湖南大学学报(自然科学版) Journal of Hunan University (Natural Sciences)

Vol. 49 No. 3 March 2022

Open Access Article

https://doi.org/10.55463/issn.1674-2974.49.3.14

Libido and Sperm Quality of the Etawah Cross-Breed Fed Urea Moringa Molasses Multinutrient Block Supplement

Nursyam Andi Syarifuddin*, Muhammad Rizal, Muhammad Riyadhi, Anis Wahdi

Department of Animal Science, Faculty of Agriculture, University of Lambung Mangkurat, Banjarbaru, Indonesia

Abstract: Moringa leaves contain high and complete nutrients, so they can be used as a constituentfor multi-nutrient blocks to increase libido and sperm quality. This study aimed to evaluate the effect of Moringa leaf flour in a multi-nutrient block on libido and sperm quality of the Etawah Cross-Breed (PE) goats. This study used a two-sample t-test on 8 PE goats aged 18.50±1.00 months, body weight 32±1.49 kg, kept in individual pens for 8 weeks, given the swamp forage ad libitum. A total of 4 animals were supplemented with urea molasses multinutrient block (UMMB) as a control, and 4 animals were supplemented with urea moringa molasses multi-nutrient block (UMMMB) as a treatment. Libido and semen quality were measured from week 4 to week 8. Measurement of scrotal circumference and blood sampling were performed at weeks 0, 5, and 8. The concentration of testosterone in plasma was analyzed using the ELISA technique. The measured variables were compared using the independent sample t-test. UMMMB supplementation did not significantly (p>0.05) increase scrotal circumference and testosterone levels. UMMMB supplementation did not significantly (p>0.05) decrease reaction time, but significantly (p<0.05) decrease mount (24.20±4.30 vs 12.93±1.58 min) and ejaculation (25.60±4.11 vs 13.97±2.23 min). UMMMB supplementation did not significantly (p>0.05) increase semen volume and sperm concentration, but significantly (p<0.05) increased total sperm motility (69.67±0.76 vs 74.67±1.46%) and sperm viability (80.37±0.68 vs 86.48±1.74%). It can be concluded that UMMMB supplementation markedly increased libido, total motility, and sperm viability of PE goats.

Keywords: libido, sperm quality, moringa, multi-nutrient block, Etawah cross-breed.

伊塔瓦杂交种饲喂尿素辣木糖蜜多营养素块补充剂的性欲和精子质量

摘要:辣木叶含有丰富而完整的营养成分,因此可用作多营养块的成分,以提高性欲和精子质量。本研究旨在评估多营养块中辣木叶粉对伊塔瓦杂交(藻红蛋白)山羊性欲和精子质量的影响。本研究对 8 只 18.50±1.00 月龄、体重 32±1.49 公斤、在单独的围栏中饲养 8 周的藻红蛋白山羊进行了两样本 t 检验,并随意给予沼泽草料。共有 4 只动物补充了尿素糖蜜多营养块作为对照,4 只动物补充了尿素辣木糖蜜多营养块作为治疗。从第 4 周到第 8 周测量性欲和精液质量。在第 0、5 和 8 周进行阴囊周长测量和采血。使用酶联免疫吸附测定技术分析血浆中睾酮的浓度。使用独立样本 t 检验比较测量变量。辣木糖蜜多营养块补充没有显着(p>0.05)增加阴囊周长和睾酮水平。尿素辣木糖蜜多营养块补充没有显着(p>0.05)减少生骑(24.20±4.30 对比 12.93±1.58 分钟)和射精(25.60±4.11 对比 13.97±2.23 分钟))。尿素辣木糖蜜多营养块补充没有显着(p>0.05)增加精液量和精子浓度,但显着(p<0.05)增加总精子活力(69.67±0.76 对比 74.67±1.46%)和精子活力(80.37± 0.68 对 86.48±1.74%)。可以得出结论,尿素辣木糖蜜多营养块补充剂显着提高了藻红蛋白山羊的性欲、总活力和精子活力。

Received: 09 December, 2021 / Revised: 22 January, 2022 / Accepted: 09 February, 2022 / Published: 28 March, 2022 Fund Project: Research Contract No. 123.20/UN.8.2/PP/2019 dated March 15, 2019, Ministry of Research, Technology and Higher

Education, Indonesia

About the authors: Nursyam Andi Syarifuddin, Muhammad Rizal, Muhammad Riyadhi, Anis Wahdi, Department of Animal Science, Faculty of Agriculture, University of Lambung Mangkurat, Banjarbaru,

关键词:性欲、精子质量、**辣木、多**营养块、**伊塔瓦**杂交

1. Introduction

The feed technology multi-nutrient blocks have been developed for a long time and have been shown to increase the productivity and reproducibility of ruminants, especially goats. Multinutrient blocks have been shown to improve the production and reproductive performance of goats [1]. The multinutritional blocks can also be used to feed lactating goats to partially replace the concentrate [2]. Urea Molasses Multinutrient Block (UMMB) has been shown to improve milk composition and milk quality of Saanen crossbreed goats[3].

Moringa (Moringa oleifera Lam) is a tree species, has great potential to be used as feed, due to its high protein contentwhich ranges between 226.0 to 268.0 g kg⁻¹dry matter (DM) [4] and its biomass production can reach 2871.8 kgha⁻¹cut⁻¹ DM [5]. Moringa leaves have been used as feed to sperm fertility of rabbits [6] increasing libido and sperm quality in Bali bulls [7] and semen characteristics of buffalo [8]. The libido and semen quality are influenced among others by nutritional factors. Nutrition controls hormone testosterone secretion, sperm production, and may promote testicular development [9].

Moringa leaves contain high and complete nutrition. Moringa leaves contain amino acids, fatty acids, macrominerals, microminerals [10], and phytochemicals [11]. Moringa leaves contain the amino acid arginine essentials for the process of spermatogenesis, putrescine precursors, spermidine, and spermine synthesis which is important for sperm motility and the capacitation of spermto fertilize ova [12].

