*, 2020, 10(4): 420–427. <https://doi.org/10.1016/j.jtcme.2020.05.004>
- [3] DI MATTEO G., SPANO M., GROSSO M., SALVO A., INGALLINA C., RUSSO M., RITIENI A., and MANNINA L. Food and COVID-19: Preventive/Co-Therapeutic Strategies Explored by Current Clinical Trials and in Silico Studies. *Foods*, 2020, 9(8): E1036. <https://doi.org/10.3390/foods9081036>
- [4] JHA A. K., KUMAR R., GOENKA M. K., and DAYAL V. M. Emerging Treatment and Prevention Strategies against COVID-19: A Brief Update. *Journal of Digestive Endoscopy*, 2020, 11(1): 69–72. <https://doi.org/10.1055/s-0040-1712547>
- [5] MCKEE D. L., STERNBERG A., STANGE U., LAUFER S., and NAUJOKAT C. Candidate Drugs against SARS-CoV-2 and COVID-19. *Pharmacological Research*, 2020, 157: 104859. <https://doi.org/10.1016/j.phrs.2020.104859>
- [6] DAI W., ZHANG B., JIANG X. M., SU H., LI J., ZHAO Y., XIE X., JIN Z., PENG J., LIU F., LI C., LI Y., BAI F., WANG H., CHENG X., CEN X., HU S., YANG X., WANG J., LIU X., XIAO G., JIANG H., RAO Z., ZHANG L. K., XU Y., YANG H., and LIU H. Structure-Based Design of Antiviral Drug Candidates Targeting the SARS-CoV-2 Main Protease. *Science*, 2020, 368(6497): 1331–1335. <https://doi.org/10.1126/science.abb4489>
- [7] XIA S., LIU M., WANG C., XU W., LAN Q., FENG S., QI F., BAO L., DU L., LIU S., QIN C., SUN F., SHI Z., ZHU Y., JIANG S., and LU L. Inhibition of SARS-CoV-2 (Previously 2019-NCoV) Infection by a Highly Potent Pan-Coronavirus Fusion Inhibitor Targeting Its Spike Protein That Harbors a High Capacity to Mediate Membrane Fusion. *Cell Research*, 2020, 30(4): 343–355. <https://doi.org/10.1038/s41422-020-0305-x>
- [8] ZHANG L., LIN D., SUN X., CURTH U., DROSTEN C., SAUERHERING L., BECKER S., ROX K., and HILGENFELD R. Crystal Structure of SARS-CoV-2 Main Protease Provides a Basis for Design of Improved α -Ketoamide Inhibitors. *Science*, 2020, 368(6489): 409–412. <https://doi.org/10.1126/science.abb3405>
- [9] XIA S., LIU M., WANG C., XU W., LAN Q., FENG S., QI F., BAO L., DU L., LIU S., QIN C., SUN F., SHI Z., ZHU Y., JIANG S., and LU L. Inhibition of SARS-CoV-2

- (Previously 2019-NCoV) Infection by a Highly Potent Pan-Coronavirus Fusion Inhibitor Targeting Its Spike Protein That Harbors a High Capacity to Mediate Membrane Fusion. *Cell Research*, 2020, 30(4): 343–355. <https://doi.org/10.1038/s41422-020-0305-x>
- [10] WALLS A. C., PARK Y. J., TORTORICI M. A., WALL A., MCGUIRE A. T., and VEESLER D. Structure, Function, and Antigenicity of the SARS-CoV-2 Spike Glycoprotein. *Cell*, 2020, 181(2): 281–292.e6. <https://doi.org/10.1016/j.cell.2020.02.058>
- [11] HARVEY A. L., EDRADA-EBEL R., and QUINN R. J. The Re-Emergence of Natural Products for Drug Discovery in the Genomics Era. *Nature Reviews Drug Discovery*, 2015, 14(2): 111–129. <https://doi.org/10.1038/nrd4510>
- [12] YANG Y., ISLAM M. S., WANG J., LI Y., and CHEN X. Traditional Chinese Medicine in the Treatment of Patients Infected with 2019-New Coronavirus (SARS-CoV-2): A Review and Perspective. *International Journal of Biological Sciences*, 2020, 16(10): 1708–1717. <https://doi.org/10.7150/ijbs.45538>
- [13] SHAHRAJABIAN M. H., SUN W., SHEN H., and CHENG Q. Chinese Herbal Medicine for SARS and SARS-CoV-2 Treatment and Prevention, Encouraging Using Herbal Medicine for COVID-19 Outbreak. *Acta Agriculturae Scandinavica, Section B — Soil & Plant Science*, 2020, 70(5): 437–443. <https://doi.org/10.1080/09064710.2020.1763448>
- [14] RUSMANA D., WAHYUDIANINGSIH R., ELISABETH M., BALQIS B., MAESAROH M., and WIDOWATI W. Antioxidant Activity of Phyllanthus Niruri Extract, Rutin and Quercetin. *The Indonesian Biomedical Journal*, 2017, 9(2): 84–90. <https://doi.org/10.18585/inabj.v9i2.281>
- [15] FRANCENIA SANTOS-SÁNCHEZ N., SALAS-CORONADO R., VILLANUEVA-CAÑONGO C., and HERNÁNDEZ-CARLOS B. Antioxidant Compounds and Their Antioxidant Mechanism. In: SHALABY E. (ed.) *Antioxidants*. IntechOpen, London, 2019. <https://doi.org/10.5772/intechopen.85270>
- [16] KHAN M. R., HUANG C., ZHAO H., HUANG H., REN L., FAIQ M., HASHMI M. S., LI B., ZHENG D., XU Y., SU H., and AN J. Antioxidant activity of thymol essential oil and inhibition of polyphenol oxidase enzyme: A case study on the enzymatic browning of harvested longan fruit. *Chemical and Biological Technologies in Agriculture*, 2021, 8(1): 61. <https://doi.org/10.1186/s40538-021-00259-y>
- [17] DALLAKYAN S., and OLSON A. J. Small-Molecule Library Screening by Docking with PyRx. In: HEMPEL J., WILLIAMS C., and HONG C. (eds.) *Chemical Biology. Methods in Molecular Biology*, Vol. 1263. Humana Press, New York, 2015: 243–250. https://doi.org/10.1007/978-1-4939-2269-7_19
- [18] JAGHOORI M. M., LEIJLEVENS B., and OLABARRIAGA S. D. 1001 Ways to run AutoDock Vina for virtual screening. *Journal of Computer-Aided Molecular Design*, 2016, 30(3): 237–249. <https://doi.org/10.1007/s10822-016-9900-9>
- [19] VISHNUVARTHAN V. J. In-Silico Screening of Flavonoids targeted for Death Receptors in Cancer by Using Hex Molecular Docking. *Journal of Young Pharmacists*, 2017, 9(2): 168–171. <https://doi.org/10.5530/jyp.2017.9.33>
- [20] KRIEGER E., and VRIEND G. New Ways to Boost Molecular Dynamics Simulations. *Journal of Computational Chemistry*, 2015, 36(13): 996–1007.

<https://doi.org/10.1002/jcc.23899>

[21] GHOSH R., CHAKRABORTY A., BISWAS A., and CHOWDHURI S. Identification of Polyphenols from *Broussonetia Papyrifera* as SARS CoV-2 Main Protease Inhibitors Using in Silico Docking and Molecular Dynamics Simulation Approaches. *Journal of Biomolecular Structure and Dynamics*, 2021, 39(17): 6747–6760. <https://doi.org/10.1080/07391102.2020.1802347>

[22] MARTÍNEZ L. Automatic Identification of Mobile and Rigid Substructures in Molecular Dynamics Simulations and Fractional Structural Fluctuation Analysis. *PLoS ONE*, 2015, 10(3): e0119264. <https://doi.org/10.1371/journal.pone.0119264>

[23] XIA S., ZHU Y., LIU M., LAN Q., XU W., WU Y., YING T., LIU S., SHI Z., JIANG S., and LU L. Fusion mechanism of 2019-nCoV and fusion inhibitors targeting HR1 domain in spike protein. *Cellular & Molecular Immunology*, 2020, 17(7): 765–767. <https://doi.org/10.1038/s41423-020-0374-2>