Moringa leaves contain minerals Zn and Se, which are important for the process of spermatogenesis. Zn minerals can function to stimulate Leydig cells in the testes to produce the hormone testosterone. Mineral Se functions as a strong antioxidant combined with amino acids to form seleno-proteins and enzymes to form selenoenzymes affecting sperm quality by preventing oxidative damage.

Moringa leaves contain lipid (DHA) as a major component in sperm tail, important in sperm motility, and fertility capacity. Moringa leaves contain vitamins B12, B9, A, E, and C which play an important role in the process of spermatogenesis[13].

Therefore, this study will use Moringa leaf flour as a constituent of multi-nutrient block supplement feed in the form of *Urea Moringa Molasses Multinutrient Block* (UMMMB) to increase the libidos and sperm qualities of PE goats.

2. Materials and Methods

2.1. Animal

This study used eight healthy PE goats aged 18.50 ± 1.00 month, bodyweight 32 ± 1.49 kg obtained from Breeding Center Pelaihari. The goats were kept in individual pens measuring $1.25 \text{ m} \times 1.00 \text{ m}$ in the form of a stage with a height of 1 m from the ground.

2.2. Experimental Design

The PE goats were divided into two groups, the first group of four goats was supplemented with UMMB as a control and the second group of four goats was supplemented with UMMMB as a treatment. The goats were given the main feed of swamp forage a mixture of *Polygonum barbatum L* and *Ischaemum polystachyum*. This study used *the independent sample t-test* to compare each observed variable.

Scrotal circumference measurements were carried out at 0, 5, and 8 weeks to know the growth of the scrotal circumference. Measurement of scrotal circumference using a measuring tape with a scale of 1 mm according to the procedures of [14]. Blood sampling in the jugular vein using a vacutainer tube at 0, 5, and 8 weeks to determine the increase in testosterone hormone levels. The plasma sample was stored at -20C until the hormonal assay. Testosterone was analyzeby using the ELISA technique in accordance procedure of [15]. Measurements of libido and semen quality were carried out once a week for five weeks from the fourth to the eighth week. Measurement of libido and semen collection was carried out at 07.00 - 10.00 WITA before feeding. Libido observation was begun from the first time to sniff the teaser (reaction time), to mount (mount), and finally to ejaculate in an artificial vagina (ejaculation). Semen collection and semen quality assessment macroscopically and microscopically according to [16].

2.3. Diet and Feeding

The PE goats kept for eight weeks were given rations according to the nutritional requirements for goats [17]. Four goats were given the main feed of swamp forage plus one block of UMMB weighing 200g, and four goats were given the main feed of swamp forage plus one block of UMMMB weighing 200g. Supplementary feed was given in the morning before forage feeding. Forage feeding is carried out after the supplement feed runs out. Provision of forage and drinking water on an *ad-lib* basis. The composition of feed supplements for UMMB and UMMMB are

shown in Table 1. The nutritional content of swamp forage, UMMB, UMMMB, used is shown in Table 2.

Table 1.Compositions of the multi-nutrient block

No	Feed ingredients (%)	UMMB	UMMMB
1.	Moringa Leaf Flour	-	30
2.	Oil Palm meal	25	-
3.	Concentrate Laying Ducks (CP144®)	5	-
4.	Fine rice bran	33	33
5.	Molasses	20	20
6.	Lime	5	5
7.	Salt	5	5
8.	Urea	5	5
9.	Mineral Mix	2	2
	Total	100	100

Table 2. Nutrient content of swamp forage, UMMB, and UMMMB (as % DM)

No	Nutrient composition (%)	Swamp Forage	UMM B	UMM MB
1.	Dry matter	28.86	69.32	66.68
2.	Ash	16.39	15.24	14.14
3.	Crude Protein	13.23	19.24	21.27
4.	Crude Fiber	22.41	6.45	7.72
5.	Ether Extract	1.22	1.95	2.45
6.	Ca	0.28	3.68	4.21
7.	P	0.34	2.15	2.72

	Continuation of Table 2							
8.	Nitrogen Free Extract	46.75	57.12	54.42				
9.	TDN	55.33	67.78	69.82				

Note: Analyzed by Laboratory of Nutrition and Animal Feedstuff, Faculty of Agriculture, Animal Science Department, Lambung Mangkurat University.

2.4. Statistical Analysis

Data (increased scrotal circumference, level testosterone, libido, and semen quality) were reported as the mean and standard error of mean analyzed using *theindependentsample t-test* with SPSS® Version 22 Software.

3. Results

3.1. Dry Matter Consumption, Nutrients Consumption, and Average Daily Gain

Table 3 shows that the consumption of dry matter, crude protein, TDN, and the average daily gain of the PE goats, supplemented with UMMB and UMMMB, were no significant differences (p>0.05). However, the Ca and P minerals consumption was significantly higher (p<0.05) in the PE goats supplemented with UMMMB compared to supplemented with the UMMB.

Table 3 The consumption of dry matter and nutrients, average daily gain, of the PE goats supplemented with UMMB and UMMMB (g/day)

No	Parameters	UMMB	UMMMB	Requirement [17]
1	Dry matter	1.338 ± 0.01	1.328 ± 0.03	810
2	Crude protein	186.42 ± 1.86	188.32 ± 3.48	63
3	TDN	654.25 ± 7.83	648.50 ± 14.55	448
4	Ca	0.009 ± 0.00^{a}	0.010 ± 0.00^{b}	0.002
5	P	0.007 ± 0.00^{a}	0.008 ± 0.00^{b}	0.001
6	Average daily gain	$0,066 \pm 0,06$	$0,074 \pm 0,08$	-

Note: Means in the same row with different superscripts differ significantly (p<0.05).

3.2. The Scrotal Circumference, Testosterone Level, and Libido

Table 4 shows that increased scrotal circumference, testosterone levels, and reactions time of the PE goats were supplemented with UMMB and UMMMB were

no significant differences(p>0.05). However, the time to mount and to ejaculation was significantly faster (p<0.05) for PE goats supplemented with UMMMB compared to PE goats supplemented with UMMB.