[24] OU X., LIU Y., LEI X., LI P., MI D., REN L., GUO L., GUO R., CHEN T., HU J., XIANG Z., MU Z., CHEN X., CHEN J., HU K., JIN Q., WANG J., and QIAN Z. Characterization of Spike Glycoprotein of SARS-CoV-2 on Virus Entry and Its Immune Cross-Reactivity with SARS-CoV. *Nature Communications*, 2020, 11(1): 1620. <https://doi.org/10.1038/s41467-020-15562-9>

[25] RAGAB D., SALAH ELDIN H., TAEIMAH M., KHATTAB R., and SALEM R. The COVID-19 Cytokine Storm; What We Know So Far. *Frontiers in Immunology*, 2020, 11: 1446. <https://doi.org/10.3389/fimmu.2020.01446>

[26] SOY M., KESER G., ATAGÜNDÜZ P., TABAK F., ATAGÜNDÜZ I., and KAYHAN S. Cytokine Storm in COVID-19: Pathogenesis and Overview of Anti-Inflammatory Agents Used in Treatment. *Clinical Rheumatology*, 2020, 39(7): 2085–2094. <https://doi.org/10.1007/s10067-020-05190-5>

[27] VASEEHARAN B., and THAYA R. Medicinal Plant Derivatives as Immunostimulants: An Alternative to Chemotherapeutics and Antibiotics in Aquaculture. *Aquaculture International*, 2014, 22: 1079–1091. <https://doi.org/10.1007/s10499-013-9729-3>

[28] WORLD HEALTH ORGANIZATION. *WHO Coronavirus (COVID-19) Dashboard*, n.d. <https://covid19.who.int/>

[29] PROTEIN DATA BANK. n.d. <https://www.rcsb.org/>

[30] "KNAPSACK" FAMILY. n.d. <http://www.knapsackfamily.com/>

参考文献:

[1] NUGRAHA R. V., RIDWANSYAH H., GHOZALI M., KHAIRI A. F. 和 ATIK N. 传统草药候选药物作为 COVID-19 的补充治疗：对其机制、优点和缺点的回顾。循证补充和替代医学，2020 年，2020 年：e2560645. <https://doi.org/10.1155/2020/2560645>

[2] PANYOD S., HO C.T. 和 SHEEN L.Y. 预防新冠肺炎的膳食疗法和草药：回顾和展望。传统与补充医学杂志，2020，10(4): 420–427. <https://doi.org/10.1016/j.jtcm.2020.05.004>

[3] DI MATTEO G., SPANO M., GROSSO M., SALVO A., INGALLINA C., RUSSO M., RITIENI A. 和 MANNINA L. 食品和新冠肺炎：预防/联合治疗策略探索

当前的临床试验和计算机研究。食品，2020，9(8): E1036。 <https://doi.org/10.3390/foods9081036>

[4] JHA A. K., KUMAR R., GOENKA M. K. 和 DAYAL V. M. 针对新冠肺炎 的新兴治疗和预防策略：简要更新。消化内镜杂志，2020，11(1): 69-72. <https://doi.org/10.1055/s-0040-1712547>

[5] MCKEE D. L., STERNBERG A., STANGE U., LAUFER S. 和 NAUJOKAT C. 针对非典-冠状病毒-2 和新冠肺炎的候选药物。药理研究，2020，157: 104859. <https://doi.org/10.1016/j.phrs.2020.104859>

[6] 戴伟，张斌，蒋晓明，苏海，李杰，赵毅，谢旭，金志，彭杰，刘菲，李春，李毅，白 F., 王 H., 程 X., 岑 X., 胡 S., 杨 X., 王 J., 刘 X., 肖 G., 蒋 H., 饶 Z., 张 L.K., XU Y., YANG H. 和 LIU H. 基于结构的针对非典-冠状病毒-2 主要蛋白酶的抗病毒候选药物的设计。科学，2020 年，368 (6497) : 1331–1335。 <https://doi.org/10.1126/science.abb4489>

[7] 夏 S., 刘敏., 王 C., 徐 W., 兰琪., 冯 S., 齐峰., 鲍丽., 杜丽., 刘 S., 秦 C., 孙 F., SHI Z., ZHU Y., JIANG S. 和 LU L. 抑制非典-冠状病毒-2 (以前是 2019-新型冠状病毒) 通过高度有效的泛环病毒融合抑制剂靶向其峰值蛋白，可容纳高能力介导膜融合。细胞研究，2020，30(4): 343–355. <https://doi.org/10.1038/s41422-020-0305-x>

[8] ZHANG L., LIN D., SUN X., CURTH U., DROSTEN C., SAUERHERING L., BECKER S., ROX K. 和 HILGENFELD R. 非典-冠状病毒-2 主要蛋白酶的晶体结构提供设计改进的 α -酮酰胺抑制剂的基础。科学，2020 年，368 (6489) : 409–412。 <https://doi.org/10.1126/science.abb3405>

[9] 夏 S., 刘敏., 王 C., 徐 W., 兰琪., 冯 S., 齐峰., 鲍丽., 杜丽., 刘 S., 秦 C., 孙 F., SHI Z., ZHU Y., JIANG S. 和 LU L. 抑制非典-冠状病毒-2 (以前是 2019-新型冠状病毒) 通过高度有效的泛环病毒融合抑制剂靶向其峰值蛋白，可容纳高能力介导膜融合。细胞研究，2020，30(4): 343–355. <https://doi.org/10.1038/s41422-020-0305-x>

[10] WALLS A. C., PARK Y. J., TORTORICI M. A., WALL A., MCGUIRE A. T. 和 VEESLER D. 非典-冠状病毒-2 刺突糖蛋白的结构、功能和抗原性。细胞，2020，181(2): 281–292.e6。 <https://doi.org/10.1016/j.cell.2020.02.058>

[11] HARVEY A. L., EDRADA-EBEL R. 和 QUINN R. J. 在基因组学时代重新出现用于药物发现的天然产物。自然评论药物发现，2015，14 (2) : 111–129。 <https://doi.org/10.1038/nrd4510>

[12] YANG Y., ISLAM M. S., WANG J., LI Y., 和 CHEN X. 中医药治疗 2019 新型冠状病毒(非典-冠状病毒-2)感染患者的回顾与展望。国际生物科学杂志，2020，16(10): 1708–1717. <https://doi.org/10.7150/ijbs.45538>

[13] SHAHRAJABIAN M. H., SUN W., SHEN H. 和 CHENG Q. 非典和非典-冠状病毒-2 治疗和预防的中草药，鼓励使用中草药治疗新冠肺炎爆发。斯堪的纳维亚农业学报，乙节 — 土壤与植物科学，2020 年，70(5) : 437–443. <https://doi.org/10.1080/09064710.2020.1763448>

[14] RUSMANA D., WAHYUDIANSIH R., ELISABETH M., BALQIS B., MAESAROH M., 和 WIDOWATI W. 叶下珠提取物、芦丁和槲皮素的抗氧化