Table 4 The increased scrotal circumference, level testosterone, and libido of the PE goatssupplemented with UMMB and MMMB

rarameters -			<i>p-</i> Value
Parameters –	Means (Means (± SEM)	
ed scrotal circumference (cm)	0.25 ± 0.22	0.48 ± 0.14	0.51
Testosterone (ng/ml):			
a. 0 weeks	0.74 ± 0.05	1.00 ± 0.02	0.10
b. 5 weeks	0.76 ± 0.13	2.51 ± 0.97	0.15
c. 8 weeks	6.58 ± 2.53	6.71 ± 3.93	0.98
Libido:			
a. Reaction time (min)	12.45 ± 3.59	6.38 ± 1.21	0.15
b.Mount (min)	24.20 ± 4.30^{a}	12.93 ± 1.58^{b}	0.04
c.Ejaculation (min)	25.60 ± 4.11^{a}	13.97±2.23 ^b	0.04
	ed scrotal circumference (cm) Testosterone (ng/ml): a. 0 weeks b. 5 weeks c. 8 weeks Libido: a. Reaction time (min) b.Mount (min)	Means (ed scrotal circumference (cm) 0.25 ± 0.22 Testosterone (ng/ml): a. 0 weeks 0.74 ± 0.05 b. 5 weeks 0.76 ± 0.13 c. 8 weeks 6.58 ± 2.53 Libido: a. Reaction time (min) 12.45 ± 3.59 b. Mount (min) 24.20 ± 4.30a c. Ejaculation (min) 25.60 ± 4.11a	Means (\pm SEM) ed scrotal circumference (cm) 0.25 ± 0.22 0.48 ± 0.14 Testosterone (ng/ml):

Note: Means in the same row with different superscripts differ significantly (p<0.05).

Fig. 1 shows the growth of scrotal circumference during rearing. The average scrotal circumference at week 0 was 23.05 vs 23.13 cm and after being reared

for eight weeks the average scrotal circumference was 23.30 vs 23.60 cm, so obtained the addition of 0.25 vs 0.48 cm.

The testosterone level in PE goats also increased along with the time of rearing and feeding with multinutrient block supplements. Testosterone levels of PE goats fed UMMMB supplementation were higher than UMMB. The increase in testosterone levels is shown in Fig. 2.

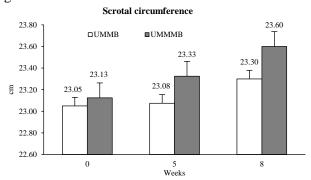


Fig. 1 Growth scrotal circumference of the PE goats

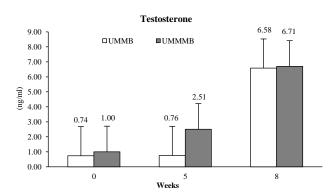


Fig. 2 Increased testosterone levels of PE goats

Comparison of the time required by the PE goats fed UMMB and UMMMB supplements to sniff a teaser, to mount, and ejaculate in an artificial vagina at 4 to 8 weeks are shown in Fig. 3.

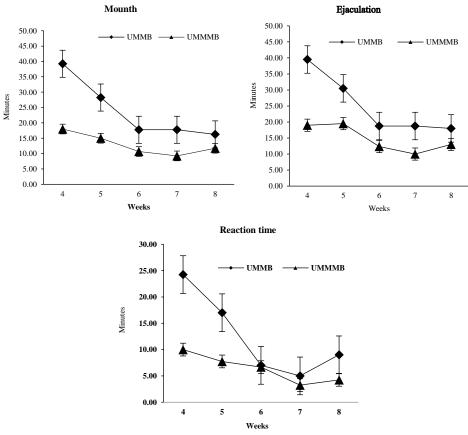


Fig. 3. Measurement of reaction time, mount, and ejaculation of PE goats

3.3. Semen Quality

Table 5 shows that semen volume, concentration, and the sperm abnormalities of PE goats supplemented with UMMB and UMMMB were no significant

differences (p>0.05). However, the total motility and viability of sperm were higher (p<0.05) in PE goats supplemented with UMMMB compared to the control goat supplemented with UMMB.

Table 5 The quality of the fresh semen of the PE goats supplemented with UMMB and UMMMB

No	Parameters –	UMMB	UMMMB	n Volue	D.f
NO		Means (± SEM)		<i>p</i> -Value	References [18]
1.	Volume (ml)	0.83 ± 0.06	0.86 ± 0.07	0.75	0.5 - 1.2
2.	Concentration (million/ml)	2.708 ± 293.73	3.258 ± 152.31	0.14	2,500 - 5,000
3.	Total motility (%)	69.67 ± 0.76^{a}	74.67 ± 1.46^{b}	0.02	$70 - 90^{1}$
4.	Viability (%)	80.37 ± 0.68^{a}	86.48 ± 1.74^{b}	0.00	60 - 80 ²

Note: Means in the same row with different superscripts differ significantly (p<0.05)

A comparison of the semen quality of PE goats fed UMMB and UMMMB supplements during rearing at 5 to 8 weeks is shown in Fig. 4.

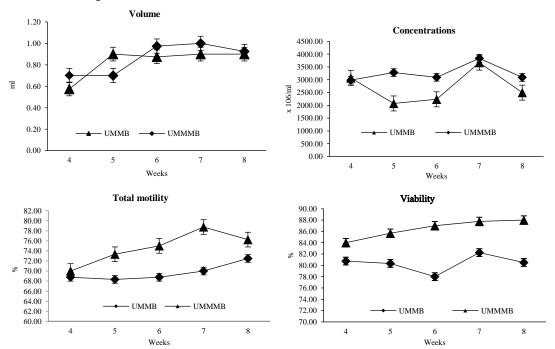


Fig. 4 Semen quality measurement results

4. Discussion

4.1. The Scrotal Circumference, Testosterone Level, and Libido

The results of this study showed that the supplementation of UMMMB significantly increased the libido of PE goats. The time required the goats to mount the teaser and ejaculate in the artificial vagina was significantly shorter than the control. However, all other reproductive variables observed such as scrotal circumference, testosterone concentration, also increased and reaction time decreased but did not show a significant difference.

UMMMB supplementation did not significantly increase the scrotal circumference of the experimental goats, presumably, the goats were more than 18 months old, i.e. an average of 18.50±1.00 months. The study by [19] on Assam goats showed that the morphometrics of seminiferous tubules and Leydig cells experienced a significant increase at the age of 6-8 months. At the age of eight months and over, the scrotal circumference will be constant (between 17-18 cm) without further significant increase. This condition probably caused the use of Moringa leaves on UMMMB did not significantly affect the increase in the scrotal circumference of the experimental goats used.

The experimental goat scrotal circumference obtained was smaller than the standard Breeding Soundness Examination in the buck [14]. It was explained that the scrotal circumference was at least 25

cm for a breed that weighed 45 kg, and most dairy goat males had a scrotal circumference of 25-28 cm when the bodyweight reached 45 kg. The experimental goat used was only 32 ± 1.49 kg, so the scrotum circumference obtained was also small (Fig. 1), because scrotal circumference was positively correlated with body weight [20]. However, the scrotal circumference obtained was still higher than the three indigenous breeds of goats in arid and semiarid agroecology of Ethiopia: Afar, Long-Eared Somali, and Woyto-Guji each; 20.5 ± 2.10 cm; 21.4 ± 1.67 cm; 20.6 ± 1.93 cm; or an average of 20.8 ± 1.94 cm [20].

Testosterone hormone in experimental goats increased along with an increase in scrotal circumference. This is because the hormone testosterone is formed in the Leydig cells of the testes. Scrotal circumference increases due to an increase in the number of Leydig cells. Leydig cells are still developing, so the production of the hormone testosterone continues to increase. The development of testosterone hormone production described by[21] it was starting from conception and the fetus is the peak producing testosterone is differentiation is needed for male sex formation and masculinization in the brain. Testosterone production then declines to a nadir in the early postpartum period. After that, the production of the hormone testosterone then increases gradually through puberty to adulthood. Furthermore, testosterone production decreases due to aging and other conditions that result in decreased testosterone levels (hypogonadism) that accompany changes in metabolism and quality of life.

Fig. 2 illustrates the increase in testosterone levels of experimental goats during maintenance and the highest at the eighth week after rearing. The testosterone level of experimental goats in the treatment group was higher than the control group but did not show a significant difference. The study by [22] showed that feed factors affect testosterone levels in male goats. The nutritional status of livestock has an impact on the growth performance and development of reproductive organs.Testicular mass. characteristics, spermatogenesis, and reproductive hormones can be influenced by feeding levels[23]. The testosterone level of experimental goats in the treatment group was higher than the control, possibly due to the effect of higher crude protein consumption, although it did not show a significant difference (Table 3) and the other possibility of Moringa leaves containing high Zn minerals in UMMMB. Moringa leaves contain the mineral Zn 25.5 - 31.03 mg/kg[10]. Zn mineralsstimulate Leydig cells to produce the normal function testosterone for hypothalamus-pituitary-testesaxis. Higher testosterone levels in the treatment group, similar to the results of our previous study in Bali cattle [7]. Diurnal changes in testosterone levels in Balinese cattle supplemented with Moringa leaves were significantly higher in the morning, afternoon, and evening. Therefore, the high protein content in UMMMB and the high Zn mineral content in Moringa leaves as a constituent of UMMMB are considered as possible triggers so that the testosterone levels of males supplemented with UMMMB are higher than the control, although it has not shown a significant difference.

Testosterone levels in experimental goats after being fed a multi-nutrient block supplement for 8 weeks were lower than those of [24] in Kacang goats, namely 18.51 ± 19.4618 ng/ml, and 29.57 ± 12.96 ng/ml. Factors affecting animal testosterone levels in the blood are a nation of origin, age, environment, disease, presence or absence of sexual stimulation, and the sensitivity of the methods [24].

Fig. 3 illustrates the better libido of PE goats during rearing fed UMMMB compared to controls. The time required to sniff a teaser, mount, and ejaculate in an artificial vagina was shorter than the control, which was 12.45 vs 6.38 min, 24.20 vs 12.93 min, and 25.60 vs 13.97 min, respectively. The time required to mount and ejaculate in an artificial vagina was significantly shorter, so the libido of PE goats supplemented with UMMMB was significantly better. Libido is influenced by testosterone levels [25] so the increase in libido in PE goats supplemented with UMMMB in this study was caused by an increase in testosterone levels. Thus, UMMMB supplementation can increase testosterone levels so that it can increase the libido of PE goats.

4.2. Semen Quality

The results of this study showed that UMMMB supplementation improved the quality of fresh semen of PE goats. Fig. 4 illustrates the semen volume, sperm concentration, total motility, and sperm viability of experimental goats supplemented with UMMMB during rearing better than controls. UMMMB supplementation significantly increased the total motility and viability of PE goats spermatozoa.

Semen volume, sperm concentration, total motility, viability, and abnormality of sperm obtained were all normal according to [18]. However, supplementation with UMMMB did not significantly increase the volume and concentration of semen. This condition is in line with the results of a study of our previous study, Moringa leaf supplementation did not significantly increase the volume and concentration of semen in Bali cattle [7].

The sperm concentration obtained in this study was higher than the results of research by [24] in Kacang goat, namely $2,763.0\pm395.0 \times 10^6$. This condition accordance with the statement of [26] and[27] that various factors are known to influence the goat semen quality such as age, breed, season, method of semen collection, extender, and centrifugation, frequency of ejaculation, techniques of breeding and even by variation among individual goats within the same herd.