- 活性。印度尼西亚生物医学杂志, 2017 年, 9(2): 84-90。
<https://doi.org/10.18585/inabj.v9i2.281>
- [15] FRANCENIA SANTOS-SÁNCHEZ N., SALAS-CORONADO R., VILLANUEVA-CAÑONGO C. 和 HERNÁNDEZ-CARLOS B. 抗氧化剂化合物及其抗氧化机制。在: SHALABY E. (编辑) 抗氧化剂。英泰克打开, 伦敦, 2019 年。
<https://doi.org/10.5772/intechopen.85270>
- [16] KHAN M. R., HUANG C., ZHAO H., HUANG H., REN L., FAIQ M., HASHMI M. S., LI B., ZHENG D., XU Y., SU H., 和 AN J. 抗氧化剂百里酚精油的活性和多酚氧化酶的抑制作用: 以龙眼果实酶促褐变为例。农业化学和生物技术, 2021, 8(1): 61. <https://doi.org/10.1186/s40538-021-00259-y>
- [17] DALLAKYAN S. 和 OLSON A. J. 通过与派接收对接进行的小分子文库筛选。在: HEMPEL J., WILLIAMS C. 和 HONG C. (编辑) 化学生物学。分子生物学方法, 卷。1263. 人道出版社, 纽约, 2015: 243-250. https://doi.org/10.1007/978-1-4939-2269-7_19
- [18] JAGHOORI M. M., LEIJLEVENS B. 和 OLABARRIAGA S. D. 1001 种运行自动停靠维娜进行虚拟筛选的方法。计算机辅助分子设计杂志, 2016, 30(3): 237-249. <https://doi.org/10.1007/s10822-016-9900-9>
- [19] VISHNUVARTHAN V. J. 通过使用十六进制分子对接对靶向癌症中死亡受体的类黄酮进行计算机筛选。青年药剂师杂志, 2017, 9(2): 168-171. <https://doi.org/10.5530/jyp.2017.9.33>
- [20] KRIEGER E. 和 VRIEND G. 促进分子动力学模拟的新方法。计算化学杂志, 2015, 36(13): 996-1007. <https://doi.org/10.1002/jcc.23899>
- [21] GHOSH R., CHAKRABORTY A., BISWAS A. 和 CHOWDHURI S. 鉴定纸莎草多酚作为非新冠病毒-2 主要蛋白酶抑制剂在计算机对接和分子动力学模拟方法中的应用。生物分子结构与动力学杂志, 2021, 39(17): 6747-6760. <https://doi.org/10.1080/07391102.2020.1802347>
- [22] MARTÍNEZ L. 分子动力学模拟和分数结构波动分析中移动和刚性子结构的自动识别。公共科学图书馆一期, 2015 年, 10(3): e0119264。
<https://doi.org/10.1371/journal.pone.0119264>
- [23] XIA S., ZHU Y., LIU M., LAN Q., XU W., WU Y., YING T., LIU S., SHI Z., JIANG S., 和 LU L. 2019 新型冠状病毒和针对刺突蛋白中人力资源 1 结构域的融合抑制剂。细胞与分子免疫学, 2020, 17(7): 765-767. <https://doi.org/10.1038/s41423-020-0374-2>
- [24] 欧 X., 刘 Y., 雷 X., 李 P., 米 D., 任 L., 郭 L., 郭 R., 陈 T., 胡 J., 向 Z., 穆 Z., 陈 X., 陈 J., 胡 K., 金 Q., 王 J., 和 钱 Z. 非典-冠状病毒-2 刺突糖蛋白对病毒进入的表征及其与非典-冠状病毒的免疫交叉反应。自然通讯, 2020, 11(1): 1620. <https://doi.org/10.1038/s41467-020-15562-9>
- [25] RAGAB D., SALAH ELDIN H., TAEIMAH M., KHATTAB R. 和 SALEM R. 新冠肺炎细胞因子风暴; 到目前为止我们所知道的。免疫学前沿, 2020 年, 11: 1446. <https://doi.org/10.3389/fimmu.2020.01446>
- [26] SOY M., KESER G., ATAGÜNDÜZ P., TABAK F., ATAGÜNDÜZ I. 和 KAYHAN S. 新冠肺炎中的细胞因子风暴: 治疗中使用的抗炎药的发病机制和概述。临床风湿病学, 2020, 39(7): 2085-2094。
<https://doi.org/10.1007/s10067-020-05190-5>
- [27] VASEEHARAN B. 和 THAYA R. 药用植物衍生物作为免疫刺激剂: 水产养殖中化疗药物和抗生素的替代品。国际水产养殖, 2014, 22: 1079-1091。
<https://doi.org/10.1007/s10499-013-9729-3>
- [28] 世界卫生组织。世卫组织冠状病毒(新冠肺炎)仪表板, 未注明日期。<https://covid19.who.int/>
- [29] 蛋白质数据库。未注明日期 <https://www.rcsb.org/>
- [30] “背包”家庭。未注明日期 <http://www.knapsackfamily.com/>