UMMMB supplementation significantly increased the total motility and viability of PE goats' sperm. This can strengthen the notion that Moringa leaves can significantly increase the total motility and viability of PE goat sperm. The results of our previous study also showedthat Moringa leaf supplementation significantly increased the total sperm motility of Bali bulls [7]. Moringa leaves can increase total motility and viability possibly because Moringa leaves contain high Ca and P. Moringa leaves contain 2,003 mg/100 g Ca and 204 mg/100 g P [10]. As in the Bali cattle study, the results of this study showed that UMMMB supplementation significantly increased Ca and P consumption, so that the high Ca and P content in Moringa leaves could be suspected as the cause of significantly higher sperm motility of goats supplemented with UMMMB.As well [28] explained that Ca serves to increase sperm motility. Phosphorus plays an important role in fertility because it plays a role in energy (ATP) transfer processes and as a second messenger (cAMP). Phosphorus deficiency in males can cause testicular degeneration. cAMP is related to sperm motility and capacitation. In addition to the, Ca and P content, the high Zn content in Moringa leaves can also be expected to increase the total motility and sperm viability of PE goats in this study. The results of a review of [29] showed that high Zn concentrations were associated with high sperm motility and viability. Zinc in the testes is essential for spermatogenesis and spermatozoa physiology by maintaining the integrity of the inner genome and structure of spermatozoa. Zinc is also effective in protecting sperm from bacteria and chromosomal damage. Due to its strong antioxidant properties, a sufficient amount of zinc in semen plasma shows a protective effect. Zinc is mainly derived from the prostate, and plays a key role in sperm motility, carrying out protective and antioxidant activities. It is also considered an antimicrobial factor against grampositive and gram-negative bacteria. Deficiency of this element can lead to failure in spermatogenesis, atrophy the seminiferous tubules. thereforehypogonadism and a high incidence of changes in sperm morphology. Therefore, further research is needed to prove the effectiveness of the minerals Ca, P, and Zn in Moringa leaves to increase sperm motility and viabilityon PE goats.

5. Conclusion

This study has successfully proven the use of feed supplement UMMMB markedly increases libido, total motility, and sperm viability of PE goats. The PE goats were supplemented UMMMB required a shorter time to mount the teaser and ejaculate into the artificial vagina. The high Zn mineral content in Moringa leaves is strongly suspected to increase the libido of PE goats. Mineral Zn stimulates the formation of Leydig cells to produce the hormone testosterone, thereby increasing libido.Moringa leaf flour that constituent UMMMB contains the high minerals Ca, P, and Zn which are strongly suspected to increase total motility and sperm viability. Minerals Ca and P function to increase sperm motility and capacitation. The mineral Zn is essential for spermatogenesis, sperm motility, and protects sperm from bacteria and chromosomal damage. Thus, the spermatozoa will become more motile and can survive.

Therefore, Moringa leaves can be used as a constituent of UMMMB to increase the libido and sperm quality of PE goats. This research still needs to be continued to prove the minerals Ca, P, and Zn in Moringa leaves can increase libido, motility and sperm viability in PE goats using Ca, P, and Zn from extracted Moringa leaves.

5.1. Acknowledgment

The authors would like to thank the Ministry of Technology and Higher Education, Research, Indonesia for supporting this research through APPLIED RESEARCH with Research Contract No. 123.20/UN.8.2/PP/2019 dated March 15, 2019. The author also expresses his deepest gratitude to the Rukun Jaya Makmur Farmers Group, Banjarbaru City, South Kalimantan for providing pens and its facilities for the research.

References

[1] SALMAN A. D., ELSHARGI K. M., AL-HABSI R. S. and AL-SADAIRI T. New Development of Feed Blocks Technology in the Sultanate of Oman. Livestock Research Rural Development, 2017, 29(6): http://www.lrrd.org/lrrd29/6/alaa29109.html

[2] RAMOS J. P. F., DE SOUSA W. H., CAVALCANTE I. T. R., OLIVEIRA J. S., SANTOS E. M., FILHO E. C. P., FREITAS F. F. D. and LEITE R. M. Multinutritional Blocks as A Food Strategy to Optimize the Use of Concentrate for Lactating Goats. Acta Scientiarum, 2019, 41: 47441. https://doi.org/10.4025/actascianimsci.v41i1.47441

[3] MOHD NOR M. F., RUSLI N. D., MAT K., HASNITA C. H., and MIRA P. Milk Composition and Milk Quality of Saanen Crossbreed Goats Supplemented by Mineral Blocks. Tropical Animal Science Journal, 2020, 43(2): 169-175. https://doi.org/10.5398/tasj.2020.43.2.169

[4] ALVARADO-RAMÍREZ E. R., JOAQUÍN-CANCINO ESTRADA-DROUAILLET В.. MARTÍNEZ-GONZÁLEZJ. C., and HERNÁNDEZ-MELÉNDEZ J. Moringa oleifera Lam: An Alternative Fodder in Livestock Production in Mexico. Agroproductividad, 2018, 11(2): 106https://revista-

agroproductividad.org/index.php/agroproductividad/article/v iew/134

[5] SANTOS R. S., EMERENCIANO NETO J. V., BONFIM B. R. S., DIFANTE G. S., BEZERRA J. D. V., LISTA F. N., GURGEL A. L. C., and BEZERRA M. G. S. Growth and Biomass Production of Moringa Cultivated in Semiarid Region as Responses to Row Spacing and Cuts. Tropical Animal Science Journal, 2021, 44(2): 183-187. https://doi.org/10.5398/tasj.2021.44.2.183.

[6] KHALIFA W. H., IBRAHIM F. M., EL MAKAWY A. I., SHARAF H. A., KHALIL W. K. B., and MAGHRABY N. A. Safety and Fertility Enhancing Role of Moringa oleifera Leaves Aqueous Extract in New Zealand Rabbit Goats. International Journal of Pharmacy, 2016, 6(1): 156-168. https://www.pharmascholars.com/current-issue.html

[7] SYARIFUDDIN N. A., TOLENG A. L., RAHARDJA D. P., ISMARTOYO, and YUSUF M. Improving Libido and Sperm Quality of Bali Bulls By Supplementation of Moringa oleifera Leaves. Media Peternakan, 2017, 40(2): 88-93. https://doi.org/10.5398/medpet.2017.40.2.88

[8] WAFA W. M., EL-NAGAR H. A., GABR A. A., and REZK M. M. Impact of Dietary Moringa oleifera Leaves Supplementation on Semen Characteristics, Oxidative Stress, Physiological Response and Blood Parameters ff Heat Stressed Buffalo Bulls. Journal of Animal and Poultry Production, 2017, 8(9): 367-379. DOI: 10.21608/jappmu.2017.46008

[9] BOLLWEIN H., JANETT F., and KASKE M. Effects of Nutrition on Sexual Development of Bulls. Animal Reproduction, 2017, 14(3): http://dx.doi.org/10.21451/1984-3143-AR1004

[10] GOPALAKRISHNAN L., DORIYA K., and KUMAR D. S. Moringa oleifera: A Review on Nutritive Importance and its Medicinal Application. Food Science and Human Wellness, 2016, 5(2) 49-56.

https://doi.org/10.1016/j.fshw.2016.04.001

[11] ADEKANMI A. A., ADEKANMI S. A., and ADEKANMI O. S. Qualitative and Quantitative Phytochemical Constituents of Moringa Leaf. International Journal of Engineering and Information Systems, 4(5): 10-17. http://www.ijeais.org/ijeais/index.php/ijeais-4-5-2020/

[12] LENIS Y. Y., ELMETWALLY M. A., MALDONADO-ESTRADA J. G., and BAZER F. W. Physiological Importance of Polyamines. *Zygote*, 2017, 25(3): 244–255.

https://doi.org/10.1017/S0967199417000120

[13] SINGH A. K., RAJAK S. K., KUMAR P., KERKETTA S., and YOGI R. K. Nutrition and Bull Fertility: A review. *Journal of Entomology and Zoology Studies*, 2018, 6(6): 635-643.

https://www.entomoljournal.com/archives/?year=2018&vol=6&issue=6&ArticleId=4517

[14] TIBARY A., BOUKHLIQ R., and El ALLALI K. Ram and Buck Breeding Soundness Examination. Revue Marocaine des Sciences Agronomiques et Vétérinaires, 2018, 6(2): 241-255.

https://core.ac.uk/download/pdf/230580575.pdf

[15] DIE DEUTSCHE RÖNTGENGESELLSCHAFT. Testosterone ELISA EIA-1559. DRG International Springfield, 2020.

https://www.google.com/url?sa=t&rct=j&q=&esrc=s&sourc e=web&cd=&cad=rja&uact=8&ved=2ahUKEwiJze6b3pz2A hVsSGwGHRyWB8gQFnoECAYQAQ&url=https%3A%2F %2Fwww.drg-diagnostics.de%2Ffiles%2Feia-1559_ifu-testosterone_2020-04-

17 endeitesfr.pdf&usg=AOvVaw0UOAp6pVpB93GpjmBC s5lK

[16] TANGA B. M., QAMAR A. Y., RAZA S., BANG S., FANG X., YOON K., and CHO J. Semen Evaluation: Methodological Advancements in Sperm Quality-Specific Fertility Assessment - A Review. *Animal Bioscience*, 2021, 34(8): 1253-1270. DOI:10.5713/ab.21.0072.

[17] NATIONAL RESEARCH COUNCIL. Nutrient Requirement of Small Ruminant (Sheep, Goat, Cervids, and New World Camelids). The National Academies Press, Washington DC, 2007. https://www.nap.edu/resource/ruminants/errata.pdf.

[18] AX R. L., DALLY M. R., DIDION B. A., LENZ R. W., LOVE C. C., VARNER D. O., HAFEZ B. and BELLIN M. E. Semen Evaluation. In: HAFEZ B., & HAFEZ E. Z. E. (eds) *Reproduction in Farm Animals. Seventh edition*. Lippincott Williams & Wilkins, Philadelphia, 2000: 365-375. https://doi.org/10.1002/9781119265306.ch25.

[19] SARMA K., & DEVI J. Morphometrical Changes of the Seminiferous Tubules and Leydig Cells in Assam Goats (*Capra hircus*) from birth to 10 months. *Journal of Applied Animal Research*, 2017, 45(1): 268-274. https://doi.org/10.1080/09712119.2016.1174127

[20] GEMEDA A. E., & WORKALEMAHU K. Body weight and scrotal-testicular biometry in three indigenous breeds of bucks in arid and semiaridagroecologies, Ethiopia. *Journal of Veterinary Medicine*, 2017, 5276106. https://doi.org/10.1155/2017/5276106

[21] ZIRKIN B. R., & PAPADOPOULOS V. Leydig Cells: Formation, Function, and Regulation. *Biology of Reproduction*, 2018, 99(1): 101–111. https://doi.org/10.1093/biolre/ioy059

[22] DE SANTIAGO A., ALVARADO J., LÓPEZ A., TRUJILLO G., ÁLVAREZ M., and MELLADO M. Effects of Testosterone Administration and Feeding Level on Reproductive Activity in Sexually Inactive Goat Bucks. *Journal of the Hellenic Veterinary Medical Society*, 2018, 69(2): 991-998. http://dx.doi.org/10.12681/jhvms.18032.

[23] WIDIYONO I., SARMIN, PUTRO P. P., and ASTUTI P. Effects of Nutrition Status on Semen Characteristics of

Kacang Goats. *Pakistan Journal Nutrition*, 2017, 16: 678-683. https://scialert.net/abstract/?doi=pjn.2017.678.683

[24] ARMANSYAH, BARAT T. E. R. P., HANDINI C. V. R., ALIZA D., SUTRIANA A., HAMDAN H., PANJAITAN B., SAYUTI A., and SIREGAR T. N. Concentration and motility of spermatozoa and testosterone level of kacang goat after seminal vesicle extract administration. *Open Veterinary Journal*, 2018, 8(4): 406–410. http://dx.doi.org/10.4314/ovj.v8i4.9

[25] SHAH S. M. H., ALI S., ZUBAIR M., JAMIL H. and AHMAD N. Effect of Supplementation of Feed with Flaxseed (Linumusitatisimum) Oil on Libido and Semen Quality of Nilli-Ravi Buffalo Bulls. *Journal of Animal Science Technology*, 2016, 58(7): 251-256. https://doi.org/10.1186/s40781-016-0107-3

[26] HAHN K., FAILING K. and WEHREND A. Effect of Temperature and Time After Collection on Buck Sperm Quality. *BMC Veterinary Research*, 2019, 15: 355. https://doi.org/10.1186/s12917-019-2135-y

[27] KULAKSIZ R., ARI U. Ç., KURU M., YILDIZ S., LEHIMCIOĞLU N. C., and ÖZTÜRKLER Y. Seasonal Variations in Testicular Measurements, Fresh Sperm Quality and Post-Thaw Sperm Motility in Gurcu Goat Goats. *Slovak Journal of Animal Science*, 2020, 53(4): 161–167. https://sjas.ojs.sk/sjas/article/view/662

[28] HARCHEGANI A. B., IRANDOOST A., MIRNAMNIHA M., RAHMANI H., TAHMASBPOUR E., and SHAHRIARY A. Possible Mechanisms for the Effects of Calcium Deficiency on Male Infertility. *International Journal of Fertility and Sterility*, 2019, 12(4): 267-272. https://pubmed.ncbi.nlm.nih.gov/30291684/

[29] SKORACKA K., EDER P., ŁYKOWSKA-SZUBER L., DOBROWOLSKA A., and KRELA-KAŹMIERCZAK I. Diet and Nutritional Factors in Male (In)fertility-Underestimated Factors. *Journal of Clinical Medicine*, 2020, 9(5): 1400. https://doi.org/10.3390/jcm9051400

参考文:

[1] SALMAN A. D., ELSHARGI K. M., AL-HABSI R. S. 和 AL-SADAIRI T. 阿曼苏丹国饲料块技术的新发展。农村 发 展 畜 牧 业 研 究 , 2017, 29(6): 109. http://www.lrrd.org/lrrd29/6/alaa29109.html

[2] RAMOS J. P. F., DE SOUSA W. H., CAVALCANTE I. T. R., OLIVEIRA J. S., SANTOS E. M., FILHO E. C. P., FREITAS F. F. D. 和 LEITE R. M. 多营养块作为优化泌乳山羊使用浓缩物的食物策略。科学学报, 2019, 41: 47441. https://doi.org/10.4025/actascianimsci.v41i1.47441

[3] MOHD NOR M. F., RUSLI N. D., MAT K., HASNITA C. H., 和 MIRA P. 补充矿物质块的萨能杂交山羊的奶成分和奶质。热带动物科学杂志, 2020, 43(2): 169-175. https://doi.org/10.5398/tasj.2020.43.2.169

[4] ALVARADO-RAMÍREZ E. R., JOAQUÍN-CANCINO S., ESTRADA-DROUAILLET B., MARTÍNEZ-GONZÁLEZJ. C., 和 HERNÁNDEZ-MELÉNDEZ J. 辣木林:墨西哥畜牧生产中的替代饲料。农业生产力, 2018, 11(2): 106-110. https://revista-agroproductividad.org/index.php/agroproductividad/article/view/134

- [5] SANTOS R. S., EMERENCIANO NETO J. V., BONFIM B. R. S., DIFANTE G. S., BEZERRA J. D. V., LISTA F. N., GURGEL A. L. C., 和 BEZERRA M. G. S. 半干旱地区种植的辣木的生长和生物量生产作为对行距和切割的响应。热带动物科学杂志, 2021, 44(2): 183-187. https://doi.org/10.5398/tasj.2021.44.2.183.
- [6] KHALIFA W. H., IBRAHIM F. M., EL MAKAWY A. I., SHARAF H. A., KHALIL W. K. B., 和 MAGHRABY N. A. 辣木叶水提取物在新西兰兔山羊中的安全性和生育力增强作用。国际药学杂志,2016,6(1):156-168. https://www.pharmascholars.com/current-issue.html
- [7] SYARIFUDDIN N. A., TOLENG A. L., RAHARDJA D. P., ISMARTOYO, 和 YUSUF M. 通过补充辣木叶提高巴厘公牛的性欲和精子质量。媒体彼得纳坎, 2017, 40(2): 88-93. https://doi.org/10.5398/medpet.2017.40.2.88
- [8] WAFA W. M., EL-NAGAR H. A., GABR A. A., 和 REZK M. M. 膳食辣木叶补充剂对热应激水牛公牛精液特征、氧化应激、生理反应和血液参数的影响。畜禽生产杂 志 , 2017, 8(9): 367-379. DOI: 10.21608/jappmu.2017.46008
- [9] BOLLWEIN H., JANETT F., 和 KASKE M. 营养对公 牛性发育的影响。动物繁殖, 2017, 14(3): 707-613 http://dx.doi.org/10.21451/1984-3143-AR1004
- [10] GOPALAKRISHNAN L., DORIYA K., 和 KUMAR D. S. 辣木:营养重要性及其药用价值综述。食品科学与人类健康,2016,5(2):49-56. https://doi.org/10.1016/j.fshw.2016.04.001
- [11] ADEKANMI A. A., ADEKANMI S. A., 和 ADEKANMI O. S. 辣木叶的定性和定量植物化学成分。 国际工程与信息系统杂志, 4(5): 10-17. http://www.ijeais.org/ijeais/index.php/ijeais-4-5-2020/
- [12] LENIS Y. Y., ELMETWALLY M. A., MALDONADO-ESTRADA J. G., 和 BAZER F. W. 多胺的 生 理 重 要 性 。 合 子 , 2017, 25(3): 244–255. https://doi.org/10.1017/S0967199417000120
- [13] SINGH A. K., RAJAK S. K., KUMAR P., KERKETTA S., 和 YOGI R. K. 营养与公牛生育能力:综述。昆虫学和动物学研究杂志,2018,6(6):635-643. https://www.entomoljournal.com/archives/?year=2018&vol=6&issue=6&ArticleId=4517
- [14] TIBARY A., BOUKHLIQ R., 和 El ALLALI K. 公羊和巴克育种健全性检查。摩洛哥农学和兽医学评论, 2018, 6(2): 241-255. https://core.ac.uk/download/pdf/230580575.pdf
- [15] DIE DEUTSCHE RÖNTGENGESELLSCHAFT. 睾酮酶联免疫吸附测定酶免疫测定-1559。德国 X 射线协会国际 斯 普 林 菲 尔 德 , 2020. https://www.google.com/url?sa=t&rct=j&q=&esrc=s&source=web&cd=&cad=rja&uact=8&ved=2ahUKEwiJze6b3pz2AhVsSGwGHRyWB8gQFnoECAYQAQ&url=https%3A%2F

- $\frac{\%2Fwww.drg-diagnostics.de\%2Ffiles\%2Feia-1559\ ifu-testosterone_2020-04-$
- $\frac{17 \ ende ites fr.pdf \& usg = AOvVaw 0 UOAp6pVpB93GpjmBC}{s51K}$
- [16] TANGA B. M., QAMAR A. Y., RAZA S., BANG S., FANG X., YOON K., 和 CHO J. 精液评估:精子质量特定生育力评估的方法学进展 综述。动物生物科学, 2021, 34(8): 1253-1270. DOI:10.5713/ab.21.0072.
- [17] NATIONAL RESEARCH COUNCIL. 小型反刍动物(绵羊、山羊、鹿类和新大陆骆驼)的营养需求。国家科学院出版社,华盛顿特区,2007. https://www.nap.edu/resource/ruminants/errata.pdf.
- [18] AX R. L., DALLY M. R., DIDION B. A., LENZ R. W., LOVE C. C., VARNER D. O., HAFEZ B. 和 BELLIN M. E. 精液评估。在: HAFEZ B., 和 HAFEZ E. Z. E. (编辑) 农场动物的繁殖。第七版。利平科特·威廉姆斯和威尔金斯,费城 , 2000: 365-375. https://doi.org/10.1002/9781119265306.ch25.
- [19] SARMA K., 和 DEVI J. 阿萨姆山羊 (山羊属) 从出生到 10 个月的细精管和睾丸间质细胞的形态变化。应用动物 研究 杂志, 2017, 45(1): 268-274. https://doi.org/10.1080/09712119.2016.1174127
- [20] GEMEDA A. E., 和 WORKALEMAHU K. 埃塞俄比亚干旱和半干旱生态环境中三种本土雄鹿品种的体重和阴 囊 睾 丸 生 物 测 定 。兽 医 杂 志 , 2017, 5276106. https://doi.org/10.1155/2017/5276106
- [21] ZIRKIN B. R., 和 PAPADOPOULOS V. 莱迪格细胞:形成、功能和调节。生殖生物学, 2018, 99(1): 101–111. https://doi.org/10.1093/biolre/ioy059
- [22] DE SANTIAGO A., ALVARADO J., LÓPEZ A., TRUJILLO G., ÁLVAREZ M., 和 MELLADO M. 睾酮给药和喂养水平对性不活跃山羊雄鹿生殖活动的影响。希腊 兽 医 学 会 杂 志 , 2018, 69(2): 991-998. http://dx.doi.org/10.12681/jhvms.18032.
- [23] WIDIYONO I., SARMIN, PUTRO P. P., 和 ASTUTI P. 营养状况对卡仓山羊精液特性的影响。巴基斯坦杂志营养, 2017, 16: 678-683. https://scialert.net/abstract/?doi=pjn.2017.678.683
- [24] ARMANSYAH, BARAT T. E. R. P., HANDINI C. V. R., ALIZA D., SUTRIANA A., HAMDAN H., PANJAITAN B., SAYUTI A., 和 SIREGAR T. N. 精囊提取物给药后卡仓山羊的精子浓度和活力和睾酮水平。开放 兽 医 杂 志 , 2018, 8(4): 406–410. http://dx.doi.org/10.4314/ovj.v8i4.9
- [25] SHAH S. M. H., ALI S., ZUBAIR M., JAMIL H. 和AHMAD N. 饲料中添加亚麻籽 (亚麻) 油对尼利-拉维水牛公牛性欲和精液质量的影响。动物科学技术杂志, 2016, 58(7): 251-256. https://doi.org/10.1186/s40781-016-0107-3

- [26] HAHN K., FAILING K. 和 WEHREND A. 收集后温度和时间对巴克精子质量的影响。生物医学中心兽医研究, 2019, 15: 355. https://doi.org/10.1186/s12917-019-2135-v
- [27] KULAKSIZ R., ARI U. Ç., KURU M., YILDIZ S., LEHIMCIOĞLU N. C., 和 ÖZTÜRKLER Y. 古尔库山羊睾丸测量值、新鲜精子质量和解冻后精子活力的季节性变化。斯洛伐克动物科学杂志, 2020, 53(4): 161–167. https://sjas.ojs.sk/sjas/article/view/662
- [28] HARCHEĞANI A. B., IRANDOOST A., MIRNAMNIHA M., RAHMANI H., TAHMASBPOUR E., 和 SHAHRIARY A. 缺钙对男性不育影响的可能机制。国际 生 育 与 不 育 杂 志 , 2019, 12(4): 267-272. https://pubmed.ncbi.nlm.nih.gov/30291684/
- https://pubmed.ncbi.nlm.nih.gov/30291684/
 [29] SKORACKA K., EDER P., ŁYKOWSKA-SZUBER L.,
 DOBROWOLSKA A., 和 KRELA-KAŹMIERCZAK I. 男
 性(不)生育力中的饮食和营养因素 被低估的因素。临
 床 医 学 杂 志 , 2020, 9(5): 1400.
 https://doi.org/10.3390/jcm9051400